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The role of STAT1-cooperative DNA binding in myocardial Infarction

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The role of STAT1-cooperative DNA binding in myocardial Infarction

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This thesis is submitted for the degree of

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The role of STAT1-cooperative DNA binding in myocardial infarction



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AFFIDAVIT

Here I declare that my doctoral thesis entitled

“The role of STAT1-cooperative DNA binding in myocardial infarction”

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Table of Contents

| | |
|--|-----|
| Abbreviations | VI |
| Acknowledgements | X |
| Abstract | XII |
| 1. Introduction..... | 1 |
| 1.1 Discovery of the JAK/STAT signal transduction pathway..... | 2 |
| 1.1.1 Interferons are categorized as cytokines | 2 |
| 1.1.2 Structure, function, and regulation of STAT proteins | 3 |
| 1.1.3 STAT1 | 5 |
| 1.1.4 STAT3..... | 6 |
| 1.1.5 Type I and II interferon signalling | 6 |
| 1.1.6 STAT1-cooperative DNA binding | 8 |
| 1.1.7 Antagonistic activities of STAT1 and STAT3 | 9 |
| 1.2 The JAK/STAT signalling pathway in cardiovascular diseases | 10 |
| 1.2.1 Myocardial infarction and leukocytes responses..... | 11 |
| 1.2.2 Regulation of leucocyte transendothelial migration by the JAK/STAT pathway ... | 12 |
| 1.3 Original hypotheses and aims of this thesis | 13 |
| 2. Materials and Methods | 14 |
| 2.1 Materials..... | 14 |
| 2.1.1 Chemicals and reagents..... | 14 |
| 2.1.2 Recombinant proteins and enzymes..... | 17 |
| 2.1.3 Water | 17 |
| 2.1.4 Kits | 17 |
| 2.1.5 Culture media | 18 |
| 2.1.6 Cell lines and animals | 18 |
| 2.1.7 shRNA lentiviral transduction particles | 19 |
| 2.1.8 Primers | 19 |
| 2.1.9 Antibodies | 21 |
| 2.1.10 Equipments | 21 |
| 2.1.11 Disposables | 23 |

| | |
|--|----|
| 2.1.12 Software | 24 |
| 2.2 Methods..... | 26 |
| 2.2.1 Cell culture..... | 26 |
| 2.2.1.1 Thawing frozen cells | 26 |
| 2.2.1.2 Harvesting of cells for cryopreservation | 26 |
| 2.2.1.3 Maintenance of cultured cells | 26 |
| 2.2.1.4 Treatment of cultured cells with different stimuli | 27 |
| 2.2.1.5 Lentiviral transduction of HL-60 cells | 27 |
| 2.2.2 Protein Analysis..... | 27 |
| 2.2.2.1 Cell lysis and fractionation..... | 27 |
| 2.2.2.2 Protein extraction of HL-60 cells..... | 28 |
| 2.2.2.3 Western blotting analysis..... | 28 |
| 2.2.2.4 Electrophoretic mobility shift assay..... | 29 |
| 2.2.2.5 Radioactive labelling of EMSA probes | 29 |
| 2.2.3 DNA analysis..... | 30 |
| 2.2.3.1 DNA isolation from mouse tails | 30 |
| 2.2.3.2 Mice genotyping..... | 30 |
| 2.2.4 RNA analysis | 31 |
| 2.2.4.1 RNA extraction from cell culture and reverse transcription | 31 |
| 2.2.4.2 Real-time PCR..... | 31 |
| 2.2.5 Animal experiments | 32 |
| 2.2.5.1 Left anterior descending coronary artery ligation in mice | 32 |
| 2.2.5.2 Immunohistochemistry of paraffin-embedded sections..... | 32 |
| 2.2.6 RNA sequencing and analysis | 34 |
| 2.2.7 Parallel plate flow chamber assay | 35 |
| 2.2.8 Statistical analysis | 35 |
| 3. Results | 36 |
| 3.1 Echocardiographic analysis demonstrated beneficial outcomes in STAT1-F77A mice after myocardial infarction..... | 36 |
| 3.2 Expression of STAT1 and STAT3 in both cardiomyocytes and infiltrating immune cells after myocardial infarction | 41 |
| 3.3 Transcriptomic analysis of F77A mice in a LAD ligation model | 42 |

| | |
|---|-----|
| 3.4 Identification of differentially expressed genes in STAT1-F77A and wild-type mice at day 1 post-myocardial infarction | 44 |
| 3.5 Gene set enrichment analysis identified common and unique pathways associated with myocardial infarction | 53 |
| 3.6 Identification of immune-related pathways differentially regulated after myocardial infarction | 59 |
| 3.7 Identification of signal pathways and cellular processes differentially regulated after LAD ligation | 65 |
| 3.8 Gene set enrichment analysis revealed changes in metabolic signature following myocardial infarction..... | 77 |
| 3.9 Differentially regulated pathways in F77A as compared to wild-type mice in the infarcted area following LAD ligation | 84 |
| 3.10 A gene expression signature indicative of immune cells infiltration detected in F77A mice following LAD ligation | 86 |
| 3.11 Analysis of STAT1-target genes associated with myocardial infarction..... | 90 |
| 3.12 Co-stimulatory effects of interferon- γ , interferon- α , and interleukin-6 in MHEC-5 cells | 91 |
| 3.13 Generation of STAT1- and STAT3-knockdown HL-60 cell lines..... | 93 |
| 3.14 DMSO-induced differentiation results in upregulation of STAT1 and STAT3 in HL-60 cells | 94 |
| 3.15 Decreased phosphorylation levels of STAT1 and STAT3 in HL-60 knockdown cells | 94 |
| 3.16 Downregulation of ezrin in STAT1- and STAT3-knockdown HL-60 cells..... | 97 |
| 3.17 Reduced transendothelial migration in HL-60 cells with decreased STAT1 and STAT3 expression | 97 |
| 4. Discussion | 99 |
| 5. References..... | 108 |

Abbreviations

| | |
|----------|--|
| A | Adenine |
| APS | Ammonium persulfate |
| ATP | Adenosine triphosphate |
| BSA | Bovine serum albumin |
| C | Cytosine |
| CAMs | Cell adhesion molecules |
| CBP/p300 | CREB-binding protein |
| CCL | C-C motif chemokine ligand |
| CCR | C-C chemokine receptor |
| CD | Cluster of differentiation |
| cDNA | Complementary deoxyribonucleic acid |
| ChIP | Chromatin immunoprecipitation |
| CXC | C-X-C motif chemokine |
| CXCL | C-X-C motif chemokine ligand |
| CXCR | CXC chemokine receptor |
| DAB | 3,3'-Diaminobenzidine |
| DBD | DNA-binding domain |
| DMEM | Dulbecco's modified Eagle medium |
| DMSO | Dimethyl sulfoxide |
| DNA | Deoxyribonucleic acid |
| dNTP | Deoxyribonucleotide triphosphate |
| dsDNA | double stranded deoxyribonucleic acid |
| DTT | Dithiothreitol |
| EDTA | Ethylenediaminetetraacetic acid |
| EF | Ejection fraction |
| EGTA | Ethylene glycol-bis(β -aminoethyl ether)-N,N,N',N'-tetraacetic acid |
| EMSA | Electrophoretic mobility shift assay |
| ERM | Ezrin, radixin, moesin |
| FAS | Fractional area shortening |
| FBS | Fetal bovine serum |
| FS | Fractional shortening |

| | |
|-------------------|---|
| G | Guanine |
| GAF | Gamma-activating factor |
| GAGE | Generally applicable gene set enrichment |
| GAPDH | Glyceraldehyde-3-phosphate dehydrogenase |
| GBPB | Glucan-binding protein B |
| GSEA | Gene set enrichment analysis |
| HEPES | 4-(2-Hydroxyethyl)-1-piperazineethanesulfonic acid |
| HIF | Hypoxia-inducible factor |
| HPRT | Hypoxanthine-guanine phosphoribosyltransferase |
| HSP | Heat shock protein |
| HUVEC | Human umbilical vein endothelial cell |
| ICAM | Intercellular adhesion molecule |
| IFIT1 | Interferon-induced protein with tetratricopeptide repeats 1 |
| IFN | Interferon |
| IFNAR | Interferon- α/β receptor |
| IFNGR | Interferon- γ receptor |
| IgE | Immunoglobulin E |
| IgG | Immunoglobulin G |
| IL | Interleukin |
| IL-R | Interleukin receptor |
| IRDYE | Infrared fluorescent dyes |
| IRF | Interferon-regulatory factor |
| ISGF | Interferon-stimulated gene factor |
| ISRE | Interferon-stimulated response element |
| JAKs | Janus kinases |
| kDa | kilodalton |
| KEGG | Kyoto encyclopedia of genes and genomes |
| LAD | Left anterior descending coronary artery |
| LAVES | Landesamt für Verbraucherschutz und Lebensmittelsicherheit |
| Log | Logarithm |
| LV | Left ventricle |
| LVID _D | Left ventricular internal dimension-diastole |
| LVIDs | Left ventricular internal dimension-systole |
| MAPK | Mitogen-activated protein kinase |

| | |
|----------------|--|
| MCP-1 | Monocyte-chemoattractant protein-1 |
| MHEC5-T | Mouse heart endothelial cell clone 5-transformed |
| MI | Myocardial infarction |
| mRNA | Messenger RNA |
| N-terminus | Amino-terminus |
| NF- κ B | Nuclear factor κ -light-chain-enhancer of activated B cells |
| NNMT | Nicotinamide N-methyltransferase |
| padj | adjusted p-value |
| PAGE | Polyacrylamide gel electrophoresis |
| PBS | Phosphate-buffered saline |
| PCR | Polymerase chain reaction |
| PI3K | Phosphatidylinositol-4,5-biphosphate 3-kinase |
| PPAR | Peroxisome proliferator-activated receptor γ |
| PVDF | Polyvinylidene fluoride membrane |
| qPCR | Quantitative polymerase chain reaction |
| Rac1 | RAS-related C3 botulinum substrate 1 |
| RNA | Ribonucleic acid |
| RPKM | Reads per kilobase of transcript per million mapped reads |
| RPMI | Roswell Park Memorial Institute |
| rRNA | Ribosomal RNA |
| SDS | Sodium dodecyl sulfate |
| SELP | Selectin P |
| shRNA | short hairpin RNA |
| SPSS | Statistical package for the social sciences |
| STAT | Signal transducer and activator of transcription |
| T | Thymine |
| TAD | Trans-activating domain |
| TBE | Tris-borate-EDTA |
| TBS | Tris-buffered saline |
| TBS-T | Tris-buffered saline-Tween 20 |
| TCA | Tricarboxylic acid |
| TEMED | Tetramethylethylenediamine |
| TGF- β | Transforming growth factor- β |
| TNF | Tumor necrosis factor |

| | |
|------------------|-----------------|
| Vol _d | Volume diastole |
| Vol _s | Volume systole |
| WT | Wild-type |

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Abstract

Myocardial infarction (MI), defined as heart damage due to prolonged ischemia, accounts for significant mortality and morbidity and has a substantial financial burden worldwide. While major advancements have been achieved in the characterization of the JAK/STAT signalling pathway in the context of stress and infection, numerous questions on its involvement in cardiac dysfunction and remodeling remain unanswered. In this study, ligation of the left anterior descending coronary artery (LAD) was performed in wild-type and knock-in mice expressing a STAT1 point mutant with defective cooperative DNA binding (STAT1-F77A), to address the role of interferon signalling in acute myocardial infarction. We hypothesized that mice with dysfunctional STAT1 signalling would show a similar phenotype to mice expressing hyperactive STAT3 with respect to its cardioprotective actions. Using echocardiographic evaluation and transcriptomic analysis, we found that a loss of STAT1 tetramerisation protected against adverse cardiac remodeling in female mice. RNA sequencing uncovered numerous immune and metabolic pathways that were differentially regulated in these mice at day 1 post-myocardial injury. The top five immune-related pathways that were upregulated post-MI in both wild-type and F77A mice were cytokine-cytokine receptor interactions, chemokine signalling pathways, cell adhesion molecules, regulation of actin cytoskeleton and leukocyte transendothelial migration, whereas the top five metabolism-related pathways downregulated post-MI were oxidative phosphorylation, citrate cycle, fatty acid metabolism, fatty acid degradation, and propanoate metabolism in both wild-type and STAT1-F77A mice. Gene set enrichment analysis revealed a downregulation of cardiac muscle contraction pathway in STAT1-F77A mice only, conceivably due to a heterogeneous cell population of infiltrating immune cells in the infarcted myocardium. In addition, the distinctive transcriptomic profile of infiltrating immune cells in the infarcted area of STAT1-F77A mice was characterized by the upregulation of several immune-activating markers to a higher order of magnitude as compared to their wild-type littermates, including chemokines and their receptors, adhesion molecules, cytokines and their receptors, growth factors and their receptors, coagulation cascade proteins, and heat shock proteins. Notably, the top ten genes differentially expressed in the infarcted area of STAT1-F77A were *Fgf23*, *Cxcl3*, *Slfn4*, *Rab44*, *Cxcl2*, *Cd177*, *Gm5483*, *Il1r2*, *Cd300lf*, and *Slfn1*. The observed upregulation of genes encoding neutrophil markers and adhesion molecules and the significant downregulation of genes engaged in oxidative phosphorylation in the STAT1-F77A mouse line as compared to wild-type mice may act as an adaptive response to reduce oxidative stress, repair cardiac damage and increase

survival. Additionally, in a parallel plate flow chamber assay, STAT1- and STAT3-knockdown HL-60 cells had reduced transendothelial migration when compared to control cells. We conclude that in the early phase of myocardial infarction, a series of transcriptional regulations can probably initiate a beneficial remodeling of the left ventricle in a mouse model expressing dysfunctional STAT1. This study has expanded our understanding of STAT1 transcriptional regulation within the context of murine myocardial infarction.

1. Introduction

Ischemic heart diseases have accounted globally for more than one million deaths in 2017, making them the first cause of early death. Myocardial infarction (MI) results from acute obstruction of a coronary artery with subsequent myocardial cell death, leading to insufficient pumping of blood to meet the body's demand or what is known as heart failure (Thygesen *et al.*, 2018). Accumulative research in mechanistic and cellular adaptations to cardiac injury has enhanced our understanding of maladaptive cardiac remodeling (Schirone *et al.*, 2017). It is largely accepted that ischemic damage to the cardiac tissue is accompanied by sterile inflammatory responses, where several cytokines play significant roles (Epelman *et al.*, 2015). Until recently, the potential involvement of the JAK/STAT pathway in cardiac repair, regeneration and metabolism has remained largely underestimated. Hence, elucidation of the JAK/STAT signalling pathway in ischemia-induced cardiac dysfunction is relevant for a deeper understanding of the disease outcome. Notably, STAT proteins are known to control a remarkable variety of fundamental biological processes, such as cellular proliferation, differentiation, growth and homeostasis (Abroun *et al.*, 2015). As transcription factors, STAT proteins are characterized by their highly conserved amino acid sequence among different species, including nematodes, insects, and vertebrates (Wang and Levy, 2012). STAT1 and STAT2 were the first two members of STAT protein family to be discovered, followed by five more, namely STAT3, STAT4, STAT5A, STAT5B, and STAT6. Apart from STAT2, all human STAT proteins form homodimers (Kisseleva *et al.*, 2002). Additionally, STATs can be found in heterodimers, e.g. STAT1:STAT2 and STAT1:STAT3, where the magnitude and nature of heterodimeric STATs are influenced by the type of activating ligands (Delgoffe and Vignali, 2013). Since the immunological actions of interferons are not restricted to anti-viral immunity, the investigation of the JAK/STAT pathway is continuously extending our knowledge and generates surprising findings.

STAT1 knock-out mice have reported profound biological defects in response to type I, II, and III interferons and less to other ligands (Akira, 1999). Non-canonical STAT signalling exerts transcriptional regulation in mammals (Majoros *et al.*, 2017), where STAT1 and STAT3 can regulate gene transcription either through binding to DNA or indirectly by several co-activators. Additionally, unphosphorylated STAT3 is known to translocate to the mitochondria and interfere with the energy production by changes in the mitochondrial cellular respiration (Wegrzyn *et al.*, 2009). This extends further connections that link STATs and cellular metabolism, which has newly entered the picture (Dodington *et al.*, 2018).

1.1 Discovery of the JAK/STAT signal transduction pathway

The history of the JAK/STAT signal transduction discovery dates back to the 1980s. At that time, research groups led by James Darnell, Ian Kerr, and George Stark were interested in the question, how gene transcription could be induced within minutes following type I interferons treatment of cells. The rate at which a signal originated at the plasma membrane and transduced to activate target genes in the nucleus, proposed the presence of few intermediate steps. Utilizing different biochemical and genetic techniques, the Darnell, Kerr and Stark laboratories uncovered a previously unrecognized direct signal transduction pathway to the nucleus by identifying the STATs as signal transducers and activators of transcription (Darnell *et al.*, 1994; Fu *et al.*, 1992). Briefly following this pioneering breakthrough, non-receptor protein tyrosine kinases (pTK) were separately discovered in signalling pathways involving interferons, erythropoietin, and growth hormone receptors (Argetsinger *et al.*, 1993; Velazquez *et al.*, 1992; Witthuhn, *et al.* 1993). Identical kinases had also come into sight earlier in screening for new pTKs and were called Janus kinases (Firmbach-Kraft *et al.*, 1990; Wilks *et al.*, 1991). Genetic complementation of interferon-unresponsive mutants with recombinant JAK and STAT proteins had made it feasible to reconstitute interferon signalling between the cell membrane and the transcription machinery in the nucleus, with merely two elements: firstly, receptor-associated JAKs that carry the recruitment side for tyrosine phosphorylation of STATs as a first step, and secondly, tyrosine-phosphorylated STATs, which translocate to the nucleus and activate target genes through binding to specific DNA elements (Levy and Darnell, 1990). Currently, the JAK/STAT signal transduction is viewed as a key pathway transducing signals to the nucleus from class I and class II cytokine receptors. Owing to its immense implication in biology, the JAK/STAT pathway is portrayed as a major signalling paradigm (Bousoik *et al.*, 2018). Thus, it is not surprising that JAKs and STATs regulate vital processes in the cell, wherein almost every intricate biological process, from embryogenesis to aging is affected by the JAK/STAT signal transduction (Binari and Perrimon 1994).

1.1.1 Interferons are categorized as cytokines

Cytokines are extracellular soluble proteins that convey cell-to-cell signals. They control a plethora of intercellular processes and regulate the recruitment of cells that are involved in inflammation, cellular growth, proliferation, differentiation, apoptosis, angiogenesis, homeostasis, tissue repair, and remodeling. Different families of cytokines bind their cognate receptors on targeted cells, which ultimately determine cellular fate from activation to

differentiation and survival (Dinarello, 2007). When classified, they can be grouped by their structure and their receptors. One group of cytokines is called interferons (IFNs), which are produced as glycoproteins by a variety of cell types to carry important cell signalling events and trigger various immune responses (Pestka *et al.*, 2007). Notably, interferons can interfere with viral replication, making it less permissive for the virus to replicate inside the cell (Isaacs and Lindenmann, 1957). Classification of interferons is based upon their receptors, to which they bind to in order to carry the signal inside the cell. For example, interferons type I binds to IFNAR (interferon α/β receptor) which consists of two chains, IFNAR1 (α chain) and IFNAR2 (β chain), whereas interferon type II binds to INFGR (interferon- γ receptor). There are many type I interferons, namely IFN- α , IFN- β , IFN- κ , IFN- δ , IFN- ϵ , IFN- τ , IFN- ω , and IFN- ζ , and only one type II interferon, which is IFN- γ (Mesev *et al.* 2019).

1.1.2 Structure, function, and regulation of STAT proteins

The STATs are differentially regulated by a multitude of signalling molecules, including interferons, interleukins, growth factors, and hormones (Ihle *et al.*, 1996). The mammalian STAT family comprises seven STAT proteins, which are STAT1, STAT2, STAT3, STAT4, STAT5a, STAT5b, and STAT6 (Kisseleva *et al.*, 2002). STAT proteins have a size range between 750-900 amino acids. Studying the genomic clustering of STATs and searching for sequence similarity in other eukaryotic species have led to the identification of homologs most closely related to STAT3 and STAT5, in organisms like *Dictyostelium* (Kawata *et al.*, 1997), *C. elegans* (Wang and Levy, 2006), and *Drosophila* (Zeidler *et al.*, 2000). In contrast to *Drosophila*, where a single STAT transmits the signal through the classical JAK/STAT pathway, the homologs in *C. elegans* and *Dictyostelium* signal through different pathways.



Figure 1: A schematic representation of STAT proteins structure. NH₂, N-terminus domain; CCD, coiled-coil domain; DBD, DNA-binding domain; LD, linker domain; SH2, Src-homology 2 domain; TS: phosphotyrosyl tail with phosphorylation sites; TAD, transactivation domain.

Utilizing multiple sequence alignment models and analyzing the crystal structure have helped in the identification of six structurally and functionally conserved domains in STAT proteins (Figure 1) (Becker *et al.*, 1998; Chen *et al.*, 1998): (1) The amino terminal domain (NH₂) being approximately 125 amino acids long, is a well conserved domain that promotes

homotypic interactions between STAT molecules (Vinkemeier *et al.*, 1998). (2) The coiled-coil domain, located between 135-315 amino acids, is a four-helix bundle that protrudes laterally ($\sim 80\text{\AA}$) from the core and associates with a number of potentially important regulatory proteins. Hence, it is implicated in nuclear import and export (Begitt *et al.*, 2000). (3) The DNA-binding domain (DBD) (320-480 amino acids), recognizes the palindromic GAS element in activated STAT homodimers (except STAT2), and participates in the process of nuclear import/export (Horvath *et al.*, 1995). (4) The linker domain, localized roughly between 480-575 amino acids, is able to structurally translate the dimerization signal to the DNA-binding motif and regulate basal nuclear export in resting cells (Hüntelmann *et al.*, 2014). (5) The SH2 domain, which is between 575-680 amino acids, is the most highly conserved motif. This domain mediates the specific recruitment to the appropriate receptor as well as the formation of active STAT dimers (Greenlund *et al.* 1994; Heim *et al.*, 1995). (6) The transcriptional activation domain (TAD) is conserved between mouse and man for every STAT member except STAT2, however, it diverges substantially between STATs. The TAD domain includes conserved serine phosphorylation sites that direct the recruitment of coactivators and in some cases regulate STAT stability (Varinou *et al.*, 2003).

Following phosphorylation of the tyrosine activation motif on the conserved tyrosine residue (701), the phosphotyrosyl residue is subsequently recognized by an SH2 domain on other STAT protomers leading to the formation of active STAT dimers. The amino terminal domain and the SH2 domain are the most conserved structures, they are also involved in protein-protein interactions, including dimerization, binding to the transcriptional co-activator CREB-binding protein (CBP/p300), and binding to the cytoplasmic receptor (Wojciak *et al.*, 2009; Zhang *et al.*, 1996). The cytoplasmic and nuclear compartment both play an important role in modulating the kinetics of STAT activation. While the activation of the JAK/STAT signal pathway leads to the formation of dimers in parallel conformation by reciprocal phosphotyrosine (pY)-SH2 domain interactions, the antiparallel alignment is present mostly in the absence of stimulation. The parallel conformation positions the two phosphorylated tyrosine residues in opposing SH2-domain pockets, thus protecting STAT1 from dephosphorylation. Therefore, the current notion is that conformational transition from the parallel to the antiparallel alignment renders the phosphotyrosine residue susceptible to enzymatic dephosphorylation and deactivation (Mertens *et al.*, 2006; Wenta *et al.*, 2008; Zhong *et al.*, 2005). Nonetheless, the exact details of these conformational shifts are still a matter of debate.

Once exported to the cytosol, STATs can enter new cycles of re-activation, nuclear import, inactivation and nuclear export. Apart from canonical tyrosine-phosphorylation, STAT-driven transcription is also regulated by a number of posttranslational modifications, including serine phosphorylation and sumoylation (Droescher *et al.*, 2011; Zhu *et al.*, 1997). Another critical aspect of the JAK-STAT signalling pathway is its short-lived nature, generally confined to less than a few hours. This decline in the cascade is achieved through the activation of opposing regulatory proteins, namely phosphatases and suppressors of cytokine signalling (SOCS) (Adams *et al.*, 1998), along with less well studied regulators such as protein inhibitor of activated STATs (PIAS) (Shuai, 2006), N-myc and STAT interactor (NMI) (Bao and Zervos, 1996), and PDZ and LIM domain-containing protein (SLIM) (Tanaka *et al.*, 2005). Other phosphatases that regulate the JAK-STAT signalling involve: protein tyrosine phosphatase non-receptor type 1 (PTP1B), protein tyrosine phosphatase non-receptor type 2 (TC-PTP), protein tyrosine phosphatase non-receptor type 6 (SHP-1), protein tyrosine phosphatase non-receptor type 11 (SHP-2), and protein tyrosine phosphatase non-receptor Type 13 (PTP-BL). Some of these regulators appear to target receptors and their associated JAKs, while others target the STATs directly and their nuclear export (Aman *et al.*, 1997; Böhmer and Friedrich, 2014).

1.1.3 STAT1

STAT1 is a crucial activator of numerous immune effector functions and regulates the expression of genes that are involved in chemokine and cytokine signalling, antigen-presentation, phagocytosis, anti-microbial, and anti-viral immunity. Therefore, expectedly humans carrying variants of STAT1 in the form of nonsense or missense mutations present clinically with a higher predisposition to bacterial and viral infections (Boisson-Dupuis *et al.*, 2012; Casanova *et al.*, 2012). The identification of STAT1 was based on screening of human complementary DNA (cDNA) libraries, that were generated from HeLa cells treated with IFN- γ and IFN- α for 45 min and 16 hours, respectively. In particular, molecular cloning and sequence analysis of the cDNA eventually led to the characterization of the two forms p91 and p84, termed as STAT1 α and STAT1 β , which are the products of alternative splicing of the same gene. Following nuclear import, STAT1 dimers that are not bound to DNA have a conformational change resulting in the exposure of a critical phosphotyrosine residue at position 701 (Staab *et al.*, 2013). This conformational shift renders STAT1 susceptible to the removal of the phosphate group by T-cell protein tyrosine phosphatase (TC-45), which makes the export of STAT1 feasible (ten Hoeve *et al.*, 2002). A compromised dissociation reaction

of STAT1 from DNA not only hampers the constant search for GAS (gamma-activated sequence) sites, but also impedes with the rate of nucleocytoplasmic shuttling and the transcriptional activity of STAT1 (Yang *et al.*, 2002). Notably, unphosphorylated STAT1 also functions as a transcription factor. However, it is quickly substituted by activated STAT1 dimers following IFN- γ stimulation (Yao *et al.*, 2017). Hence, unphosphorylated STAT1 stays for several days in the cell, leading to enhanced signal transduction following re-exposure to low doses of interferon (Cheon and Stark, 2009). Additionally, experimental data suggested the existence of unphosphorylated homodimers, STAT1:STAT2 and STAT1:STAT3 heterodimers before cytokine stimulus (Ho *et al.*, 2016).

1.1.4 STAT3

In response to inflammation, binding of acute-phase response factor (APRF) to IL-6-responsive elements in the DNA of target genes is known to activate the transcription of acute-phase proteins (APPs) (Gabay *et al.*, 1999; Wegenka *et al.*, 1993). Analysis of APRF revealed a mouse cDNA encoding the 770 amino acid-long homodimers of STAT3. In addition, Northern blot detected a single mRNA transcript (4.8 kb) in both mouse and human, corresponding to a protein product of 88 kDa. Stimulation of mouse liver cells for 3 hours with IL-6 has shown a 5-fold induction of STAT3 mRNA levels (Akira *et al.*, 1994; Zhong *et al.*, 1994). Expression of STAT3 transcripts has been found to be ubiquitous in all tissues examined. Previous research demonstrated that STAT3 can transduce signals from the IL-6 and IL-10 families, as well as granulocyte-colony stimulating factor (G-CSF), leptin, IL-21, IL-27, and potentially IFN-Is (Akira *et al.*, 1997). Moreover, *in vitro* studies have recently shown that different growth factors and oncogenes are inducing the activation of STAT3, which is associated with the expression of anti-apoptotic/pro-survival genes (Yeh *et al.*, 2009). Deletion of the *Stat3* gene in mice resulted in an early lethal embryonic phenotype at embryonic day 6.5–7.5 (Takeda *et al.*, 1997), while tissue specific *Stat3* deletions demonstrated an inflammatory phenotype with decreased transformative capacity in cancer cells and tumor regression in animal models (Yu *et al.*, 2009). Moreover, overexpression of the active *Stat3* allele, using dominant negative inhibitors and RNA silencing, was associated with increased transformation capacity (Bromberg *et al.*, 1998; Gough *et al.*, 2009; Vultur *et al.*, 2005).

1.1.5 Type I and II interferon signalling

The JAK-STAT1 pathway is activated by binding of type I and II interferons to their cognate receptors on the plasma membrane, followed by the activation of non-covalently attached

Janus-activated kinases (JAKs) bound to these receptors (Darnell *et al.*, 1994; Levy and Darnell, 2002). The mammalian family of JAKs consists of four ubiquitously expressed members: JAK1, JAK2, JAK3, and TYK2, but the expression of JAK3 is primarily limited to the hematopoietic cells. In type I interferon signalling, binding of several ligands induces the dimerization of type I interferon receptor subunits and the phosphorylation of their associated JAKs on specific tyrosine residues. This is followed by the phosphorylation of other tyrosine-residues on the cytoplasmic receptor domains, creating docking sites for latent cytoplasmic STAT molecules, that bind through their Src-homology-2 (SH2) domain. Then, the STATs become phosphorylated by the JAKs on a conserved tyrosine residue near their C-terminus. Upon phosphorylation, the dissociation of STAT molecules from the receptor complex ensues, followed by dimerization via reciprocal phosphotyrosine (pY)-SH2 domain interactions. A complex which comprises STAT1, STAT2, and IRF9, called interferon-stimulated gene factor 3 (ISGF3), is subsequently formed. The ISGF3 complex binds DNA on interferon-sensitive response element (ISRE) sequences (5'-AGTTTCN₂TTTC-3'), to induce gene transcription of target genes. However, in type II interferon signalling, interferon- γ receptor subunits (IFNGR1 and IFNGR2) associate with JAK1 and JAK2, which phosphorylate exclusively STAT1. Two STAT1 monomers form a dimeric complex called gamma-activation factor (GAF), that after translocation to the nucleus and binds GAS sites (5'-TTCN₃GAA-3') to activate gene transcription of IFN- γ -driven target genes. Thus, type I and II interferons translate extracellular signals and mediate differential transcriptional responses (Figure 2) (Lee and Ashkar, 2018). In addition, STATs nuclear translocation is regulated through two pathways, which are the importin α -5 dependent pathway and the Ran (RAs-related Nuclear protein) nuclear import pathway (McBride and Reich, 2003).

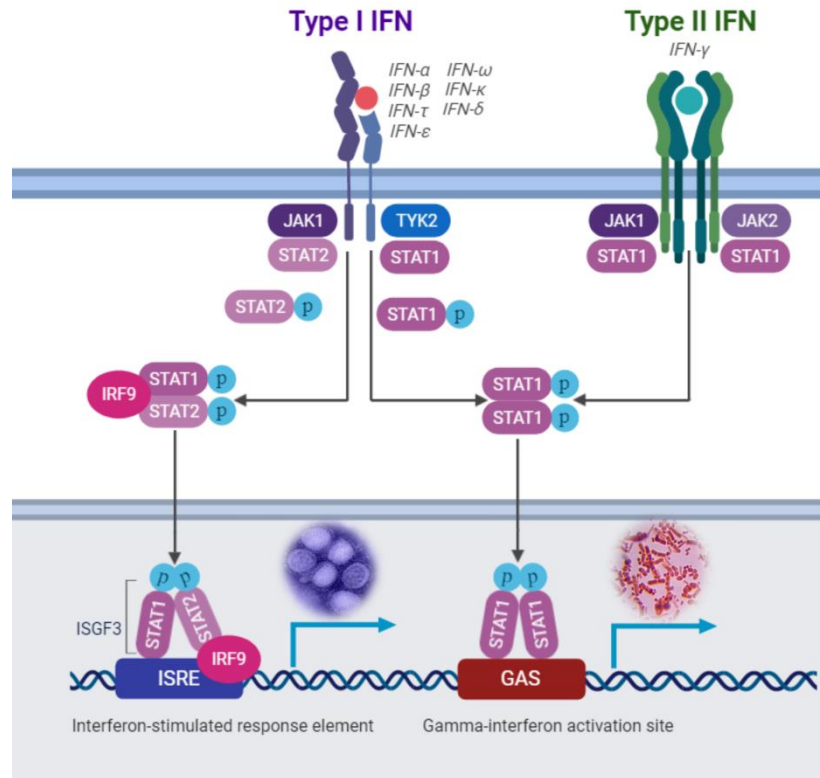


Figure 2: Differential regulation of type I and II interferons signalling pathways. *IFN-α* (type I IFN) leads to the formation of STAT1-STAT2 heterodimers, which also require IRF9 (interferon-regulatory factor 9). This heterotrimeric complex known as ISGF3 (interferon-stimulated gene factor 3) drives transcription at so-called ISREs (IFN-stimulated response elements) sites. *IFN-γ* (type II IFN) results in the formation of STAT1-STAT1 homodimers that are known as GAF, which binds to other DNA sequence, termed GAS sites, to drive transcription.

1.1.6 STAT1-cooperative DNA binding

It has been shown that phage lambda will enter a lytic or a lysogenic cycle by performing a swift switch utilizing cooperative DNA binding (Murray and Gann, 2007). Similarly, STAT dimers polymerize on tandem GAS and ISRE sites through N-terminal domain mediated interactions (Xu *et al.*, 1996). Begitt and colleagues studied the role of cooperative DNA binding in type I and type II interferon signalling. The foundation of their work was the generation of a mutant mouse line (STAT1-F77A), which is defective in polymerizing additional STAT dimers on the DNA, as a result of a single amino acid substitution, namely alanine for phenylalanine at position 77 in the N-terminus domain. Infection experiments using this knock-in mouse line emphasized the physiological significance of STAT1 tetramerisation in immunity. Although STAT1-F77A mice expressed similar levels of STAT1 and responded similarly to vesicular stomatitis virus (VSV) infection, as compared to wild-type littermates, they had a severe defect in antibacterial immunity, exhibiting a higher susceptibility to infection with *Listeria monocytogenes*. Additionally, STAT1-F77A-

fibroblasts have shown an impaired induction of IFN- γ -regulated genes as compared to IFN- α -regulated genes. Furthermore, using chromatin immunoprecipitation and electrophoretic mobility shift assays, STAT1-F77A recruitment to GAS sites was impeded upon IFN- γ stimulation as compared to wild-type cells, but not upon stimulation with IFN- α (Begitt *et al.*, 2014).

1.1.7 Antagonistic activities of STAT1 and STAT3

Phosphorylation of STAT proteins by JAKs is essential for facilitating STAT homo- and hetero-dimerization in a parallel dimer conformation. Upon binding of STATs to the receptors, phosphorylation of tyrosine residue 701 in STAT1 and tyrosine residue 705 in STAT3 occurs, followed by STATs dimerization and nuclear translocation to activate responsive genes. Accordingly, tyrosine-phosphorylated STAT1 can form dimers with STAT3 and with STAT2, and the balance between STAT1:STAT3 heterodimers and their STAT3 homodimers will define the transcriptional activity in a cell. Different group of cytokines have been shown to activate distinct STAT proteins. This differential potency of activation for certain cytokines is best exemplified by IFN- γ , a potent activator of STAT1, and STAT3 mediating signalling from interleukin-6 (IL-6) family members, in particular IL-6, leukemia inhibitory factor (LIF) and cardiotrophin-1 (CT-1). Remarkably, STAT1 and STAT3 can substitute for each other absence following IFN- γ and IL-6 stimulation, this is characterized by GAS-dependent gene activation in STAT1-null cells, and enhanced activation of STAT1 in STAT3-null cells resulting in IFN- γ -like response (Costa-Pereira *et al.*, 2002). Every STAT protein has a modular structure comprised of distinct domains. STAT1 and STAT3 display certain homology within the coiled-coiled domain, the DNA-binding domain, the linker domain, the Src-homology (SH2) domain and the carboxy-terminal transactivation domain (TD). To the contrary, the amino-terminal domain is dramatically less conserved between STAT1 and STAT3, signifying this region as a mediator of cellular responses that are unique to distinctive STAT proteins. Even though STAT1 and STAT3 share very comparable conserved structure, both have potential antagonistic effects of their gene products, which regulate processes such as cellular proliferation and apoptosis. For example, several studies have established that STAT1 plays an important role as a tumor suppressor promoting apoptosis in multiple cell types, however, STAT3 exhibits anti-apoptotic activities. Additionally, while STAT3 promotes proliferation and transformation in a wide spectrum of carcinomas, STAT1 is reported to have anti-proliferative activities (Table 1).

Table 1: Antagonistic activities of STAT1 and STAT3

| STAT1 | STAT3 |
|--------------------|---------------------------------|
| anti-proliferative | pro-proliferative |
| pro-apoptotic | pro-survival |
| pro-inflammatory | immunosuppressive |
| anti-angiogenic | pro-angiogenic & pro-metastatic |

1.2 The JAK/STAT signalling pathway in cardiovascular diseases

Several animal models have been developed to investigate the role of various signalling pathways in cardiovascular function under normal or pathologically conditions, including the JAK/STAT signalling pathway, which is a key regulator of several cardiovascular pathologies. Studies have been performed on its function in atherosclerosis, hypertension, myocardial infarction, hypertrophy, myocarditis, and ischemia-reperfusion-induced cardiac injury (El-Adawi *et al.*, 2003; Mascareno *et al.*, 2001; Ortiz-Muñoz *et al.*, 2009; Satou and Gonzalez-Villalobos, 2012; Zhang *et al.*, 2013). In the myocardium, STATs regulate the expression of inflammation- and extracellular matrix-related genes as well as genes regulating apoptosis, angiogenesis and proliferation (Figure 3).

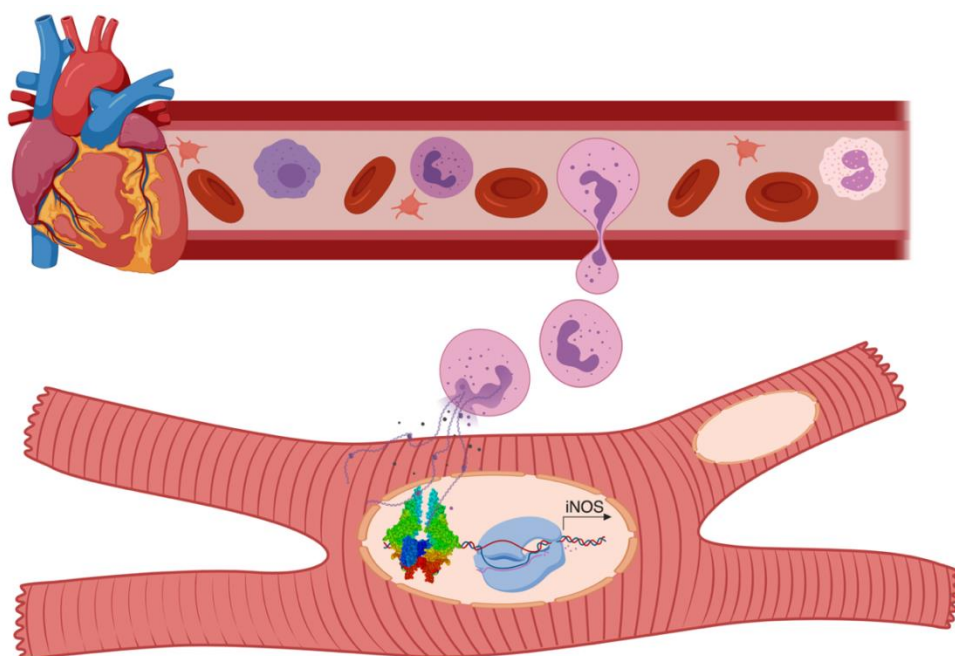


Figure 3: STAT1 and STAT3 regulate transcriptional processes in the ischemic heart. *The balance between the activation state of both members of the STAT protein family in the cardiomyocytes as well as in the immune infiltrating cells mediates the remodeling process after myocardial infarction.*

1.2.1 Myocardial infarction and leukocytes responses

Myocardial infarction (MI) triggers a cascade of events which recruit different types of immune cells such as neutrophils, macrophages, lymphoid cells, and mast cells to orchestrate several inflammatory responses. Following a cardiac insult, leukocytes move out of the circulation towards the site of injury, guided by a gradient of chemoattractant peptides, known as chemokines (Altara *et al.*, 2016; Frangogiannis, 2014; Gomez *et al.*, 2018; Meng *et al.*, 2016; Nahrendorf and Swirski, 2013; Pinto *et al.*, 2012). Recruited monocytes and neutrophils are initially mobilized from their original niche in the bone marrow to the spleen, generating myeloid cells that take part in multiple repair processes (Lambert *et al.*, 2008; Nahrendorf *et al.*, 2007). Heart-infiltrating immune cells perform a complex role clearing debris and stabilizing the heart wall, through a tuned balance between residents and recruited/differentiated hematopoietic progenitors (Heidt *et al.*, 2014; Massa *et al.*, 2005; Nahrendorf and Swirski, 2016). Hence, trafficking of immune cells shapes the outcome following myocardial infarction by profoundly influencing cardiac repair, fibrosis, regeneration and scar formation, and exerting either pro-inflammatory or anti-inflammatory actions (Forte *et al.*, 2018; Ruparelia *et al.*, 2015). Activated chemokines signal through G-protein-coupled receptors, which are expressed on various immune cells. Dissociation of the α - and β - γ -subunits of G-proteins leads to downstream signalling cascades which ultimately result in changes in cell polarity and motility through small GTPases (Zweemer *et al.*, 2014). For example, highly coordinated migration and velocity of neutrophils towards sites of injury are established and maintained by cell adhesion molecules (CAMs). Notably, CAMs underpin a crucial cross-talk between innate and adaptive immune cells. They can either anchor the cell to the substratum or transduce signals between adjacent cells to reshape their migratory responses and dynamically remodel the organization of their actin cytoskeleton.

The migration of neutrophils into extravascular tissue can be viewed as a series of interactions which is mediated by (1) the integrin family, (2) the immunoglobulin superfamily, (3) selectins, and (4) cadherins. CAMs can also be classified according to the role they play to (1) recognize antigens, (2) adhere to each other and to the extracellular matrix, and (3) carry co-stimulation signal (Cavallaro and Dejana, 2011). Additionally, activated chemokine receptors stimulate a rapid elevation of diacylglycerol and cytosolic calcium levels to induce effector functions and coordinate microvascular remodeling, including nitric oxide production and release of reactive oxygen species (ROS) (Cavalera and Frangogiannis, 2014; Saporov *et al.*, 2017). The ensuing removal of dead cells sustains a local environment that supports cardiomyocyte repair. Platelet activation is another hallmark of acute myocardial infarction,

which can be stimulated by collagen, von Willebrand factor (VWF), thromboxane A2 (TXA2), adenosine diphosphate (ADP), and thrombin. The proceeding thrombus formation within the ventricle increases the myocardium stiffness (Dutta *et al.*, 2012). Furthermore, myocardial ischemia reprograms catabolic and anabolic pathways in the heart to adjust for new requirements of energy acquisition and substrate utilization, and mediate repair processes, cell survival, and growth (Meyer and Voigt, 2017; Wende *et al.*, 2017). Nearly a century ago, Otto Warburg found that cancer cells favor the metabolization of glucose via aerobic glycolysis. Similarly, deregulated metabolism with increased glycolysis has emerged as a significant hallmark of ischemic injury in the heart (Chen *et al.*, 2018). The goal in near future would be to enhance our understanding of the metabolism mediated through the JAK/STAT signalling and propose a rational basis to reprogram metabolic pathways for an improved cardiac repair and regeneration (Doenst *et al.*, 2013).

1.2.2 Regulation of leucocyte transendothelial migration by the JAK/STAT pathway

Cell migration is an intricate, synchronized process in which numerous parts of the cell are involved, including surface receptors, intracellular signalling proteins, and the cytoskeleton. Cumulating evidence has highlighted the role of inflammatory cytokines and transcription factors as crucial mediators of cell migration and polarization (Dustin and Chan, 2000; Nieto *et al.*, 1997; Randolph, 2001). One of the best studied examples in cellular polarity is the epithelial-to-mesenchymal transition, in which cells lose epithelial polarity and attachment to adjacent cells (Lamouille *et al.*, 2014). The inverse happens when migrating cells arrive at their target location, build an epithelium and/or integrate into a previous epithelial tissue (Muller, 2015). Examples of such transitions taking place are demonstrated by neural crest development in vertebrates (Bronner and LeDouarin, 2012). Similarly, heart regeneration depends on overlooked temporal and spatial roles for macrophages and neutrophils, where they mediate extracellular matrix regulation. Consequently, improper migration and polarization can potentially affect repair processes and thus contribute to the development of heart failure. STAT1 was found to be essential for IL-6 expression and the increased adhesion and migration of monocytes across the blood-brain barrier, using different *in-vitro* models in human immunodeficiency virus 1 (HIV-1) infection (Yang *et al.*, 2009). Another study has shown that depletion of STAT1 in a fibroblast cell line resulted in a reduction of cell migration (Xie *et al.*, 2001). Whereas, IFN- γ treatment arrested monocyte chemotaxis by modulating the organization of the cytoskeleton via RAC/CDC42 pathways (Hu *et al.*, 2008). Recent studies investigating the loss of STAT3 expression had revealed an elevation of Rac1

activity in murine embryonic fibroblasts, which mediated a random type of migration through decreasing directional persistence and the assembly of actin stress fibers (Marcella *et al.*, 2005; Teng *et al.*, 2009). These findings suggest a major role of STAT1 and STAT3 in chemokine-induced leucocyte transendothelial migration.

1.3 Original hypotheses and aims of this thesis

There is accumulating evidence defining a critical role for cytokines in post-myocardial infarction remodeling. While there is ample evidence indicating a pivotal cardioprotective role of STAT3 in acute and chronic stress in the heart (Zouein *et al.*, 2015), only a small proportion of the studies carried out have investigated the functional role of STAT1. It is proposed that STAT1 deficiency in the heart has a cardioprotective role by enhancing autophagy (McCormick *et al.*, 2012). Accordingly in this thesis, the pleiotropy of STAT1 action and the nature of its transcriptional program in the failing myocardium was studied, with its subsequent impact on cardiac function using genome-wide profiling of the cardiac transcriptome in infarcted areas of wild-type and STAT1 tetramerisation-deficient mice (STAT1-F77A).

This project investigates the biological roles of STAT1-cooperative DNA binding in a murine model of myocardial infarction. In order to test our hypothesis, the following aims were established to:

- 1) understand the role of STAT1 as a transcription factor and the function of its downstream effector genes responding to myocardial infarction, as well as their potential cross talk in normal and dysfunctional signaling,
- 2) investigate the potential role of the antiviral program “type-II interferon response” in pathophysiological remodeling, by identifying signalling pathways that are differentially regulated between wild-type and transgenic STAT1-F77A animals following injury of the myocardium, and
- 3) perform proof-of-concept experiments to study the functional role of STAT1 and STAT3 in mediating transendothelial migration in a neutrophil-like cell line, by generating STAT1- and STAT3-knockdown HL-60 cells.

2. Materials and Methods

2.1 Materials

2.1.1 Chemicals and reagents

Table 2: List of chemicals and reagents

| Name | Company |
|---|--|
| 3,3'-Diaminobenzidine (DAB) | Sigma-Aldrich, Taufkirchen, Germany |
| 4-(2-Hydroxyethyl)-1-piperazineethanesulphonic acid (HEPES, Pufferan $\geq 99.5\%$, p. a.) | Carl Roth, Karlsruhe, Germany |
| Agarose | Carl Roth |
| Ammonium persulphate (APS) | Carl Roth |
| Ampicillin | Sigma-Aldrich |
| Bovine serum albumin (BSA) Fraction V | Carl Roth |
| Bromophenol blue | Fisher Bioreagents, Pittsburgh, USA |
| Chloroform | Sigma-Aldrich |
| Complete mini proteinase inhibitor cocktail | Roche, Rotkreuz, Switzerland |
| Deoxy-adenosine triphosphate (dATP) [^{33}P]-labelled | Hartmann Analytic, Braunschweig, Germany |
| Deoxynucleotide triphosphates (dNTP) | Carl Roth |
| Dimethyl sulphoxide (DMSO) | Applichem, Darmstadt, Germany |
| Dithiothreitol (DTT) | Applichem |
| Entellan | Merck Millipore, Darmstadt, Germany |
| Eosin Y | Merck Millipore |
| Ethanol (99.8%) | Carl Roth |
| Ethidium bromide solution (0.625 g/ml) | VWR Life Science, Erlangen, Germany |
| Ethylene glycol-bis(β -aminoethyl ether)-N,N,N',N'-tetraacetic acid (EGTA) | Carl Roth |
| Ethylenediamine-tetraacetic acid (EDTA) | Sigma-Aldrich |
| Fetal bovine serum (FBS) | Biochrom, Berlin, Germany |
| Fluoromount G | Southern Biotech, Birmingham, USA |
| Formaldehyde solution (37%) | Carl Roth |
| L-Glutamine | Thermo Fisher Scientific, Langenselbold, |

| | |
|---|---------------------------------------|
| | Germany |
| Glycerol, ROTIPURAN $\geq 99.5\%$, p. a. | Carl Roth |
| Glycine | Carl Roth |
| HEPES, cell culture grade | Carl Roth |
| Hydrogen peroxide solution (30%) | Carl Roth |
| IGEPAL CA-630 | Sigma-Aldrich |
| Isopropanol | Carl Roth |
| Kanamycin | Sigma-Aldrich |
| Lithium chloride | Carl Roth |
| Magnesium chloride | Sigma-Aldrich |
| Mayer's haemalum | Merck Millipore |
| Mercaptoethanol | Sigma-Aldrich |
| Methanol | Merck Millipore |
| Methylene blue with (3%) acetic acid | Stemcell, Cologne, Germany |
| Pefabloc | Roche |
| Penicillin/streptomycin | Biochrom |
| Phenol/chloroform/isoamylalcohol (25:24:1) | Carl Roth |
| Phosphate-buffered saline (PBS) | Life Technologies, Darmstadt, Germany |
| Polyacrylamide gel electrophoresis running buffer (SDS-PAGE) | Carl Roth |
| Poly-deoxyinosinic-deoxycytidylic acid (Poly-dIdC) | Sigma-Aldrich |
| Ponceau-S | Sigma-Aldrich |
| Potassium chloride | Merck Millipore |
| Puromycin | Sigma-Aldrich |
| Radioimmunoprecipitation assay buffer (RIPA) | Merck Millipore |
| Rotiphorese gel 30 (acrylamid/bisacrylamid (37.5:1)-solution) | Carl Roth |
| Rotiphorese gel 40 (acrylamid/bisacrylamid (29:1)-solution) | Carl Roth |
| Roti-plast paraffin | Carl Roth |
| Sodium acetate | Carl Roth |
| Sodium azide | Thermo Fisher Scientific |
| Sodium bicarbonate | Carl Roth |

| | |
|------------------------------------|-------------------------------|
| Sodium chloride | Carl Roth |
| Sodium chloride solution (0.9%) | B. Braun, Melsungen, Germany |
| Sodium citrate | Carl Roth |
| Sodium deoxycholate | Applichem |
| Sodium dodecyl sulphate (SDS) | Carl Roth |
| Sodium orthovanadate | Acros Organics, Geel, Belgium |
| Sodium pyruvate | Thermo Fisher Scientific |
| Sulfuric acid | Carl Roth |
| Tetramethylethylenediamine (TEMED) | Carl Roth |
| Tris-base | Carl Roth |
| Tris-HCl | Carl Roth |
| Triton X-100 | Carl Roth |
| TRIzol reagent | Thermo Fisher Scientific |
| Tween-20 | Carl Roth |
| Xylene | Merck Millipore |

Table 3: List of drugs

| Name | Company |
|-------------------------------------|--|
| Cepetor (medetomidine, 1 mg/ml) | Cp-pharma, Burgdorf, Germany |
| Fentanyl (0.05 mg/ml) | Rotexmedica, Trittau, Germany |
| Flumazenil(0.5mg/ml) | Inresa Arzneimittel, Freiburg, Germany |
| Isoflurane | Abbvie, Wiesbaden, Germany |
| Ketamine | Medistar, Ascheberg, Germany |
| Midazolam (5 mg/ml) | Rotexmedica |
| Metamizol (500 mg/ml) | Zentiva Pharma, Frankfurt am Main, Germany |
| Atipamezole hydrochloride (5 mg/ml) | Cp-pharma |
| Buprenorphine hydrochloride | Reckitt Benckiser, Slough, UK |
| Xylazine hydrochloride | Ecuphar, Greifswald, Germany |

2.1.2 Recombinant proteins and enzymes

Table 4: List of recombinant proteins and enzymes

| Name | Company |
|--|--------------------------------|
| IFN- α , human, recombinant from <i>E. coli</i> | Biomol, Hamburg, Germany |
| IFN- γ , human, recombinant from <i>E. coli</i> | Biomol |
| IL-6, human recombinant | Biomol |
| InnuTaq DNA polymerase | Analytic Jena |
| InnuTaq HOT-A DNA polymerase | Analytic Jena |
| Proteinase K | Carl Roth |
| Taq polymerase | Ares Bioscience, Köln, Germany |
| Trypsin | Sigma-Aldrich |
| <i>Tsp509I</i> | Thermo Fisher Scientific |

2.1.3 Water

Table 5: List of water

| | |
|--|---|
| Double-distilled H ₂ O | PURELAB Plus purification system, Elga Labwater, Celle, Germany |
| Molecular biology grade water | 5 Prime, Heidelberg, Germany |
| DNase-, RNase- and proteinase-free water | |

2.1.4 Kits

Table 6: List of kits

| Name | Company |
|--|---|
| ABsolute Blue QPCR SYBR Green Mix | Thermo Fisher Scientific |
| dsDNA 905 Reagent Kit | Advanced Analytical Technologies, Heidelberg, Germany |
| PeqGold total RNA Kit | VWR Life Science |
| QuantiFluor dsDNA System | Promega |
| Thermo Scientific Verso cDNA Synthesis Kit | Thermo Fisher Scientific |
| TruSeq stranded mRNA prep Kit | Illumina, California, USA |

2.1.5 Culture media

Table 7: List of media

| Name | Company |
|---|--------------------------------|
| Dulbecco's modified Eagle's medium (DMEM) (Glucose 4.5 g/l, 580 mg/l L-glutamine, 110 mg/l Na-pyruvate) | Biochrom |
| DMEM (Glucose 4.5 g/l, without L-glutamine, 110 mg/l Na-pyruvate) | Biochrom |
| RPMI (Roswell Park Memorial Institute medium) 1640 | Lonza, Cologne, Germany |
| Endothelial cell growth medium | PromoCell, Heidelberg, Germany |

2.1.6 Cell lines and animals

Table 8: List of cell lines

| Cell line | Characteristics |
|-----------|--|
| HL-60 | Human suspension cell line derived from peripheral blood acute promyelocytic leukemia, exhibit phagocytic activity and responsiveness to chemotactic stimuli |
| HUVEC | Human umbilical vein endothelial cells are cells derived from the endothelium of veins from the umbilical cord |

Table 9: List of animals

| Animal model | Description |
|---------------------|---|
| STAT1-F77A knock-in | Mouse line carrying a point mutation resulting in a substitution of alanine in position 77 for phenylalanine on a C57BL6/N background (Charles Rivers, Sulzfeld, Germany) (Begitt <i>et al.</i> , 2014) |

Mice were kept in individually ventilated cages in the University Medical Centre Animal Facility, Göttingen. Experiments performed were approved by the Lower Saxony State Office for Consumer Protection and Food Safety (Niedersächsisches Landesamt für Verbraucherschutz und Lebensmittelsicherheit, LAVES).

2.1.7 shRNA lentiviral transduction particles

Table 10: List of STAT MISSION shRNA lentiviral transduction particles

| | |
|--|---|
| Signal transducer and activator of transcription 1, 91kDa | |
| SHCLNV-NM_007315 | |
| TRCN0000280021 | CCGGCTGGAAGATTTACAAGATGAACTCGAGTTCATCTTGTAATCTTCCAGTTTTTG |
| TRCN0000280024 | CCGGCCCTGAAGTATCTGTATCCAACCTCGAGTTGGATACAGATACTTCAGGGTTTTTG |
| TRCN0000004267 | CCGGCTGGAAGATTTACAAGATGAACTCGAGTTCATCTTGTAATCTTCCAGTTTTT |
| TRCN0000004265 | CCGGCCCTGAAGTATCTGTATCCAACCTCGAGTTGGATACAGATACTTCAGGGTTTTT |
| TRCN0000004266 | CCGGCGACAGTATGATGAACACAGTCTCGAGACTGTGTTTCATCATACTGTCGTTTTT |
| Signal transducer and activator of transcription 3 (acute-phase response factor) | |
| SHCLNV-NM_003150 | |
| TRCN0000329888 | CCGGGGCGTCCAGTTCCTACTAACTCGAGTTTAGTAGTGAAGTGGACGCCTTTTTTG |
| TRCN0000329886 | CCGGGCAAAGAATCACATGCCACTTCTCGAGAAGTGGCATGTGATTCTTTGCTTTTTG |
| TRCN0000329887 | CCGGGCACAATCTACGAAGAATCAACTCGAGTTGATTCTTCGTAGATTGTGCTTTTTG |
| TRCN0000020840 | CCGGGCTGACCAACAATCCCAAGAACTCGAGTTCTTGGGATTGTTGGTCAGCTTTTT |
| TRCN0000020843 | CCGGGCAAAGAATCACATGCCACTTCTCGAGAAGTGGCATGTGATTCTTTGCTTTTT |

2.1.8 Primers

Table 11: List of oligonucleotide primers for genotyping

| Gene | Sequence |
|--------------------|----------------------------------|
| <i>Stat1</i> -F77A | F: 5'-CGCGAATTGCTAATAAAACAA G-3' |
| | R: 5'-TGCTGCTGAGTCCAAATAAAG-3' |
| <i>Stat1</i> -WT | F: 5'-GGCTGACCTTGGAACCTTGTG-3' |
| | R: 5'-AAGAGTCAGCAGGGGTCTGA-3' |

Table 12: List of oligonucleotide primers for quantitative PCR (qPCR)

| Gene | Sequence |
|---------------|---|
| <i>Bcl2l1</i> | F: 5'-GCCTTTTCTCCTTTGGCGG-3' R: 5'-TCCACAAAAGTGTCACAGCC-3' |
| <i>Cd34</i> | F: 5'-AGGCTGATGCTGGTGCTAG-3' R: 5'-AGTCTTTCGGGAATAGCTCTG-3' |
| <i>Cox7a</i> | F: 5'-AAAACCGTGTGGCAGAGAAG-3' R: 5'-CAGCGTCATGGTCAGTCTGT-3' |
| <i>Csf3r</i> | F: 5'-TGAACCTACACCCAGGCCTTC-3' R: 5'-GTGCATGAGGCAGGATAGGT-3' |
| <i>Cxcl2</i> | F: 5'-GCAAGGCTAACTGACCTGGA-3' R: 5'-AGGCACATCAGGTACGATCC-3' |
| <i>Cxcl9</i> | F: 5'-GGAGTTCGAGGAACCCTAGT-3' R: 5'-AGGCAGGTTTGATCTCCGTT-3' |
| <i>Gbp2b</i> | F: 5'-AAGGGCATCTGGATGTGGTG-3' R: 5'-TCTGGTTGTCACCTTCTGC-3' |
| <i>Hprt</i> | F: 5'-AGTCCCAGCGTCGTGATTAG-3' R: 5'-TGATGGCCTCCCATCTCCTT-3' |
| <i>Irf1</i> | F: 5'-ACCCTGGCTAGAGATGCAGA-3' R: 5'-TGCTTTGTATCGGCCTGTGT-3' |
| <i>Kras</i> | F: 5'-CAAGAGCGCCTTGACGATACA-3' R: 5'-CCAAGAGACAGGTTTCTCCATC-3' |
| <i>Nnmt</i> | F: 5'-GAAGGCAACAGAATGAAGGGAC-3' R: 5'-TTCCTGAGGGCAGTGCGATA-3' |
| <i>Rb1</i> | F: 5'-ACTCCGTTTTTCATGCAGAGACTAA-3' R: 5'-GAGGAATGTGAGGTATTGGTGACA-3' |
| <i>Sca1</i> | F: 5'-AGGAGGCAGCAGTTATTGTGG-3' R: 5'-CGTTGACCTTAGTACCCAGGA-3' |
| <i>Stat1</i> | F: 5'-CCGTTTTTCATGACCTCCTGT-3' R: 5'-TGAATATTCCCCGACTGAGC-3' |
| <i>Thy1</i> | F: 5'-TCCAGAATCCAAGTCGGAAC-3' R: 5'-GTTATTCTCATGGCGGCAGT-3' |

Primers were purchased from Sigma with a grade of purification as desalted. NCBI-BLAST tool was used for primer design.

2.1.9 Antibodies

Table 13: List of primary antibodies

| Name | Company | Cat-No. |
|------------------------|---|---------|
| Ezrin | Cell Signaling Technology, Frankfurt am Main, Germany | 3145 |
| Moesin (C-15) | Santa Cruz Biotechnology, Heidelberg, Germany | sc-6410 |
| STAT1(D1K9Y) | Cell Signaling Technology | 14994 |
| pSTAT1 (Tyr701)(58D6) | Cell Signaling Technology | 9167 |
| STAT3 (D1B2J) | Cell Signaling Technology | 30835 |
| STAT3 (H-190) | Santa Cruz Biotechnology | sc-7179 |
| pSTAT3 (Tyr705) (D3A7) | Cell Signaling Technology | 9145 |
| GAPDH (14C10) | Cell Signaling Technology | 2118 |
| β-actin (8H10D10) | Cell Signaling Technology | 3700 |

Table 14: List of secondary antibodies

| Name | Company | Cat-No. |
|--|--|-----------|
| Donkey anti-rabbit secondary antibody IRDye 800CW IgG (H + L) | LI-COR Biosciences | 926-32213 |
| Donkey anti-mouse secondary antibody IRDye 800CW IgG (H + L) | LI-COR Biosciences | 926-32212 |
| Donkey anti-Goat secondary antibody IRDye 680RD IgG (H + L) | LI-COR Biosciences | 925-68074 |
| Biotinylated anti-rabbit IgG (H+L) derived from goat | Vector Laboratories, Peterborough, United Kingdom | BA-1000 |

2.1.10 Equipments

Table 15: List of equipment

| Name | Company |
|------------------------------------|---|
| 5075 ELV autoclave | Tuttnauer Europe B.V., Breda, Netherlands |
| Bandelin Sonopuls | Bandelin electronic, Berlin, Germany |
| Biological safety cabinet class II | Thermo Fisher Scientific |
| BioPhotometer plus | Eppendorf, Hamburg, Germany |

| | |
|--|---|
| Centrifuge 5804 R | Eppendorf |
| Digital video camera C10600 ORCA-R2 | Hamamatsu, Hamamatsu, Japan |
| Fragment analyzer | Advanced Analytical Technologies |
| Freezing container | Nalgene, Rochester, USA |
| Illumina HiSeq 4000 | Illumina |
| Incubator B 5061 EC-CO2 | Heraeus, Hanau, Germany |
| Microscope incubator | Solent Scientific |
| Microcentrifuge 5415 R | Eppendorf |
| Microscope, BX51 | Olympus, Tokyo, Japan |
| Microscope, fluorescent, Axiovert 200M | Zeiss, Oberkochen, Germany |
| Microscope, inverted, Axiovert 40 CFL | Zeiss |
| Microtome SM 2000 R | Leica Microsystems, Wetzlar, Germany |
| Mini vent type 845 | FMI GmbH, Seeheim-Jungenheim, Germany |
| Multifuge 1S-R | Heraeus |
| Neubauer-modified counting chamber | Paul Marienfeld, Königshofen, Germany |
| Odyssey CLx imaging system | LI-COR Biosciences, Bad Homburg vor der Höhe, Germany |
| Olympus IX81 time-lapse inverted fluorescence microscope | Olympus |
| Orbital shaker | VWR Life Science |
| Pipettes, type research (P1000, P200, P100, P10, P2.5) | Eppendorf |
| Power supply unit | Biometra, Göttingen, Germany |
| QuantiFluor dsDNA system | Promega, Mannheim, Germany |
| Real-time cycler mastercycler ep gradient | Eppendorf |
| Rocker duomax 1030 | Heidolph, Schwabach, Germany |
| SDS-PAGE gel running system | Biometra |
| Semi-dry blotting cell | Peqlab Biotechnologie, Erlangen, Germany |
| Sonicator UW 2070 | Bandelin electronic |
| Speedmill tissue homogenizer | Analytic Jena, Jena, Germany |
| Surgical forceps | Fine Science Tools, Heidelberg, Germany |
| Surgical scissors | Fine Science Tools |

| | |
|---|------------------------------------|
| Thermo shaker TS1 | Biometra |
| Tissue dehydration system TP1028 | Leica Microsystems |
| UVsolo TS imaging system | Analytic Jena |
| Vevo 2100 System | Visualsonics, Toronto, Canada |
| Waterbath GFL | Schuett-biotec, Göttingen, Germany |
| XCell II blot module | Thermo Fisher Scientific |
| XCell SureLock mini-cell electrophoresis system | Thermo Fisher Scientific |

2.1.11 Disposables

Table 16: List of disposables

| Name | Company |
|--|--------------------------------------|
| Cell culture flasks, T75 | Sarstedt, Nümbrecht, Germany |
| Cell culture plates, 6-well, 24-well, 96-well | Sarstedt |
| Cell scraper 25 cm | Sarstedt |
| Cryovials | Nunc, Roskilde, Denmark |
| Ethilon 9-0 BV-4 5.0 mm 3/8c EH7448G surgical silk | Ethicon, Norderstedt, Germany |
| Falcon tubes, 15 ml, 50 ml | Sarstedt |
| Filter (0.22 µm) | Carl Roth |
| Glass cover slip | Thermo Fisher Scientific |
| Glass microscope slide | Thermo Fisher Scientific |
| Injection needles | B. Braun, Melsungen, Germany |
| Insulin syringe, 29 G | B. Braun |
| Microreaction tube, 0.2 ml, 1.5 ml, 2 ml | Eppendorf |
| Minisart NY 25 syringe filter | Sartorius, Göttingen, Germany |
| Nitrocellulose membrane | GE Healthcare, Braunschweig, Germany |
| PCR-plate (96 well) | Nunc |
| Petri dishes, 10 cm | Corning, New York, USA |
| Pipette tips (P1000, P100, P10) | Sarstedt |
| Prolene 6-0 C-1 13 mm 3/8c 8889H surgical silk | Ethicon |

| | |
|--|---------------------------------------|
| Polyvinylidene fluoride membrane Immobilon-P (PVDF) | Merck Millipore |
| Serological pipettes, 5 ml, 10 ml | Sarstedt |
| Syringe, 20 ml, 10 ml | Becton Dickinson, Heidelberg, Germany |
| Tissue culture dish 20 cm | Nunc |
| Tissue culture plates, 96 well, 24 well, 6 well | Nunc |

2.1.12 Software

Table 17: List of software

| Name | Description | Distributor |
|-------------------------------------|---|--|
| BaseCaller | Base calling for sequence files | Illumina |
| bcl2fastq (version 2.17.1.14) | Formats conversion software | Illumina |
| biomaRt (version 2.32.1) | Gene annotation to database mining | Bioconductor R package |
| BLAST | Basic local alignment search tool | National Human Genome Research Institute, Bethesda, USA |
| CorelDraw | Vector graphics editor | Corel, Ottawa, Canada |
| DESeq2 package (version 1.14.1.) | Differential gene expression analysis based on the negative binomial distribution | Bioconductor R package |
| FastQC (version 0.11.7) | Quality control check on sequences | Babraham Institute, Cambridge, UK |
| featureCounts (version 1.4.5-p1) | Assigning sequence reads to genomic features | Bioconductor R package |
| FlowJo | Flow cytometry data analysis | FlowJo LLC, Ashland, USA |
| GraphPad Prism | Analysis and graphing solution | GraphPad, San Diego, USA |
| IBM SPSS | Statistical software, Version 24 | IBM, Kassel, Germany |
| ImageJ | Image processing software | NIH, Bethesda, USA |
| ImageStudio Lite | Image processing software | LI-COR Biosciences |
| Inkscape 2 | Vector graphics editor | Inkscape |
| Realplex ep | qPCR running and data analysis software | Eppendorf |

| | | |
|----------------------------------|--|--|
| RStudio | Open-source integrated development environment for statistical computing | RStudio Inc. Boston, USA |
| SigmaPlot | Scientific data analysis and graphing | Systat Software, Erkrath, Germany |
| Snapgene | DNA cloning and PCR tools | GSL Biotech, Chicago, USA |
| STAR aligner (version 2.5.2a) | NGS read aligner | National Human Genome Research Institute |
| Vevo 2100 | Ultrasound analysis software for image data | FUJIFILM, Tokyo, Japan |
| Volocity Software | 3D analysis of fluorescence images | PerkinElmer, Baesweiler, USA |
| Windows Office | Spreadsheet for calculation and graphing tools | Microsoft, Washington, USA |

2.2 Methods

2.2.1 Cell culture

2.2.1.1 Thawing frozen cells

To unfreeze cells, a cryovial containing 2×10^6 cells was thawed by gentle agitation in a water bath set at 37°C . When most of the ice started to melt, the vial was removed from the water bath and added immediately to 10 ml of pre-warmed culture medium. The cell suspension was centrifuged at $65 \times g$ for 6 min at room temperature. Supernatant was carefully aspirated, leaving the pellet undisturbed to be resuspended in 10 ml of pre-warmed culture medium and cultured in a T75 flask. MHEC5-T cells were cultured in DMEM, supplemented with 10% FBS and 1% penicillin/streptomycin. HL-60 cells were cultured in RPMI 1640, supplemented with 10% FBS and 1% penicillin/streptomycin, in the additional presence of L-glutamine and $1 \mu\text{g/ml}$ puromycin.

2.2.1.2 Harvesting of cells for cryopreservation

Cells were harvested by trypsinization after washing with PBS for adherent cells, or by centrifugation for suspension cells. Adherent cells were incubated with trypsin for 2 min, and then centrifuged at $65 \times g$ for 5 min. Cryopreservation medium was prepared by adding DMSO to FBS at a final concentration of 10% (v/v), which was placed on ice until ready to use. The cell pellet was resuspended in 1 ml of cold freezing medium at the recommended viable cell density (1.5×10^6 – 10×10^6). The cryovials were stored at -80°C .

2.2.1.3 Maintenance of cultured cells

Cells were maintained at 37°C in 5% CO_2 under a humidified atmosphere and passaged every 2 to 3 days. MHEC5-T cells were washed with PBS and subsequently trypsinized for 2 min at 37°C . Trypsinization was stopped by adding 5 ml of complete culture medium. An aliquot from this suspension was used to seed a new T75 flask with 1:10 dilution. HL-60 cells were counted and then subcultured at a density of 0.5×10^6 cells per ml. Cells were grown at densities no greater than 2.5×10^6 per ml. For protein and RNA analysis, MHEC5-T cells were seeded in a 6-well plate at a density of approximately 1.5×10^5 cells per cm^2 , and HL-60 cells were seeded in a 6-well plate at a density of 2×10^6 cells per ml. Absence of mycoplasma infection was routinely checked in the cell lines.

2.2.1.4 Treatment of cultured cells with different stimuli

MHEC5-T were treated with the respective stimulus (e.g. IFN- γ , IL-6) after removing the medium and replacing it with cytokine-containing medium at the indicated concentration. For stimulating HL-60 cells, without removing the media, the cytokine was added from an intermediate stock media to achieve the indicated concentration. For RNA extraction, cells were exposed 16 h prior to stimulation to a medium which had 0.5% FBS (low serum-culture medium). Stimuli concentrations were the following: human IFN- γ 50 ng/ml, murine IFN γ 5 ng/ml, and murine IFN- α 5 ng/ml, human IL-6 10 ng/ml, and murine IL-6 10 ng/ml.

2.2.1.5 Lentiviral transduction of HL-60 cells

Expression of STAT1 and STAT3 genes were knockdown in HL-60 cells using MISSION TRC shRNA lentiviral transduction particles (SIGMA) with pLKO.1 plasmid, using standard puromycin selection method. At the first day, 100 μ l of HL-60 cells were plated at a density of 1×10^6 cells per ml in a complete medium in 96-well plates prior to transduction. The lentiviral stock was slowly thawed on ice and then added to each well with a multiplicity of infection (MOI) of 5. At day 4, 100 μ l of puromycin was added to the transfected cells with a concentration of 1 μ g/ml. Cells were observed under the microscope for viability and passaged by adding fresh puromycin-supplemented media according to the cell density. Cells were transferred to a 6-well plate and subsequently cultured in a T25 and then a T75 flask, as they continued to proliferate.

2.2.2 Protein Analysis

2.2.2.1 Cell lysis and fractionation

Cells grown on a 6-well plate were washed with PBS, then lysed on ice for 5 min with 50 μ l of complete cytosolic protein lysis buffer (20 mM HEPES, 10 mM KCl, 1 mM EDTA, 100 μ M Na₃VO₄, 10% glycerol, 0.1% IGEPAL CA-630, supplemented with 1% Complete proteinase inhibitor cocktail, 3 mM DTT, 0.4 mM Pefabloc, pH 7.4). Adherent cells were harvested using a cell scraper and then transferred to a 1.5 ml tube. Cells were spun down for 15 sec at 4°C and $16100 \times g$. The supernatant was transferred to a fresh tube and centrifuged again for 5 min at 4°C and $16100 \times g$. The resulting supernatant was collected as a cytosolic extract. The pellet from the first centrifugation was lysed with 50 μ l of complete nucleic protein lysis buffer (20 mM HEPES, 420 mM KCl, 1 mM EDTA, 100 μ M Na₃VO₄, 20% glycerol, supplemented with 1% complete proteinase inhibitor cocktail, 3 mM DTT, and 0.4 mM Pefabloc, pH 7.4) and incubated for 30 min on ice, followed by centrifugation for 15 min

at 4°C and 16100 × g. The supernatant was collected as a nucleic extract. The cytosolic and nucleic extracts were pooled and stored at -80°C for further use. In total, 14 µl of 6× Laemmli buffer (350 mM Tris-HCl, 8% SDS, 30% glycerol, 10% mercaptoethanol, 0.04% bromophenol blue, pH 7.4) were added to each protein extract. Samples were then boiled at 95°C for 3 min and stored at -20°C for Western blotting analysis.

2.2.2.2 Protein extraction of HL-60 cells

HL-60 cells were harvested by centrifugation at 65 × g. Pellets were lysed in 300 µl of 2.5× protein loading buffer (200 mM Tris-HCl pH 6.8, 8% SDS (v/v), 20% glycerol (v/v), 0.4% bromophenol blue (w/v)). Protein extracts were sonicated for few pulses and subsequently incubated at 95°C for 5 minutes to complete protein denaturation. Finally, samples were stored at -20°C until further analysis.

2.2.2.3 Western blotting analysis

Protein samples were loaded into the wells of a two-phase 10% SDS-PAGE gel, along with the molecular weight marker. The stacking gel solution was made by adding 25% stacking gel buffer (500 mM Tris-HCl, 0.4% SDS, pH 6.8), 16% Rotiphorese Gel 30, 0.6% APS, and 0.2% TEMED in double-distilled H₂O. The separating gel solution was made by mixing 25% separating gel buffer (1.5 mM Tris-HCl, 0.2% SDS, pH 8.8), 33.1% Rotiphorese Gel 30, 0.03% APS, and 0.16% TEMED in double-distilled H₂O. SDS-PAGE running buffer (25 mM Tris-base, 192 mM glycine, 0.1% SDS, pH 8.6) was used to run the gels at 11 mA. A semi-dry blotting system was used at 80 mA for 90 min with transfer buffer (25 mM Tris-base, 150 mM glycine, 10% methanol, pH 8.0). A polyvinylidene difluoride (PVDF) membrane was activated with methanol for 1 min and rinsed with transfer buffer before preparing the stack. The membrane was subsequently blocked for 1 h at room temperature using blocking buffer, 4% BSA in Tris-buffered saline (TBS: 137 mM NaCl, 10 mM Tris-HCl, pH 7.4) supplemented with 0.1% Tween-20 (TBS-T). The membrane was incubated with the appropriate dilutions of primary antibody in blocking buffer overnight at 4°C. The blot was washed five times for 5 min with TBS-T before being incubated with the diluted secondary antibody (prepared in blocking buffer) for 1 h at room temperature in the dark. The membrane was washed five times with TBS-T and once with TBS, 5 min each. For signal reading, the blot was scanned using the LI-COR Odyssey CLx imaging system. For reblotting, the membrane was washed three times for 15 min in TBS-T, then incubated overnight with a new primary antibody for next day development. When necessary, the blot was stripped with stripping buffer (62.5 mM Tris-HCl, 2% SDS, 0.7% mercaptoethanol, pH 6.8) at 60°C for 1 h.

Afterward, it was thoroughly washed with tap water followed by four times TBS-T for 15 min. The blot was blocked with blocking buffer for 1 h at room temperature and incubated overnight with a new primary antibody. The dilutions of used antibodies were as follows: anti-ezrin 1:1000, anti-GAPDH 1:5000, anti- β -actin 1:5000, anti-phosphotyrosine-STAT1 1:1000, anti-phosphotyrosine-STAT3 1:1000, anti-STAT1 1:1000, and anti-STAT3 1:1000. The IRDye 800CW-conjugated donkey anti-rabbit secondary antibody used for detection was diluted 1:15000, and IRDye 800CW-conjugated donkey anti-mouse secondary antibody was diluted 1:15000.

2.2.2.4 Electrophoretic mobility shift assay

Native protein samples were incubated with radioactively labelled DNA probes in the presence of the non-specific competitor poly-dIdC. A native polyacrylamide gel was used to run the samples. The reaction mix contained 0.2 μ l of the DNA probe, which was added to 1 μ l poly-dIdC (2 mg/ml), 1.3 μ l DTT (100 mM), 3.5 μ l H₂O, and 2.5 μ l 5 \times EMSA loading buffer (100 mM HEPES, 200 mM KCl, 5 mM MgCl₂, 2.5 mM EDTA, 0.5 mM EGTA, 20% Ficoll). The reaction mix was incubated for the indicated time points with 4.5 μ l of protein extracts. In supershift reactions, the STAT1 C-24 antibody was used, while the STAT3 H-190 antibody was used as a negative control. The reactions were loaded onto an equilibrated non-denaturing 4.8% TBE-polyacrylamide gel (12% Rotiphorese Gel 40, 2.4% TBE (89 mM Tris-base, 89 mM boric acid, 2 mM EDTA), 2% APS, 0.1% TEMED) and run using 0.25 \times TBE at 400 V. After the run, Whatman blotting paper was pressed against the gel and the gel was then vacuum-dried. The radioactive signal was recorded using a phospho-imager film and read by FLA-5100 scanner using TINA v 2.0 software.

2.2.2.5 Radioactive labelling of EMSA probes

Oligonucleotides were diluted at a concentration of 100 mmol/l in DNase-, RNase- and proteinase-free water and incubated at 95°C in a water bath. Mixtures were cooled overnight at room temperature for proper annealing and stored at -20°C. Five units of Klenow fragment from *E. coli* DNA polymerase I were mixed with 5 μ l of 10 \times Eco-Pol buffer, 0.1 ng annealed oligonucleotides, and 6 μ l of [³³P]-labelled ATP and incubated for 25 min at room temperature. Following that, an excess of non-radioactive dNTPs (6.5 mM of each dNTP) was added for 5 min. The reaction was stopped by the addition of 1 μ l of 0.5 mM EDTA. Free nucleotides were removed by centrifugation at 700 \times g for 3 min.

2.2.3 DNA analysis

2.2.3.1 DNA isolation from mouse tails

Tail biopsy samples were lysed in 500 µl of tail extraction buffer (100 mM Tris, 5 mM EDTA, 200 mM NaCl, 0.2% SDS, pH 8.5) with 10 µl proteinase K (20 mg/ml) at 56°C overnight under continuous shaking at 250 rpm. Proteinase K was inactivated by incubating the samples for 5 min at 90°C. Samples were centrifuged at $16100 \times g$ in 10°C for 15 min. 350 µl of the resulting supernatant was collected for the DNA precipitation step by adding 500 µl of isopropanol. The samples were vortexed and subsequently centrifuged for 5 min at $16100 \times g$. The supernatant was removed, and the pellet was washed twice with 70% ethanol. The air-dried pellets were then resuspended in 50 µl of TE buffer (10 mM Tris-base, 1 mM EDTA, pH 7.5). Heating of DNA samples was performed at 48°C for 30 min. The samples were stored at 4°C overnight to achieve a complete resuspension in TE buffer.

2.2.3.2 Mice genotyping

Genotyping of STAT1-F77A knock-in mice was performed by combining PCR with a restriction fragment length polymorphism (RFLP) technique. The amplified fragment was generated by mixing the following components in a single reaction mix for each sample: the primer pair *stat1*-F77A at a final concentration of 0.4 µmol/l, 2.5 µl of 10× reaction buffer, 0.5 µl of 50 mM MgCl₂ solution, 1 µl of dNTP mix (10 µmol/l), 17.25 µl of DNase-, RNase- and proteinase-free water, and 0.25 µl of Taq polymerase. In total, 1.5 µl of purified DNA solution was added to each reaction. The following PCR program was run for the reaction mixtures: 95°C for 3 min, 41 cycles of denaturation at 95°C for 15 s, annealing at 61°C for 15 s, and extension at 72°C for 15 sec and a final elongation step at 72°C for 7 min. A 1% agarose gel was run at 85 mA for 45 min to detect the generated PCR product with the UVsolo TS gel documentation system. Digestion of the fragment was performed by incubating 5 µl of the product with 0.3 µl of *Tsp509I*, 1.5 µl of reaction buffer B and 7.2 µl of DNase-, RNase- and proteinase-free water at 65°C overnight. The digestion reaction mix was analyzed with 3% agarose gel run at 85 mA for 60 min. As the F77A mutation causes a loss of *Tsp509I* site, a 234 bp fragment was detected in homozygous mice, while the wildtype-derived PCR product showed two fragments of equal size.

2.2.4 RNA analysis

2.2.4.1 RNA extraction from cell culture and reverse transcription

Cells were cultured in a starvation medium for 16 h and then stimulated with the respective cytokine for the specified time duration. PeqGold total RNA kit (VWR) was used for RNA isolation. For adherent cells, culture medium was removed and cells were washed with PBS. 400 μ l of RNA Lysis Buffer T were added directly to the cells on ice. The lysate was transferred directly into a DNA Removing Column placed in a 2.0 ml Collection Tube and centrifuged at $12000 \times g$ for 1 min at room temperature. The flow-through lysate was transferred into a new 1.5 ml tube and an equal volume of 70% ethanol was added to the lysate and mixed thoroughly by vortexing. The lysate was then added directly to the membrane of a PerfectBind RNA column in a new 2.0 ml collection tube. This assembly was centrifuged at $10000 \times g$ for 1 min. The flow-through liquid and the collection tube were discarded. The column was placed in a fresh 2.0 ml collection tube and washed by adding 500 μ l of RNA Wash Buffer I followed by a centrifuge step for 15 sec at $10000 \times g$. A 600 μ l of completed RNA Wash Buffer II was added to the column and centrifuged for 15 sec at $10000 \times g$. The flow-through liquid was discarded, and the wash step was repeated. The column was placed in the collection tube and centrifuged for 2 min at $10000 \times g$ to completely dry the column matrix. To elute RNA, the column was placed into a fresh 1.5 ml microcentrifuge tube and 50 μ l of sterile RNase-free water were added directly to the binding matrix and centrifuged for 1 min at $5000 \times g$. The eluted RNA was used directly for cDNA synthesis or stored at -80°C . Verso cDNA Synthesis kit was used to reverse transcribe the RNA to create cDNA, according to the manufacturer's instructions. The reaction mixture was as follows: 4 μ l of $5\times$ cDNA synthesis buffer, 2 μ l of dNTP mix, 1 μ l of RNA primer (anchored Oligo dT), 1 μ l of RT Enhancer, 1 μ l of Verso Enzyme Mix, and 3 μ l of DNase-, RNase- and proteinase-free H_2O with 8 μ l of the purified RNA. The amount of necessary RNA had been determined experimentally. The reverse transcription cycling program was 30 min at 42°C , followed by inactivation cycle at 95°C for 2 min. The samples were stored at -20°C .

2.2.4.2 Real-time PCR

Real-time reverse transcription PCR was used to enable detection and measurement of gene transcription products. The PCR cycling program was as follows: 95°C for 4 min, 38 cycles of denaturation at 95°C for 45 sec, annealing at 61°C for 45 sec, and extension at 72°C for 120 sec, with a final elongation at 72°C for 10 min. A master mix was prepared for each reaction containing 10 μ l of SYBR green, 0.28 μ l of each primer (5 $\mu\text{mol/l}$), and 9.44 μ l of

H₂O. Using a semi-skirted 96-well-plate, 19 µl of the master mix was distributed to each well with 1 µl of the cDNA reaction sample. GAPDH or HPRT were used as a reference housekeeping gene and each sample was run in duplicate. The $2^{-\Delta\Delta CT}$ method was used to analyze the data exported from the realplex ep program.

2.2.5 Animal experiments

2.2.5.1 Left anterior descending coronary artery ligation in mice

STAT1-F77A knock-in mice were used in the myocardial infarction model (LAVES approved with reference number 3392 42502-04-13/1226). Surgery was performed on healthy young female animals at the age of 8-12 weeks. To provide basic analgesia, metamizol in a final concentration of 2 mg/ml was added to drinking water three days before surgery. Mice were intraperitoneally anaesthetized with 10 µl/g body weight of 5% Cepetor, 10% midazolam, and 10% fentanyl in 0.9% NaCl solution and stabilized in the supine position. A small incision of the trachea was introduced to insert a tube for artificial respiration (150 µl volume and 150 strokes/min). To expose the heart, a lateral cut of the pectoral muscles was carried to expose the rib cage and spread the fourth intercostal space. Using Ethilon 9-0 BV-4 5.0 mm 3/8c EH7448G surgical silk, ligation of the left anterior descending artery was performed to induce infarction. Cardiac apex started discoloration into white as a measure of proper occlusion of the coronary vessel. Sham-operated mice underwent the same procedure except occlusion of the left anterior descending artery. Following successful ligation, retractors were disconnected, and suturing of the skin was performed using Prolene 6-0 C-1 13 mm 3/8c 8889H surgical silk. Intraperitoneal injection of 10 µl/g body weight of narcosis antagonist, 5% atipamezole, 10% flumazenil, in sterile 0.9% NaCl was administered. Mice were given 1% buprenorphin in sterile 0.9% NaCl at a dose of 15 µl/g body weight and placed on a pre-warmed plate at 37°C until awake. Mice were sacrificed through CO₂ asphyxiation and hearts were collected at day 1 following the surgery. The whole heart was washed with 1× PBS and the infarcted areas from the left ventricle were excised with a surgical blade. Samples were collected in TRIzol and stored at -80°C until further analysis.

2.2.5.2 Immunohistochemistry of paraffin-embedded sections

Mice were sacrificed through CO₂ asphyxiation. In LAD ligation experiments, the heart was surgically removed and washed in PBS before being stored in 4% formaldehyde solution at 4°C. After 24 hours, organs were transferred from paraformaldehyde (PFA) into PBS and stored at 4°C until further processing. For paraffin embedding, organs were prepared through

dehydration using ascending concentrations of ethanol and finally pure xylene. Casting paraffin-embedded organs in blocks was performed using the EG1160 tissue embedder. The blocks were then cut into sections at 3 μm thickness using the SM 2000 R microtome. Tissues were left to stretch in a water bath at 45°C before transferring them into a slide to dry at 50°C. To monitor protein expression and localization of proteins in tissues, deparaffinization and rehydration were performed by xylene incubation for 15 min twice, then by using descending concentrations of ethanol (99%, 96%, 70%, desalted H_2O) twice for 2 min per each solution. Citrate buffer (8.13 mM sodium citrate, pH 6.0) was used for boiling the samples for 15 min in a microwave. Samples were cooled on ice followed by washing with double-distilled H_2O followed by PBS. A solution of 3% H_2O_2 in PBS was used for 20 min at 4°C to block peroxidase activity in the tissue sections. A blocking solution consisting of 10% FBS in PBS, supplemented with 0.05% Triton X-100, was used to incubate the slides for 1 h at room temperature. Primary antibodies were incubated in 10% FBS in PBS overnight at 4°C. Tissue slides were washed three times in PBS before secondary antibody incubation, which was diluted in 10% FBS in PBS at 1:1000 for 1 h at room temperature. Tissue slides were washed four times with PBS followed by incubation for 1 h with streptavidin/peroxidase solution diluted at 1:1000 in PBS. A 3,3'-diaminobenzidine (DAP) staining was used at development stage (2.5% stock was used by diluting it 1:250 in PBS and adding 0.01% H_2O_2). Reaction time was between 5 to 10 min and was stopped by rinsing with double-distilled water. Slides were incubated in filtered Mayer's haemalum solution for 10 min followed by washing in running tap water for 10 min. Slides were then washed twice with desalted water and dehydrated with ascending concentrations of ethanol (70%, 96%, 99%) twice for 2 min each. Tissues were incubated in xylene for 15 min twice. Entellan was used to mount the samples to be examined using a light microscope. Dilutions of primary antibodies were as the following: anti-STAT1; 1:200, anti-STAT3; 1:200.

Table 18: Evaluation of STAT staining distribution and intensity in heart tissue

| | |
|---|---|
| 0 | No specific staining |
| 1 | Less than half of the left ventricle |
| 2 | 50% to 90% of the left ventricle |
| 3 | Entire left ventricle but little or no infiltration of the septum or right ventricle |
| 4 | Entire left ventricle and moderate to significant infiltration of the right ventricle or the septum |

| | |
|---|--|
| 0 | No specific staining |
| 1 | Single cells at low density |
| 2 | Small accumulations, moderate density |
| 3 | Many or large accumulations |
| 4 | High, even density of positively stained cells |

2.2.6 RNA sequencing and analysis

Sequencing of RNA samples was conducted at the Microarray and Deep-Sequencing Facility (Transcriptome and Genome Analysis Laboratory (TAL), Göttingen). RNA was extracted using TRIzol isolation protocol. Quality and integrity of RNA samples were assessed with a fragment analyzer using the standard sensitivity RNA analysis kit. All samples selected for sequencing exhibited an RNA integrity number over 8. For library preparation, 500 ng of total RNA was used. Samples were immediately fragmented (300 bp) and cDNA subsequently synthesized. RNA-seq libraries were performed using TruSeq stranded mRNA prep kit with a modified strand-specific massively-parallel cDNA sequencing mRNA-seq protocol. The protocol was first optimized during the ligation step by diluting the adapters concentration to increase ligation efficiency (>94%), and finally reducing the number of PCR cycles to avoid PCR duplication artifacts as well as primer dimers in the final library product. Using strand-specificity protocols, differences in coverage, agreement with known annotations, and accuracy for expression profiling were observed. For accurate quantitation of cDNA libraries, a fluorometric-based system called QuantiFluor dsDNA was used. The size of final cDNA libraries was determined by using the dsDNA 905 reagent kit, exhibiting a sizing of 300 bp in average. Libraries were pooled and sequenced on an Illumina HiSeq 4000 generating 50 bp single-end reads (30-40 million reads/sample). Generation of raw data files and their quality check were performed, and sequence images were transformed with BaseCaller to bcl files,

which was demultiplexed to FASTQ files with bcl2fastq. The sequencing quality was asserted using FastQC. Mapping of sequenced reads and read counting for all samples was performed. Sequences were aligned to the reference genome *Mus musculus* mm10 (version 89) using the STAR aligner, allowing for 2 mismatches within 50 bases (Dobin *et al.*, 2013). Subsequently, read counting was performed using featureCounts (Liao *et al.*, 2014). Differential expression analysis was also performed and read counts were analyzed in the R/Bioconductor environment (version 3.4.2) using DESeq2 package (Love *et al.*, 2014). Candidate genes were filtered using an absolute \log_2 fold-change >1 and FDR(False Discovery Rate)-corrected p -value <0.05 . Gene annotation was performed using *Mus musculus* entries via biomaRt R package (Durinck *et al.*, 2009). The following R packages were used for gene set enrichment analysis and data visualization: ggplot2, kableExtra, pheatmap, d3heatmap, gage, dplyr, pathview, biomaRt, grid, VennDiagram, and ggrepel.

2.2.7 Parallel plate flow chamber assay

All cells were resuspended in HL-60 media (RPMI-1640, 10% FCS, 1% penicillin/streptomycin) supplemented with 25 mM HEPES buffer prior to the perfusion assay. Addition of HEPES provided supplemental buffering at a pH 7.2 through 7.6, as CO₂ supply was absent in the assay. A monolayer of human umbilical vein endothelial cell (HUVEC) was grown to confluency and stimulated overnight with 10 ng/ml TNF- α . On the next day, analysis of HL-60 cells interacting with HUVEC was performed at 37°C in a 5% CO₂ incubator and visualized using Olympus IX81 time-lapse inverted fluorescence microscope linked to a Hamamatsu C10600 ORCA-R2 digital video camera. A Harvard syringe pump generated a physiological shear stress of blood flow with a flow rate of 0.30 ml/min which corresponds to 1.5 dyn/cm³. HL-60 cells were perfused at a density of 1×10^6 cells/ml over TNF- α activated HUVEC, grown in plastic dishes for 25 minutes. The footage was recorded from the moment the cells were visible using a 4 \times objective. Images were saved into a video file using Volocity Imaging software.

2.2.8 Statistical analysis

IBM SPSS and R statistical software were used. Descriptive statistics were calculated from the data (e.g. mean, confidence interval of the mean, median, minimum, maximum, first quantile, third quantile, standard deviation, and standard error). A t-test for independent samples was used to compare the means for two groups. A p -value of 0.05 was chosen as level of significance.

3. Results

3.1 Echocardiographic analysis demonstrated beneficial outcomes in STAT1-F77A mice after myocardial infarction

The left ventricular size and function of STAT1-F77A and wild-type mice were assessed using sham-operated mice (as control) or mice that were subjected to ligation of the left anterior descending coronary artery (LAD), as published in Theresa Riebling's doctoral thesis, summarized in Tables 21 and 22 and depicted in Figure 4. The left ventricular end-systolic inner diameter in STAT1-F77A mice was significantly decreased as compared to wild-type mice on the third day (mean=3.92 mm, 95%-CI=3.59–4.25 mm vs mean=4.39 mm, 95%-CI=4.15–4.64 mm, $p=0.036$) and at the end of the first week (mean=4.38 mm, 95%-CI=3.70–5.05 mm vs mean=5.40 mm, 95%-CI=4.60–6.20 mm, $p=0.049$) after LAD ligation. Similarly, the left ventricular end-systolic area in STAT1-F77A mice was significantly lower as compared to wild-type mice on the third day (mean=10.24 mm², 95%-CI=8.92–11.55 mm² vs mean=12.79 mm², 95%-CI=11.54–14.05 mm², $p=0.0068$) and at the end of the first week (mean=11.55 mm², 95%-CI=7.79–15.31 mm² vs mean=18.09 mm², 95%-CI=13.54–22.65 mm², $p=0.03$) after LAD ligation. The systolic epicardial area was reduced at day 7 post-MI compared to wild-type mice (mean=22.6 mm², 95%-CI=18.36–26.84 mm² vs mean=29.52 mm², 95%-CI=24.32–34.71 mm², $p=0.039$). Additionally, the left ventricular end-diastolic inner diameter in STAT1-F77A mice was significantly decreased on the third day compared to wild-type mice (mean=4.39 mm, 95%-CI=4.14–4.64 mm vs mean=4.79 mm, 95%-CI=4.64–4.94 mm, $p=0.0035$). Correspondingly, the end-diastolic area in STAT1-F77A mice was significantly lower on the third day compared to wild-type mice (mean=13.57 mm², 95%-CI=12.17–14.98 mm² vs mean=15.83 mm², 95%-CI=14.81–16.85 mm², $p=0.013$).

Table 19 illustrates the most common parameters used to evaluate the function of the left ventricle including the equations used to calculate them, which are fractional area shortening (FAS), fractional shortening (FS), end-systolic volume (Vol_s), end-diastolic volume (Vol_d), and ejection fraction (EF). In our data, the fractional area shortening of the left ventricle in STAT1-F77A mice was significantly increased at the end of the first week following myocardial infarction as compared to wild-type mice (mean=26.92%, 95%-CI=20.36–33.48% vs mean=16.13%, 95%-CI=9.70–22.56%, p=0.024). Likewise, the ejection fraction of the left ventricle in STAT1-F77A mice was significantly improved at day 7 following MI as compared to wild-type mice (mean=30.11%, 95%-CI=22.64–37.57% vs mean=18.6%, 95%-CI=11.26–25.95%, p=0.03). Furthermore, the end-systolic volume of the left ventricle in STAT1-F77A mice was significantly decreased as compared to wild-type mice on the third day (mean=62.44 mm³, 95%-CI=52.42–72.45 mm³ vs mean=79.49 mm³, 95%-CI=69.93–89.05 mm³, p=0.039) and at the end of the first week (mean=72.00 mm³, 95%-CI=42.88–101.12 mm³ vs mean=119.69 mm³, 95%-CI=84.14–155.23 mm³, p=0.039). Moreover, the end-diastolic volume of the left ventricle in STAT1-F77A mice was significantly lower on the third day as compared to wild-type mice (mean=85.82 mm³, 95%-CI=74.52–97.12 mm³ vs mean=101.90 mm³, 95%-CI=93.03–110.78 mm³, p=0.045).

Table 19: Equations used in the calculation of left ventricular function parameters

| | |
|------------------|---|
| FAS | $(\text{Area}_d - \text{Area}_s) / \text{Area}_d * 100$ |
| FS | $(\text{LVID}_d - \text{LVID}_s) / \text{LVID}_d * 100$ |
| Vol _s | $(5 / 6) * (\text{Area}_s * L_s)$ |
| Vol _d | $(5 / 6) * (\text{Area}_d * L_d)$ |
| EF | $(\text{Vol}_d - \text{Vol}_s) / \text{Vol}_d * 100$ |

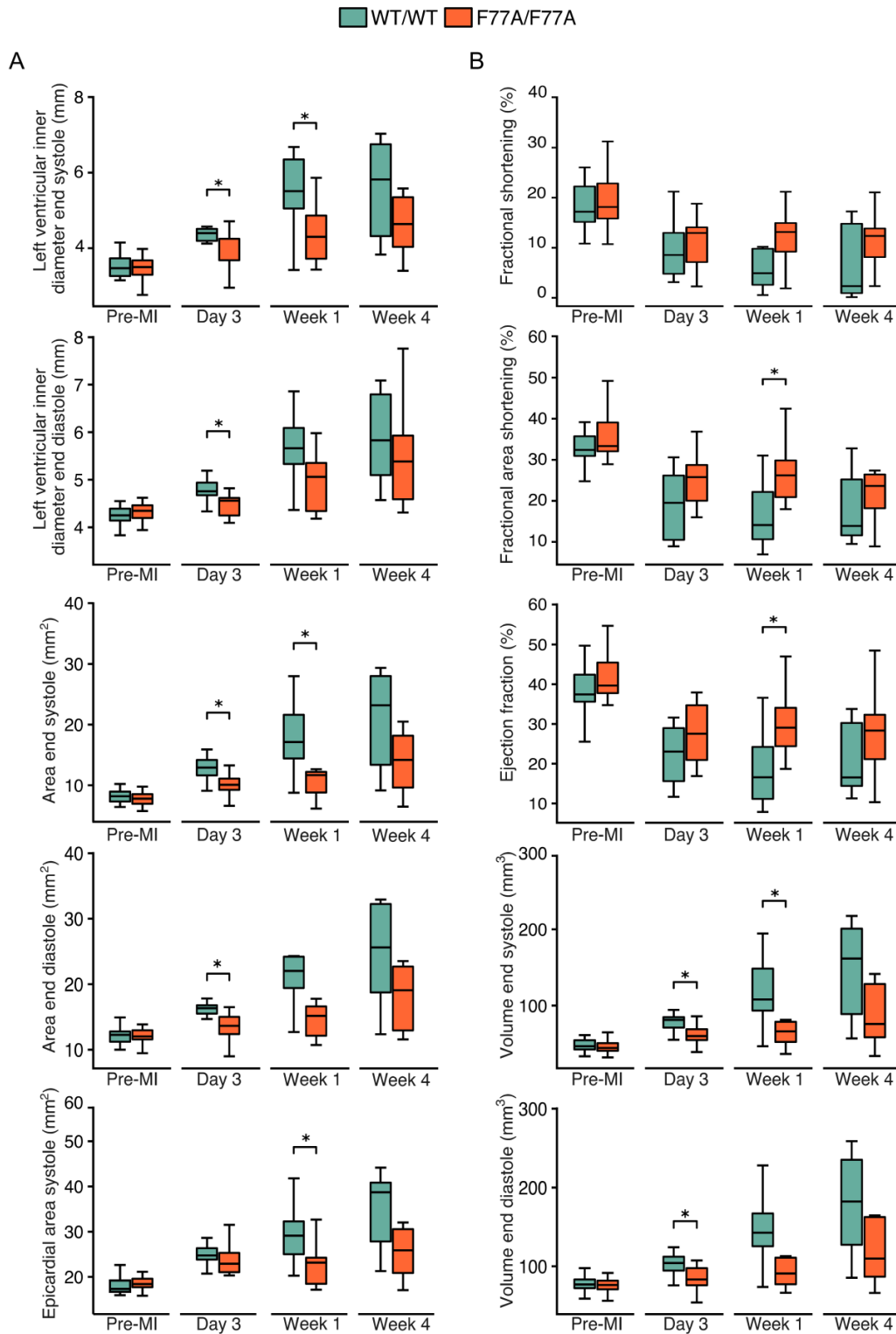


Figure 4: Echocardiographic assessment of hemodynamic parameters in wild-type and STAT1-F77A knock-in mice. Before LAD ligation ($n=23$ and $n=23$, respectively), 3 days, ($n=12$ versus $n=12$), one week ($n=9$ versus $n=8$) and four weeks ($n=7$ versus $n=8$) following LAD ligation. $*p \leq 0.05$ by independent-samples t -test.

Table 20: Measurements of important structural cardiac parameters in wild-type and STAT1-F77A knock-in mice.

| | Time | Genotype | Mean (CI) | Min | Q1 | Median | Q3 | Max | p-value |
|--|--------|------------|---------------------|-------|-------|--------|-------|-------|---------|
| End-systolic left ventricular inner diameter (mm) | Pre-MI | WT | 3.50 (3.38-3.63) | 3.15 | 3.26 | 3.47 | 3.73 | 4.15 | 0.99 |
| | | STAT1-F77A | 3.47 (3.34-3.59) | 2.76 | 3.3 | 3.5 | 3.68 | 3.98 | |
| | Day 3 | WT | 4.39 (4.15-4.64) | 3.58 | 4.19 | 4.4 | 4.51 | 5.03 | 0.036* |
| | | STAT1-F77A | 3.92 (3.59-4.25) | 2.95 | 3.68 | 3.79 | 4.25 | 4.71 | |
| | Week 1 | WT | 5.40 (4.6-6.20) | 3.42 | 5.05 | 5.51 | 6.35 | 6.68 | 0.049* |
| | | STAT1-F77A | 4.38 (3.7-5.05) | 3.43 | 3.72 | 4.3 | 4.86 | 5.86 | |
| | Week 4 | WT | 5.55 (4.26-6.83) | 3.83 | 4.32 | 5.82 | 6.75 | 7.03 | 0.52 |
| | | STAT1-F77A | 4.94 (3.75-6.13) | 3.4 | 4.03 | 4.64 | 5.35 | 7.94 | |
| End-diastolic left ventricular inner diameter (mm) | Pre-MI | WT | 4.29 (4.18-4.4) | 3.83 | 4.14 | 4.25 | 4.39 | 5.01 | 0.52 |
| | | STAT1-F77A | 4.31 (4.22-4.4) | 3.94 | 4.19 | 4.35 | 4.46 | 4.62 | |
| | Day 3 | WT | 4.79 (4.64-4.94) | 4.33 | 4.68 | 4.76 | 4.94 | 5.19 | 0.0035* |
| | | STAT1-F77A | 4.39 (4.14-4.64) | 3.46 | 4.25 | 4.56 | 4.62 | 4.82 | |
| | Week 1 | WT | 5.60 (5.05-6.16) | 4.36 | 5.33 | 5.66 | 6.09 | 6.86 | 0.061 |
| | | STAT1-F77A | 4.96 (4.43-5.49) | 4.18 | 4.34 | 5.06 | 5.36 | 5.98 | |
| | Week 4 | WT | 5.9 (4.94-6.85) | 4.57 | 5.1 | 5.83 | 6.8 | 7.09 | 0.45 |
| | | STAT1-F77A | 5.47 (4.52-6.43) | 4.31 | 4.59 | 5.39 | 5.93 | 7.76 | |
| End-systolic area (%) | Pre-MI | WT | 8.28 (7.68-8.88) | 6.43 | 7.34 | 8.2 | 8.98 | 11.96 | 0.38 |
| | | STAT1-F77A | 7.92 (7.32-8.52) | 5.77 | 6.97 | 7.8 | 8.54 | 11.55 | |
| | Day 3 | WT | 12.79 (11.54-14.05) | 9.1 | 11.64 | 12.93 | 14.17 | 15.91 | 0.0068* |
| | | STAT1-F77A | 10.24 (8.92-11.55) | 6.64 | 9.27 | 10.09 | 11.11 | 13.27 | |
| | Week 1 | WT | 18.09 (13.54-22.65) | 8.76 | 14.42 | 17.14 | 21.62 | 27.96 | 0.030* |
| | | STAT1-F77A | 11.55 (7.79-15.31) | 6.17 | 8.83 | 11.7 | 12.22 | 21.12 | |
| | Week 4 | WT | 20.64 (12.79-28.50) | 9.18 | 13.38 | 23.19 | 28.01 | 29.35 | 0.38 |
| | | STAT1-F77A | 16.43 (7.76-25.10) | 6.51 | 9.63 | 14.18 | 18.19 | 39.39 | |
| End-diastolic area (mm ²) | Pre-MI | WT | 12.2 (11.6-12.81) | 10.01 | 11.2 | 12.27 | 12.79 | 15.89 | 0.95 |
| | | STAT1-F77A | 12.22 (11.55-12.90) | 9.47 | 11.56 | 12.01 | 12.96 | 16.62 | |
| | Day 3 | WT | 15.83 (14.81-16.85) | 12.3 | 15.52 | 16.34 | 16.79 | 17.81 | 0.013* |
| | | STAT1-F77A | 13.57 (12.17-14.98) | 9 | 12.39 | 13.63 | 15 | 16.49 | |
| | Week 1 | WT | 21.29 (16.79-25.79) | 12.7 | 19.4 | 22.02 | 24.2 | 32.24 | 0.061 |
| | | STAT1-F77A | 15.54 (11.5-19.58) | 10.72 | 12.13 | 15.15 | 16.6 | 25.74 | |
| | Week 4 | WT | 24.68 (17.06-32.31) | 12.34 | 18.73 | 25.62 | 32.24 | 32.91 | 0.27 |
| | | STAT1-F77A | 20.52 (11.86-29.19) | 11.58 | 12.95 | 19.07 | 22.71 | 43.23 | |
| Epicardial systolic area (mm ²) | Pre-MI | WT | 18.19 (17.4-18.98) | 15.93 | 16.73 | 17.34 | 19.23 | 22.66 | 0.29 |
| | | STAT1-F77A | 18.61 (17.78-19.44) | 14.57 | 17.71 | 18.4 | 19.58 | 23.67 | |
| | Day 3 | WT | 25.02 (23.57-26.47) | 20.73 | 23.83 | 24.75 | 26.35 | 28.62 | 0.17 |
| | | STAT1-F77A | 23.21 (20.18-26.24) | 14.12 | 21.04 | 22.91 | 25.33 | 31.5 | |
| | Week 1 | WT | 29.52 (24.32-34.71) | 20.27 | 25.03 | 29.1 | 32.28 | 41.8 | 0.039* |
| | | STAT1-F77A | 22.6 (18.36-26.84) | 17.17 | 18.47 | 23.15 | 24.22 | 32.66 | |
| | Week 4 | WT | 34.49 (26.27-42.72) | 21.26 | 27.83 | 38.71 | 40.83 | 44.16 | 0.15 |
| | | STAT1-F77A | 28.97 (17.9-40.04) | 17.07 | 20.87 | 25.86 | 30.56 | 59.27 | |

Before MI (n=23 and n=23, respectively), and at different time points after MI: 3 days, (n=12 versus n=12), one week (n=9 versus n=8) and four weeks (n=7 versus n=8). Mean, lower and upper quartile of the mean, minimum, first (Q1) and third (Q3) quartiles, median, and maximum are shown below.

Table 21: Assessment of important functional cardiac parameters in wild-type and STAT1-F77A knock-in mice

| | Time | Genotype | Mean (CI) | Min | Q1 | Median | Q3 | Max | p-value |
|---|--------|------------|------------------------|-------|--------|--------|--------|--------|---------|
| Fractional shortening (%) | Pre-MI | WT | 18.35 (16.43-20.28) | 10.84 | 15.16 | 17.19 | 22.27 | 26.06 | 0.49 |
| | | STAT1-F77A | 19.54 (17.19-21.90) | 10.74 | 15.84 | 18.14 | 22.84 | 31.27 | |
| | Day 3 | WT | 9.47 (5.85-13.08) | 3.11 | 4.81 | 8.56 | 12.98 | 21.21 | 0.56 |
| | | STAT1-F77A | 10.88 (7.06-14.71) | 2.28 | 7.13 | 12.96 | 14.1 | 18.8 | |
| | Week 1 | WT | 6.79 (1.82-11.77) | 0.55 | 2.58 | 4.93 | 9.81 | 21.67 | 0.11 |
| | | STAT1-F77A | 12.23 (7.50-16.95) | 1.88 | 9.22 | 13.13 | 14.95 | 21.19 | |
| Fractional area shortening (%) | Pre-MI | WT | 32.38 (29.86-34.91) | 21.25 | 30.93 | 32.38 | 35.73 | 45.46 | 0.22 |
| | | STAT1-F77A | 35.26 (32.23-38.29) | 18.62 | 32.04 | 33.33 | 39.07 | 49.9 | |
| | Day 3 | WT | 19.33 (14.07-24.58) | 8.9 | 10.51 | 19.53 | 26.17 | 30.58 | 0.21 |
| | | STAT1-F77A | 24.72 (20.16-29.29) | 15.97 | 20.02 | 25.76 | 28.74 | 36.84 | |
| | Week 1 | WT | 16.13 (9.7-22.56) | 6.98 | 10.65 | 14.09 | 22.14 | 31.01 | 0.024* |
| | | STAT1-F77A | 26.92 (20.36-33.48) | 17.96 | 20.88 | 26.2 | 29.84 | 42.42 | |
| Ejection fraction (%) | Pre-MI | WT | 37.82 (35.23-40.41) | 25.52 | 35.6 | 37.46 | 42.4 | 49.68 | 0.11 |
| | | STAT1-F77A | 41.08 (38.23-43.93) | 25.37 | 37.76 | 39.65 | 45.43 | 54.71 | |
| | Day 3 | WT | 22.27 (17.45-27.08) | 11.65 | 15.57 | 23.04 | 28.93 | 31.61 | 0.10 |
| | | STAT1-F77A | 27.39 (22.25-32.54) | 16.9 | 20.91 | 27.53 | 34.71 | 37.89 | |
| | Week 1 | WT | 18.60 (11.26-25.95) | 7.84 | 11.14 | 16.59 | 24.23 | 36.59 | 0.030* |
| | | STAT1-F77A | 30.11 (22.64-37.57) | 18.69 | 24.41 | 29.05 | 34.08 | 47 | |
| End-systolic volume (mm ³) | Pre-MI | WT | 48.50 (44.3-52.7) | 33.58 | 42.47 | 46.68 | 54.67 | 75.21 | 0.25 |
| | | STAT1-F77A | 45.08 (41.36-48.8) | 32.28 | 40.73 | 44.56 | 50.86 | 64.85 | |
| | Day 3 | WT | 79.49 (69.93-89.05) | 55.23 | 71.23 | 81.4 | 85.01 | 108.4 | 0.039* |
| | | STAT1-F77A | 62.44 (52.42-72.45) | 39.05 | 54.82 | 60.11 | 69.01 | 86 | |
| | Week 1 | WT | 119.69 (84.14-155.23) | 46.72 | 93.47 | 108.07 | 148.48 | 194.47 | 0.039* |
| | | STAT1-F77A | 72.00 (42.88-101.12) | 36.61 | 52.32 | 66.16 | 78.59 | 148.88 | |
| End-diastolic volume (mm ³) | Pre-MI | WT | 145.13 (83.69-206.57) | 57.03 | 88.86 | 161.71 | 200.87 | 217.71 | 0.27 |
| | | STAT1-F77A | 108.43 (37.11-179.76) | 34.04 | 58.41 | 75.94 | 128.26 | 299.99 | |
| | Pre-MI | WT | 77.69 (72.7-82.68) | 59.02 | 72.24 | 76.89 | 83.34 | 110.86 | 0.78 |
| | | STAT1-F77A | 76.46 (71.59-81.32) | 56.03 | 70.81 | 76.27 | 81.72 | 106.57 | |
| | Day 3 | WT | 101.9 (93.03-110.78) | 75.72 | 94.42 | 104.22 | 112.07 | 124.1 | 0.045* |
| | | STAT1-F77A | 85.82 (74.52-97.12) | 53.89 | 75.85 | 83.24 | 97.68 | 107.4 | |
| End-diastolic volume (mm ³) | Week 1 | WT | 143.94 (108.35-179.53) | 73.67 | 125.49 | 142.63 | 167.09 | 228.05 | 0.061 |
| | | STAT1-F77A | 100.36 (68.71-132.01) | 66.42 | 77.08 | 90.74 | 110.85 | 183.09 | |
| | Week 4 | WT | 178.84 (116.48-241.2) | 85.47 | 127.35 | 182.2 | 235.29 | 258.94 | 0.22 |
| | | STAT1-F77A | 139.31 (66.55-212.07) | 66.05 | 86.83 | 109.76 | 162.51 | 334.46 | |

Before MI ($n=23$ and $n=23$, respectively), and at different time points after MI: 3 days, ($n=12$ versus $n=12$), one week ($n=9$ versus $n=8$) and four weeks ($n=7$ versus $n=8$). Mean, lower and upper quartile of the mean, minimum, first (Q1) and third (Q3) quartiles, median, and maximum are shown below.

3.2 Expression of STAT1 and STAT3 in both cardiomyocytes and infiltrating immune cells after myocardial infarction

In order to analyze the expression profile of STAT1 and STAT3 proteins, immunohistochemical staining was performed in sections of infarcted area for STAT1-F77A and wild-type mice undergoing LAD ligation or sham operation. Immunohistochemical analysis of staining intensity and distribution revealed expression of STAT1 and STAT3 in both cardiomyocytes and infiltrating immune cells at day 3 post-MI, while no significant differences in their expression levels were identified among the two groups (Figure 5).

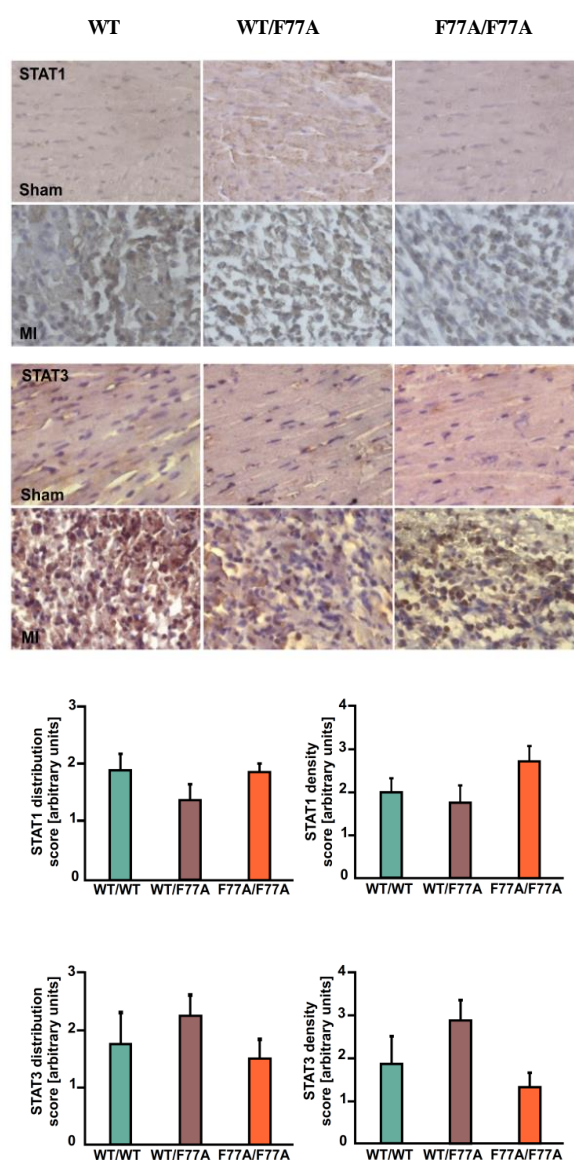


Figure 5: Immunohistochemical analysis of STAT1 and STAT3 expression in the infarcted area at day 3 post-MI. Semi-quantitative analysis of STAT staining distribution and intensity are presented as means \pm SD.

3.3 Transcriptomic analysis of F77A mice in a LAD ligation model

With the aim to analyze the left ventricular global transcriptional profile, we constructed 12 RNA libraries and sequenced them (Figure 6). Samples were collected from sham-operated or LAD-ligation-operated mice expressing either wild-type or mutant STAT1. The quality controls as well as the number of pooled mice for extracted RNA are shown in Table 22 and Figure 7.

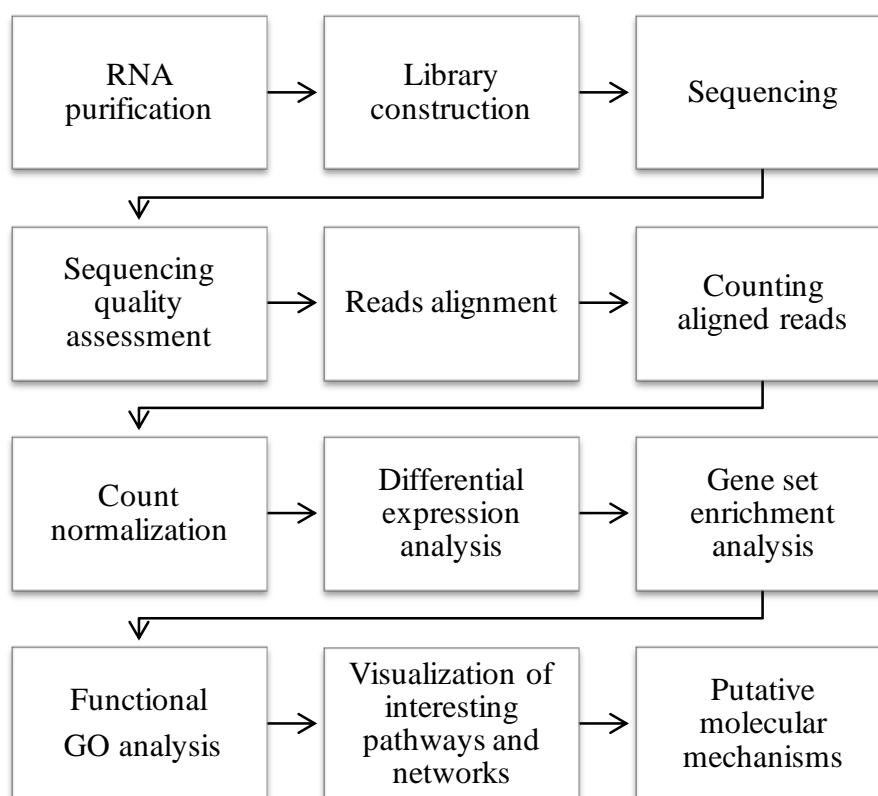


Figure 6: RNA-seq pipeline to establish transcriptomic signature of the infarcted area.

Table 22: Quality assessment of total RNA in the infarcted heart tissue samples.

| Sample ID | N | Concentration (ng/μl) | Yield (μg) | A260/A280 | A260/A230 | RIN | 28s/18s |
|-------------|---|--------------------------|------------|-----------|-----------|-----------|-----------|
| WT-Sham-1 | 4 | 711.46±197.94 | 35.57±9.9 | 1.92±0.05 | 1.61±0.33 | 9.35±0.16 | 1.40±0.19 |
| WT-Sham-2 | 4 | 728.99±68.55 | 36.45±3.43 | 1.88±0.04 | 1.35±0.28 | 9.88±0.08 | 1.33±0.22 |
| WT-Sham-3 | 3 | 910.82±130.15 | 45.54±6.51 | 1.97±0.03 | 1.73±0.33 | 10.00±0 | 1.43±0.29 |
| WT-MI-1 | 4 | 341.19±59.23 | 17.06±2.96 | 1.87±0.01 | 2.08±0.07 | 8.35±0.28 | 1.65±0.27 |
| WT-MI-2 | 4 | 524.07±80.14 | 26.20±4.01 | 1.90±0.02 | 1.66±0.25 | 9.45±0.06 | 1.73±0.2 |
| WT-MI-3 | 3 | 348.60±45.23 | 17.43±2.26 | 1.83±0.01 | 2.01±0.06 | 9.87±0.13 | 1.67±0.09 |
| F77A-Sham-1 | 2 | 493.65±12.11 | 24.68±0.61 | 1.89±0.07 | 1.54±0.52 | 9.00±0.6 | 1.40±0 |
| F77A-Sham-2 | 1 | 435.19 | 21.76 | 1.88 | 2.19 | 10.00 | 1.20 |
| F77A-Sham-3 | 1 | 768.92 | 38.45 | 1.95 | 1.44 | 10.00 | 1.70 |
| F77A-MI-1 | 4 | 316.85±59.01 | 15.84±2.95 | 1.86±0.01 | 1.70±0.24 | 8.03±0.08 | 1.35±0.05 |
| F77A-MI-2 | 3 | 331.88±82.68 | 16.59±4.13 | 1.80±0.06 | 1.61±0.51 | 8.23±0.03 | 1.27±0.09 |
| F77A-MI-3 | 3 | 378.46±114.19 | 18.92±5.71 | 1.88±0.01 | 1.63±0.25 | 8.67±0.09 | 2.80±1.55 |

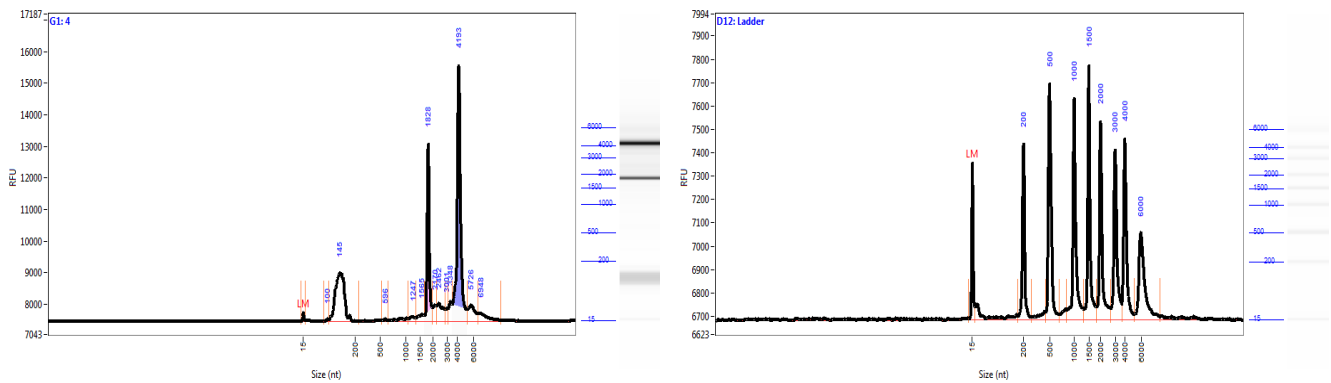


Figure 7: Separation of high-quality total RNA extracted from infarcted heart tissue. A) The purple and pink peaks represent the large (28S rRNA) and small (18S rRNA) ribosomal RNA peaks, respectively. The small RNA region includes the peaks and smears below 200 nucleotides, which is well defined, as these are the small and large rRNA subunits, providing a high RQN (RNA quality number) of 10. All samples selected for sequencing exhibited an RNA integrity number over 8. B) Separation of HS RNA Ladder on a Fragment Analyzer using the Standard Sensitivity RNA analysis kit (DNF-471).

3.4 Identification of differentially expressed genes in STAT1-F77A and wild-type mice at day 1 post-myocardial infarction

To identify differentially expressed genes (DEGs) following MI in wild-type and mutant mice, a $|\log_2(\text{fold change})| > 1$ and a $p\text{-value} < 0.05$ were used as standards. We identified 1912 differentially expressed genes in wild-type mice. However, as a consequence of LAD ligation, the number of differentially regulated genes was 4055 genes in STAT1-F77A, which is significantly higher as compared to wild-type mice. Furthermore, subsequent to MI induction, we found 194 DEGs that were unique in wild-type mice but were not significantly modulated in STAT1-F77A mice, and 2337 DEGs that are unique in STAT1-F77A mice, but not changed in wild-type mice post-MI. Moreover, we identified 1718 DEGs that were commonly expressed in wild-type and STAT1-F77A mice. Of the 2337 uniquely differentially expressed genes in the STAT1-F77A following MI, there were 909 upregulated and 1428 downregulated genes, whereas, of the 194 uniquely differentially expressed genes in wild-type animals, there were 111 upregulated and 83 downregulated genes (Figure 8). The hierarchical clustering of RPKM (reads per kilobase of transcript per million mapped reads) values of the differentially expressed genes for both wild-type (MI vs sham) and STAT1-F77A (MI vs sham) comparisons in the four groups (wild-type sham, wild-type MI, STAT1-F77A sham, STAT1-F77A MI) are shown in Figure 9.

The top 10 genes that are differentially expressed in wild-type following MI are the following: *Hspa1a* (heat shock protein family a (hsp70) member 1a), *Hspa1b* (heat shock protein family a (hsp70) member 1b), *Fgf23* (fibroblast growth factor 23), *Cxcl2* (C-X-C motif chemokine ligand 2), *Ngp* (neutrophilic granule protein), *Fosb* (fosb proto-oncogene, ap-1 transcription factor subunit), *Aa467197* (expressed sequence aa467197), *Cxcl10* (C-X-C motif chemokine ligand 10), *Ccl4* (c-c motif chemokine ligand 4), *Cd177* (human neutrophil alloantigen 2a). Table 23 illustrates the top 20 differentially expressed genes in wild-type (LAD ligation vs sham) comparison with the \log_2 fold change, padj, \log_{10} padj and baseMean values shown.

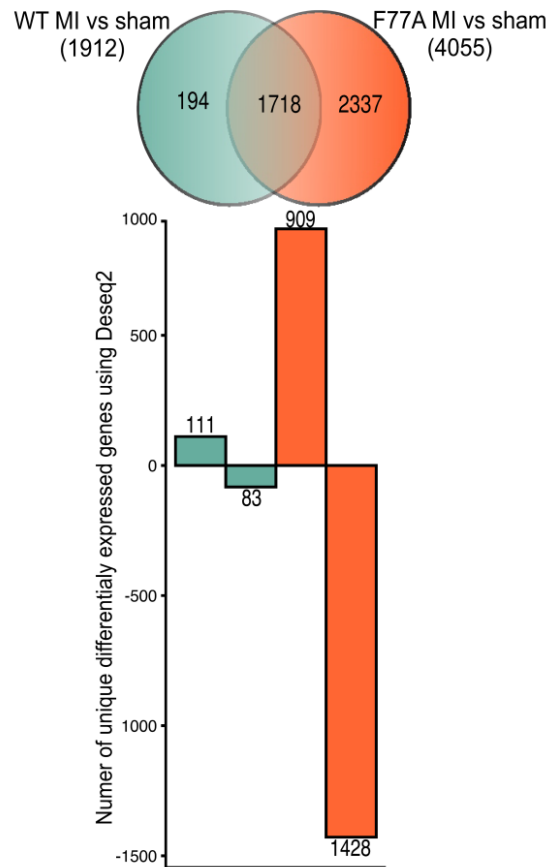


Figure 8: The number of unique genes that are upregulated or downregulated in wild-type and STAT1-F77A following LAD ligation.

Table 23: Top 20 genes differentially expressed in wild-type following LAD ligation

| Gene | log ₂ fold change | padj | log ₁₀ padj | baseMean |
|-----------------|------------------------------|----------|------------------------|----------|
| <i>Hspa1a</i> | 3.77 | 9.56e-11 | 1.00e+01 | 1851.08 |
| <i>Hspa1b</i> | 3.66 | 4.59e-10 | 9.34e+00 | 2979.38 |
| <i>Fgf23</i> | 3.65 | 1.91e-10 | 9.72e+00 | 216.89 |
| <i>Cxcl2</i> | 3.62 | 4.60e-11 | 1.03e+01 | 1167.84 |
| <i>Ngp</i> | 3.43 | 2.40e-08 | 7.62e+00 | 49.94 |
| <i>Fosb</i> | 3.39 | 1.53e-08 | 7.82e+00 | 478.94 |
| <i>AA467197</i> | 3.31 | 4.37e-08 | 7.36e+00 | 690.60 |
| <i>Cxcl10</i> | 3.28 | 3.62e-09 | 8.44e+00 | 422.35 |
| <i>Ccl4</i> | 3.27 | 1.02e-09 | 8.99e+00 | 393.55 |
| <i>Cd177</i> | 3.25 | 1.28e-09 | 8.89e+00 | 406.20 |
| <i>Thbs1</i> | 3.24 | 5.26e-11 | 1.03e+01 | 12485.02 |
| <i>Hcar2</i> | 3.23 | 1.46e-08 | 7.84e+00 | 337.96 |
| <i>Cxcl3</i> | 3.21 | 1.20e-08 | 7.92e+00 | 1422.77 |
| <i>Asprv1</i> | 3.21 | 6.28e-11 | 1.02e+01 | 290.54 |
| <i>Ptgs2</i> | 3.18 | 3.82e-08 | 7.42e+00 | 1411.21 |
| <i>Slc7a11</i> | 3.17 | 1.36e-08 | 7.87e+00 | 422.98 |
| <i>Retnlg</i> | 3.16 | 1.20e-11 | 1.09e+01 | 797.23 |
| <i>Il6</i> | 3.16 | 7.46e-09 | 8.13e+00 | 532.09 |
| <i>Selp</i> | 3.16 | 5.26e-11 | 1.03e+01 | 1177.32 |
| <i>Il1r2</i> | 3.15 | 1.98e-08 | 7.70e+00 | 1517.34 |

In addition, the top 10 genes that are differentially expressed in STAT1-F77A (LAD ligation vs sham) comparison are the following: *Fgf23* (fibroblast growth factor 23), *Cxcl3* (C-X-C motif chemokine ligand 3), *Slfn4* (schlafen 4), *Rab44* (rab44, member ras oncogene family), *Cxcl2* (C-X-C motif chemokine ligand 2), *Cd177* (human neutrophil alloantigen 2a), *Gm5483* (predicted gene 5483), *Il1r2* (interleukin 1 receptor type 2), *Cd300lf* (cd300 molecule like family member f), *Slfn1* (schlafen family member 12). Table 24 illustrates the top 20 differentially expressed genes in STAT1-F77A following LAD ligation with the log₂fold change, padj, log₁₀padj and baseMean values shown.

Table 24: Top 20 genes differentially expressed in STAT1-F77A following LAD ligation

| Gene | log ₂ fold change | padj | log ₁₀ padj | baseMean |
|----------------|------------------------------|----------|------------------------|----------|
| <i>Fgf23</i> | 4.89 | 1.29e-19 | 1.89e+01 | 216.89 |
| <i>Cxcl3</i> | 4.76 | 9.07e-20 | 1.90e+01 | 1422.77 |
| <i>Slfn4</i> | 4.64 | 8.59e-21 | 2.01e+01 | 5478.06 |
| <i>Rab44</i> | 4.52 | 1.37e-17 | 1.69e+01 | 788.41 |
| <i>Cxcl2</i> | 4.51 | 1.38e-18 | 1.79e+01 | 1167.84 |
| <i>Cd177</i> | 4.49 | 3.60e-19 | 1.84e+01 | 406.20 |
| <i>Gm5483</i> | 4.45 | 1.53e-16 | 1.58e+01 | 136.50 |
| <i>Il1r2</i> | 4.41 | 2.53e-17 | 1.66e+01 | 1517.34 |
| <i>Cd300lf</i> | 4.40 | 4.25e-22 | 2.14e+01 | 1932.80 |
| <i>Slfn1</i> | 4.34 | 9.07e-20 | 1.90e+01 | 1233.29 |
| <i>Ptgs2</i> | 4.27 | 2.51e-15 | 1.46e+01 | 1411.21 |
| <i>Cxcr2</i> | 4.19 | 3.23e-18 | 1.75e+01 | 1119.46 |
| <i>Slc7a11</i> | 4.19 | 9.62e-16 | 1.50e+01 | 422.98 |
| <i>Hcar2</i> | 4.18 | 5.19e-15 | 1.43e+01 | 337.96 |
| <i>Sh2d5</i> | 4.15 | 1.04e-13 | 1.30e+01 | 925.10 |
| <i>Adamts4</i> | 4.13 | 5.50e-14 | 1.33e+01 | 3382.60 |
| <i>Trem1</i> | 4.13 | 2.77e-17 | 1.66e+01 | 631.05 |
| <i>Hspa1a</i> | 4.12 | 5.50e-14 | 1.33e+01 | 1851.08 |
| <i>Mirt2</i> | 4.12 | 4.52e-13 | 1.23e+01 | 69.10 |
| <i>Il6</i> | 4.09 | 1.18e-15 | 1.49e+01 | 532.09 |

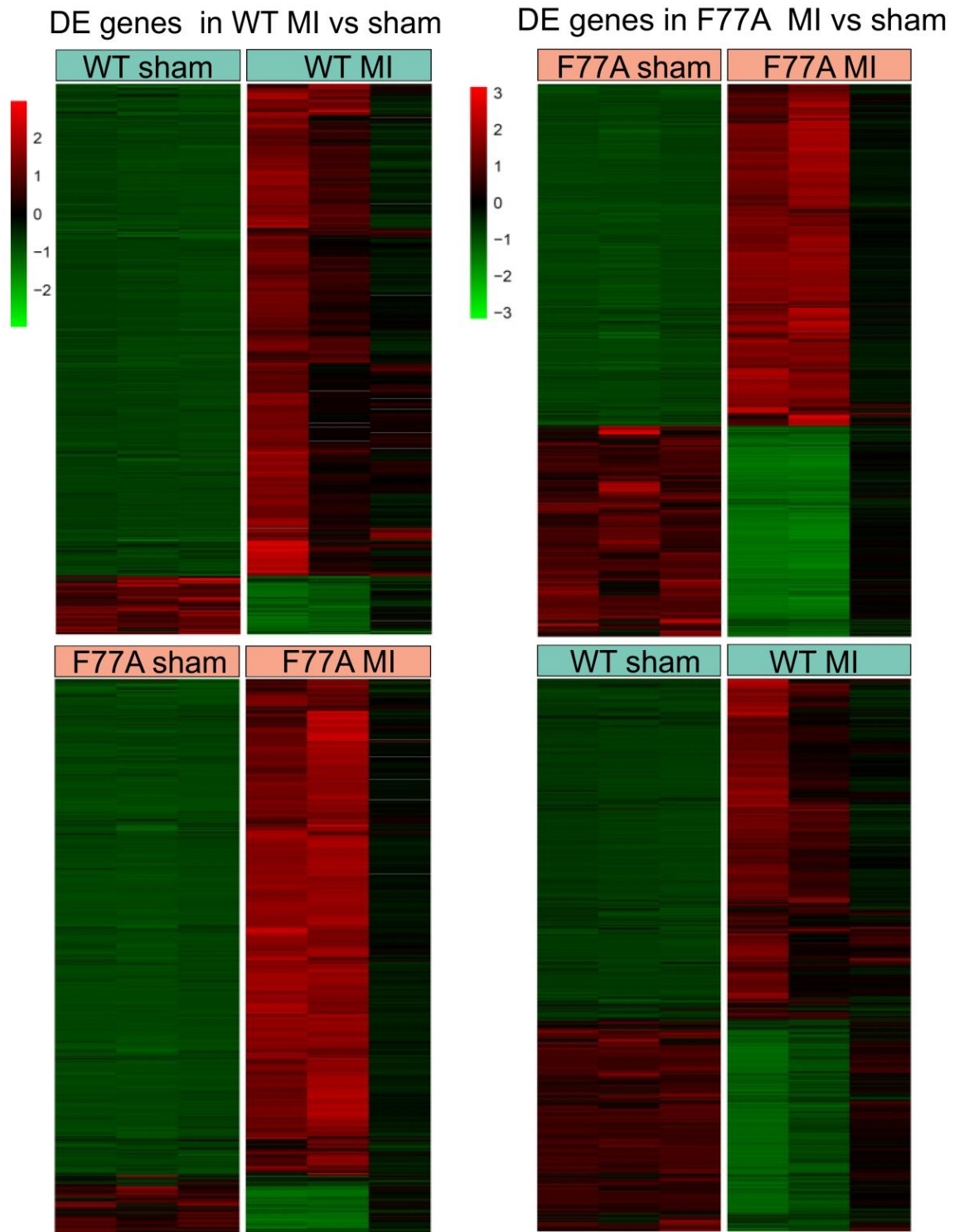


Figure 9: Hierarchical clustering of RPKM values in wild-type and STAT1-F77A mice in both sham-operated and LAD-ligation-operated mice. The first heat map was generated using the list of genes that are differentially expressed in wild-type (MI vs sham) comparison (in total 1912 genes). The second heat map was generated using genes that are differentially expressed in STAT1-F77A (MI vs sham) comparison (in total 4055 genes).

To depict the distribution of differentially regulated genes unique in unaffected myocardium and infarcted areas from the hearts of wild-type and STAT1-F77A mice, a Venn diagram representation was used. Figure 10 demonstrates the number of differentially expressed genes found in the following four groups: wild-type (MI vs sham) upregulated, wild-type (MI vs sham) downregulated, STAT1-F77A (MI vs sham) upregulated, and STAT1-F77A (MI vs sham) downregulated. The overlap of the two ovals in the top of the diagram is 1598, which is the number of differentially expressed genes that are shared between wild-type (MI vs sham) upregulated and F77A (MI vs sham) upregulated lists. The overlap of the two ovals in the bottom of the diagram is 120, which is the number of the differentially expressed genes that are shared between the wild-type (MI vs sham) downregulated and F77A (MI vs sham) downregulated lists. The areas of no overlap represent the number of uniquely (not shared) differentially expressed genes in each respective comparison.

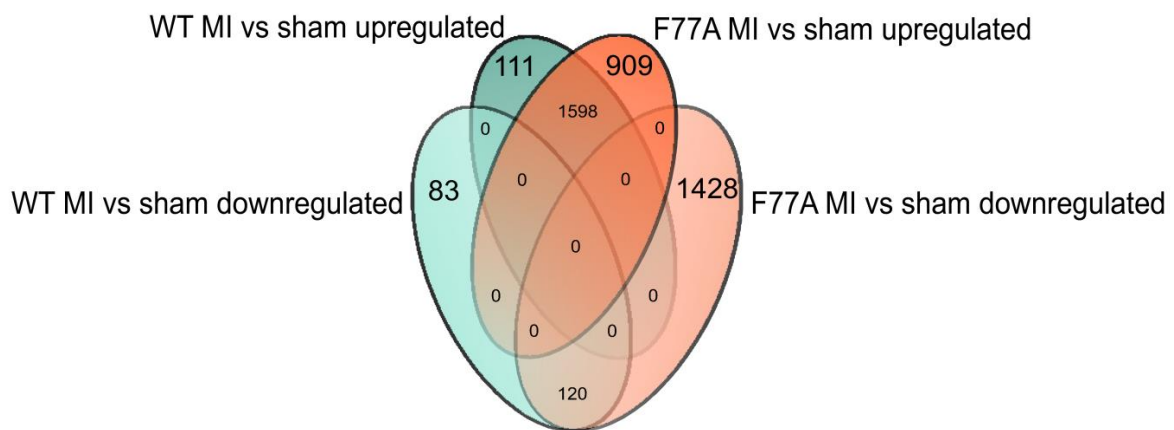


Figure 10: The numbers of differentially expressed genes in the infarcted area common and unique in wild-type and STAT1-F77A mice. *The following pairwise comparisons are shown: wild-type (MI vs sham) upregulated, wild-type (MI vs sham) downregulated, STAT1-F77A (MI vs sham) upregulated, and STAT1-F77A (MI vs sham) downregulated.*

Additionally, we used volcano plots to represent DESeq2 differential expression analysis results in animals who underwent ligation of the LAD vs sham operation in wild-type and STAT1-F77A mice, with y-axis as $-\log_{10}$ -adjusted p -value and the x-axis as the \log_2 -fold change (Figure 11 and Figure 12). In each plot, every point represents an individual transcript. The vertical lines represent a threshold of 1-fold change, either upregulated (right side) or downregulated (left side), while the horizontal lines represent an adjusted p -value <0.05 . Colored points correspond to the uniquely differentially expressed genes in the following groups: 111 genes in wild-type MI vs sham upregulated (dark-turquoise), 83 genes in wild-type MI vs sham downregulated (light-turquoise), 909 genes in F77A MI vs sham upregulated (dark-orange), 1428 genes in F77A MI vs sham downregulated (light-orange), which corresponds to Figure 10 as well.

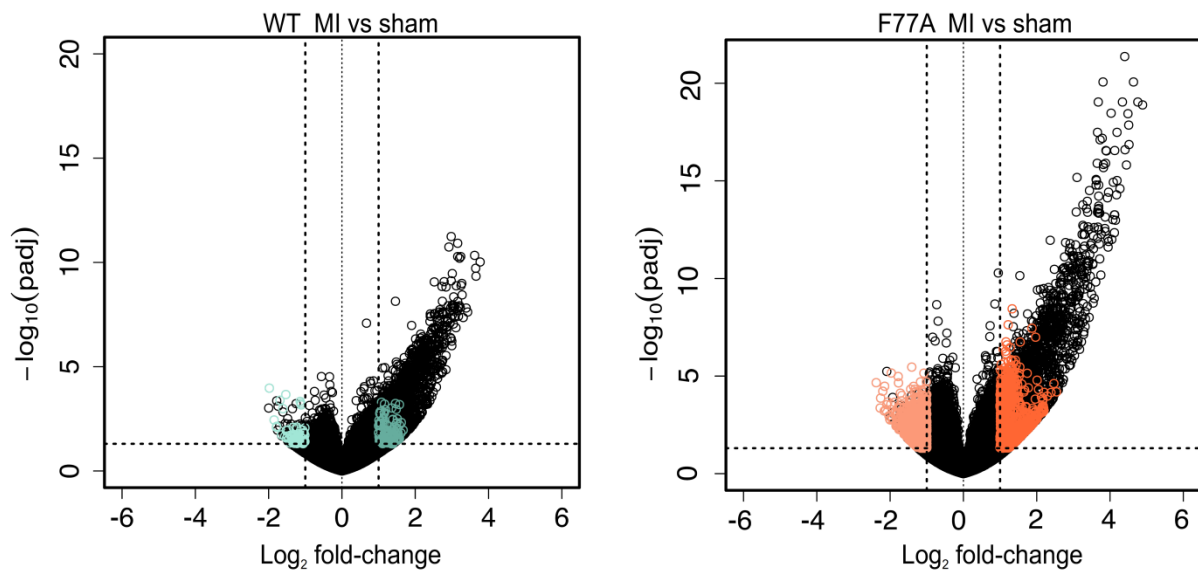


Figure 11: Volcano plot representation of differentially expressed genes in wild-type and STAT1-F77A mice post-myocardial infarction vs sham operated. Colored points correspond to uniquely upregulated or downregulated genes in the following pairwise comparisons: wild-type (MI vs sham) upregulated, wild-type (MI vs sham) downregulated, STAT1-F77A (MI vs sham) upregulated, and STAT1-F77A (MI vs sham) downregulated.

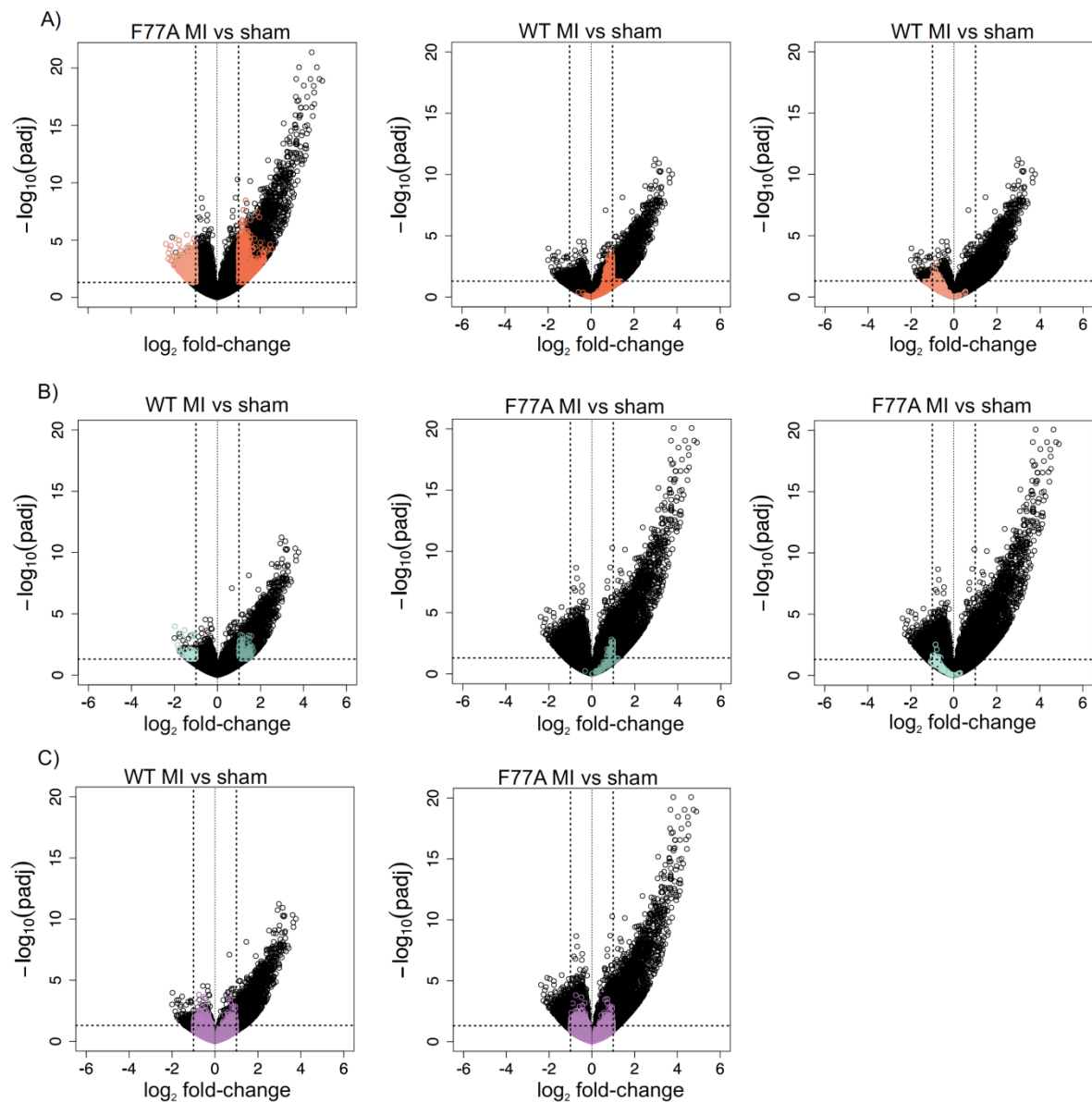


Figure 12: Volcano plot representation of uniquely differentially expressed genes in wild-type and STAT1-F77A (MI vs sham operated). (A) STAT1-F77A MI vs sham comparison. Colored points correspond to uniquely upregulated or downregulated genes in the STAT1-F77A MI vs sham comparison. The same genes are highlighted in the wild-type MI vs sham plots separately. (B) Wild-type MI vs sham. Colored points correspond to uniquely upregulated or downregulated genes in the wild-type MI vs sham comparison and the same genes are highlighted in the STAT1-F77A MI vs sham plots separately. (C) Colored points correspond to non-differentially expressed genes in the wild-type MI vs sham comparison and the same is highlighted in the F77A MI vs sham plot.

Tables 25 and 26 illustrate the top 20 uniquely upregulated or downregulated differentially expressed genes in wild-type (LAD ligation vs sham) comparison, respectively.

Table 25: Top 20 uniquely upregulated genes in wild-type following LAD ligation

| Gene | log ₂ fold change | padj | log ₁₀ padj | baseMean |
|----------------------|------------------------------|----------|------------------------|----------|
| <i>Olfm2</i> | 1.67 | 1.10e-02 | 1.96e+00 | 7.21 |
| <i>Unc13c</i> | 1.62 | 4.75e-03 | 2.32e+00 | 18.22 |
| <i>Gp5</i> | 1.61 | 1.50e-02 | 1.82e+00 | 17.47 |
| <i>Cacna1e</i> | 1.57 | 6.56e-04 | 3.18e+00 | 18.02 |
| <i>Mospd4</i> | 1.57 | 2.77e-02 | 1.56e+00 | 4.27 |
| <i>Jakmip1</i> | 1.57 | 6.29e-03 | 2.20e+00 | 15.90 |
| <i>1700016P03Rik</i> | 1.56 | 2.92e-02 | 1.53e+00 | 7.58 |
| <i>Snord58b</i> | 1.54 | 2.51e-02 | 1.60e+00 | 5.60 |
| <i>Depdc1b</i> | 1.53 | 1.47e-02 | 1.83e+00 | 6.53 |
| <i>Ighg2b</i> | 1.53 | 3.14e-02 | 1.50e+00 | 3.81 |
| <i>Crhbp</i> | 1.52 | 3.23e-02 | 1.49e+00 | 3.59 |
| <i>Col2a1</i> | 1.52 | 1.90e-02 | 1.72e+00 | 6.18 |
| <i>Lrtm2</i> | 1.52 | 1.00e-02 | 2.00e+00 | 13.59 |
| <i>Gm10288</i> | 1.51 | 2.55e-02 | 1.59e+00 | 9.14 |
| <i>Igha</i> | 1.51 | 3.18e-03 | 2.50e+00 | 46.31 |
| <i>Spic</i> | 1.50 | 8.90e-03 | 2.05e+00 | 11.16 |
| <i>Igsf11</i> | 1.50 | 4.84e-03 | 2.32e+00 | 44.37 |
| <i>Gm17619</i> | 1.50 | 3.40e-02 | 1.47e+00 | 4.77 |
| <i>Gm23301</i> | 1.48 | 5.33e-03 | 2.27e+00 | 18.45 |
| <i>Cebpd</i> | 1.48 | 1.11e-03 | 2.95e+00 | 1707.34 |

Table 26: Top 20 uniquely downregulated genes in wild-type following LAD ligation

| Gene (wild-type unique down) | log ₂ fold change | padj | log ₁₀ padj | baseMean |
|------------------------------|------------------------------|----------|------------------------|----------|
| <i>Slitrk6</i> | -1.01 | 3.86e-02 | 1.41e+00 | 50.93 |
| <i>Dact2</i> | -1.01 | 3.85e-02 | 1.41e+00 | 51.37 |
| <i>Adamts17</i> | -1.01 | 8.55e-03 | 2.07e+00 | 44.52 |
| <i>Ccdc173</i> | -1.01 | 1.92e-02 | 1.72e+00 | 38.83 |
| <i>Ripk4</i> | -1.02 | 3.48e-02 | 1.46e+00 | 17.04 |
| <i>Gm17080</i> | -1.03 | 2.89e-02 | 1.54e+00 | 33.97 |
| <i>Gm15853</i> | -1.03 | 1.45e-02 | 1.84e+00 | 30.76 |
| <i>E330037G11Rik</i> | -1.03 | 2.69e-02 | 1.57e+00 | 19.74 |
| <i>Fgf14</i> | -1.05 | 3.86e-02 | 1.41e+00 | 70.66 |
| <i>Pdzrn4</i> | -1.05 | 2.62e-02 | 1.58e+00 | 133.42 |
| <i>Rassf9</i> | -1.05 | 4.06e-02 | 1.39e+00 | 626.03 |
| <i>Ict1os</i> | -1.06 | 2.60e-02 | 1.59e+00 | 17.01 |
| <i>Gm28523</i> | -1.06 | 1.60e-02 | 1.80e+00 | 56.84 |
| <i>Gm43075</i> | -1.06 | 4.64e-02 | 1.33e+00 | 12.46 |
| <i>Gm45809</i> | -1.06 | 4.61e-02 | 1.34e+00 | 10.64 |
| <i>Gm6581</i> | -1.06 | 4.93e-02 | 1.31e+00 | 11.72 |
| <i>BC065397</i> | -1.07 | 1.10e-02 | 1.96e+00 | 92.85 |
| <i>5033417F24Rik</i> | -1.07 | 1.92e-02 | 1.72e+00 | 23.30 |
| <i>Syt15</i> | -1.08 | 2.79e-02 | 1.55e+00 | 43.60 |
| <i>Gm20518</i> | -1.08 | 1.91e-02 | 1.72e+00 | 18.63 |

Tables 27 and 28 illustrate the top 20 uniquely upregulated or downregulated differentially expressed genes in STAT1-F77A (LAD ligation vs sham) comparison, respectively.

Table 27: Top 20 uniquely upregulated genes in STAT1-F77A following LAD ligation

| Gene (F77A unique UP) | log ₂ fold change | padj | log ₁₀ padj | baseMean |
|-----------------------|------------------------------|----------|------------------------|----------|
| <i>Tnfsf15</i> | 2.54 | 6.45e-05 | 4.19e+00 | 3.93 |
| <i>Trdc</i> | 2.47 | 2.43e-05 | 4.61e+00 | 10.88 |
| <i>Gm10800</i> | 2.39 | 6.96e-05 | 4.16e+00 | 140.52 |
| <i>Gm19610</i> | 2.29 | 1.06e-04 | 3.97e+00 | 6.86 |
| <i>Muc2</i> | 2.20 | 6.83e-05 | 4.17e+00 | 26.84 |
| <i>Khdc3</i> | 2.20 | 5.70e-04 | 3.24e+00 | 4.04 |
| <i>Gm40304</i> | 2.20 | 5.74e-04 | 3.24e+00 | 4.81 |
| <i>Gm15922</i> | 2.19 | 4.36e-04 | 3.36e+00 | 10.23 |
| <i>Gm38575</i> | 2.19 | 6.16e-04 | 3.21e+00 | 3.21 |
| <i>Gm10801</i> | 2.18 | 4.00e-04 | 3.40e+00 | 16.00 |
| <i>Gm16853</i> | 2.18 | 3.40e-04 | 3.47e+00 | 4.66 |
| <i>Slc22a20</i> | 2.17 | 6.31e-04 | 3.20e+00 | 3.17 |
| <i>5830428M24Rik</i> | 2.17 | 1.46e-04 | 3.84e+00 | 19.90 |
| <i>Chrb3</i> | 2.16 | 7.11e-04 | 3.15e+00 | 5.36 |
| <i>Gpr15</i> | 2.16 | 1.95e-04 | 3.71e+00 | 7.39 |
| <i>Irgc1</i> | 2.15 | 5.52e-04 | 3.26e+00 | 4.32 |
| <i>Gm10717</i> | 2.15 | 6.31e-04 | 3.20e+00 | 4.74 |
| <i>Gm13748</i> | 2.14 | 3.99e-04 | 3.40e+00 | 8.37 |
| <i>E230014E18Rik</i> | 2.13 | 1.18e-04 | 3.93e+00 | 16.59 |
| <i>Gm42141</i> | 2.12 | 5.47e-04 | 3.26e+00 | 9.37 |

Table 28: Top 20 uniquely downregulated genes in STAT1-F77A following LAD ligation

| Gene | log ₂ fold change | padj | log ₁₀ padj | baseMean |
|----------------------|------------------------------|----------|------------------------|----------|
| <i>Cd36</i> | -1.00 | 4.51e-03 | 2.35e+00 | 72449.49 |
| <i>Rmnd1</i> | -1.00 | 6.19e-03 | 2.21e+00 | 851.78 |
| <i>Gnb3</i> | -1.00 | 1.24e-02 | 1.91e+00 | 168.96 |
| <i>Dusp23</i> | -1.00 | 8.89e-05 | 4.05e+00 | 454.24 |
| <i>Rgs5</i> | -1.00 | 6.41e-03 | 2.19e+00 | 22728.88 |
| <i>Nipal3</i> | -1.00 | 5.40e-03 | 2.27e+00 | 1366.59 |
| <i>Zfp606</i> | -1.00 | 3.36e-03 | 2.47e+00 | 613.67 |
| <i>Myom2</i> | -1.00 | 2.38e-03 | 2.62e+00 | 69623.68 |
| <i>Macrodl</i> | -1.00 | 2.76e-04 | 3.56e+00 | 5880.34 |
| <i>D630003M21Rik</i> | -1.00 | 2.89e-02 | 1.54e+00 | 465.21 |
| <i>Uqcrfs1</i> | -1.00 | 8.79e-04 | 3.06e+00 | 28587.72 |
| <i>Fam13c</i> | -1.00 | 1.72e-02 | 1.76e+00 | 291.44 |
| <i>Cxxc4</i> | -1.00 | 2.06e-02 | 1.69e+00 | 183.16 |
| <i>Kcnb1</i> | -1.00 | 5.70e-03 | 2.24e+00 | 2996.43 |
| <i>Gm9947</i> | -1.00 | 4.03e-02 | 1.39e+00 | 10.70 |
| <i>Camk2b</i> | -1.00 | 2.07e-03 | 2.68e+00 | 573.18 |
| <i>Cd59b</i> | -1.00 | 6.76e-03 | 2.17e+00 | 54.54 |
| <i>Gm10604</i> | -1.00 | 4.75e-02 | 1.32e+00 | 12.84 |
| <i>Gm13657</i> | -1.00 | 4.62e-02 | 1.34e+00 | 17.66 |
| <i>Spa17</i> | -1.01 | 5.45e-03 | 2.26e+00 | 72.01 |

3.5 Gene set enrichment analysis identified common and unique pathways associated with myocardial infarction

To gain biological insight into the physiological role of cooperative DNA binding of STAT1 transcription factor on the global transcriptional regulation in acute myocardial infarction, we aimed to characterize the transcriptome of normal and infarcted heart tissue on day 1 from both wild-type and STAT1-F77A mice following LAD ligation. The top enriched pathways in the myocardial tissue of wild-type and STAT1-F77A animals post-MI were identified. To explore shared functions among differentially expressed genes, a common approach is to integrate the biological information, such as Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG), for classifying major biological themes of gene sets. In these analyses, we tested for coordinated differential expression over gene sets from KEGG pathways rather than changes of individual genes. Generally applicable gene-set enrichment analysis was performed using the GAGE package on the differentially expressed genes. The \log_2 -fold changes for all the genes from the DEseq2 differential expression analysis were used as an input for GAGE analysis. We assumed that perturbations over a specific pathway may propose mechanistic changes post-myocardial infarction. In addition, gene expression profiles that are genotype dependent can be identified. GAGE uses fold changes for each gene to calculate mean and standard deviation of fold changes for a specific gene set (pathway), as well as for the whole transcriptome as a background, generating *t*-test statistics and a *p*-value for each gene set including the background in the respective comparison. Finally, to integrate and visualize enriched pathways, the Pathview R package was used to render the data (Gentleman *et al.*, 2004). KEGG pathways that were identified as significantly differentially regulated were visualized using PathView, where upregulated genes are displayed in red and downregulated genes are in green and an unregulated gene in grey. The pathway is considered to be generally upregulated if the majority of genes are colored in red. A higher enrichment mean corresponds to a shifting of gene set constituents towards either end of the ranked list representing strongly upregulated or downregulated pathways.

The differentially regulated pathways demonstrate that differentially expressed genes were mainly enriched in four main categories, which are (1) immune processes, (2) signal transduction, (3) cellular processes and (4) metabolism. The enrichment scores for both immune- and extracellular matrix remodeling-related pathways were significantly higher, while metabolism-related pathways had higher negative enrichment scores in both wild-type and STAT1-F77A after myocardial infarction. The top gene set enrichment analysis (GSEA)-identified pathways for wild-type (LAD ligation vs sham) comparison with *q*-value < 0.1 are

shown in Table 29 and Figure 13, and similarly for STAT1-F77A (LAD ligation vs sham) comparison in Table 30 and Figure 14, while Figure 15 lists both genotypes.



Figure 13: List of KEGG differentially regulated pathways in wild-type mice post-MI.
Table 29: List of KEGG differentially regulated pathways following LAD ligation in wild-type mice.

| | KEGG ID | KEGG pathway (wild-type MI vs sham) | mean | p value | q value | size |
|---------------------|----------|--|------|----------|----------|------|
| Immune Responses | mmu04060 | Cytokine-cytokine receptor interaction | 9.1 | 4.66e-18 | 1.03e-15 | 204 |
| | mmu04062 | Chemokine signalling pathway | 5.2 | 1.70e-07 | 4.16e-06 | 167 |
| | mmu04514 | Cell adhesion molecules (CAMs) | 4.5 | 4.80e-06 | 8.80e-05 | 117 |
| | mmu04810 | Regulation of actin cytoskeleton | 4.3 | 9.56e-06 | 1.40e-04 | 195 |
| | mmu04670 | Leukocyte transendothelial migration | 4.3 | 1.32e-05 | 1.82e-04 | 95 |
| | mmu04510 | Focal adhesion | 2.3 | 1.03e-02 | 5.29e-02 | 192 |
| | mmu04611 | Platelet activation | 2.1 | 1.66e-02 | 8.10e-02 | 116 |
| | mmu04520 | Adherens junction | 2.1 | 1.91e-02 | 9.14e-02 | 70 |
| | mmu04620 | Toll-like receptor signalling pathway | 4.7 | 2.93e-06 | 5.86e-05 | 85 |
| | mmu04666 | FcγR-mediated phagocytosis | 3.2 | 8.39e-04 | 6.84e-03 | 85 |
| | mmu04640 | Hematopoietic cell lineage | 6.7 | 2.00e-10 | 2.20e-08 | 84 |
| | mmu04659 | Th17 cell differentiation | 3.3 | 6.32e-04 | 5.56e-03 | 93 |
| | mmu04657 | IL-17 signalling pathway | 6.1 | 4.73e-09 | 2.60e-07 | 78 |
| | mmu04660 | T cell receptor signalling pathway | 3.3 | 5.44e-04 | 5.44e-03 | 93 |
| | mmu04658 | Th1 and Th2 cell differentiation | 3 | 1.69e-03 | 1.32e-02 | 79 |
| | mmu04662 | B cell receptor signalling pathway | 3.3 | 5.96e-04 | 5.56e-03 | 69 |
| | mmu04650 | Natural killer cell mediated cytotoxicity | 4.8 | 1.93e-06 | 4.25e-05 | 96 |
| | mmu04672 | Intestinal immune network for IgA production | 3.4 | 6.20e-04 | 5.56e-03 | 34 |
| | mmu04610 | Complement and coagulation cascades | 3.5 | 3.91e-04 | 4.09e-03 | 59 |
| | mmu04664 | Fc epsilon RI signalling pathway | 2.2 | 1.54e-02 | 7.69e-02 | 61 |
| | mmu04612 | Antigen processing and presentation | 4.6 | 6.35e-06 | 1.07e-04 | 62 |
| Signal Transduction | mmu04151 | PI3K-Akt signalling pathway | 3.2 | 7.66e-04 | 6.48e-03 | 306 |
| | mmu04630 | Jak-STAT signalling pathway | 3.6 | 1.89e-04 | 2.18e-03 | 124 |
| | mmu04668 | TNF signalling pathway | 5.6 | 4.48e-08 | 1.62e-06 | 108 |
| | mmu04064 | NF-κB signalling pathway | 5.6 | 5.15e-08 | 1.62e-06 | 89 |
| | mmu04621 | NOD-like receptor signalling pathway | 5.4 | 7.44e-08 | 2.04e-06 | 146 |
| | mmu04010 | MAPK signalling pathway | 4.3 | 8.88e-06 | 1.40e-04 | 272 |
| | mmu04380 | Osteoclast differentiation | 6.4 | 4.91e-10 | 3.60e-08 | 122 |
| | mmu04066 | HIF-1 signalling pathway | 2.9 | 2.25e-03 | 1.60e-02 | 95 |
| | mmu04350 | TGF-β signalling pathway | 2.7 | 3.95e-03 | 2.63e-02 | 73 |
| | mmu04015 | Rap1 signalling pathway | 2.6 | 4.93e-03 | 3.03e-02 | 188 |
| | mmu04622 | RIG-I-like receptor signalling pathway | 2.6 | 4.95e-03 | 3.03e-02 | 52 |
| | mmu04390 | Hippo signalling pathway | 2.9 | 1.98e-03 | 1.45e-02 | 138 |
| | mmu04115 | p53 signalling pathway | 2.4 | 8.59e-03 | 4.61e-02 | 69 |
| | mmu04623 | Cytosolic DNA-sensing pathway | 4 | 7.89e-05 | 1.02e-03 | 46 |
| Cellular Processes | mmu04144 | Endocytosis | 2.7 | 3.74e-03 | 2.57e-02 | 240 |
| | mmu04110 | Cell cycle | 2.6 | 4.95e-03 | 3.03e-02 | 119 |
| | mmu04218 | Cellular senescence | 2.9 | 1.87e-03 | 1.42e-02 | 155 |
| | mmu04210 | Apoptosis | 3.8 | 1.02e-04 | 1.24e-03 | 131 |
| | mmu04217 | Necroptosis | 2.4 | 8.96e-03 | 4.69e-02 | 121 |
| | mmu03010 | Ribosome | 2.5 | 6.99e-03 | 3.94e-02 | 130 |
| | mmu04145 | Phagosome | 5.9 | 5.96e-09 | 2.62e-07 | 137 |
| | mmu04146 | Peroxisome | -4.5 | 8.56e-06 | 4.71e-04 | 78 |
| | mmu03040 | Spliceosome | 2.5 | 6.16e-03 | 3.63e-02 | 127 |
| | mmu04141 | Protein processing in endoplasmic reticulum | 2.5 | 6.26e-03 | 3.63e-02 | 155 |
| | mmu04723 | Retrograde endocannabinoid signalling | -3.2 | 6.97e-04 | 1.92e-02 | 122 |
| | mmu03008 | Ribosome biogenesis in eukaryotes | 3.6 | 2.05e-04 | 2.26e-03 | 72 |
| Metabolism | mmu00190 | Oxidative phosphorylation | -6.3 | 1.28e-09 | 2.82e-07 | 116 |
| | mmu00020 | Citrate cycle (TCA cycle) | -3.9 | 1.84e-04 | 6.74e-03 | 30 |
| | mmu01212 | Fatty acid metabolism | -2.8 | 3.51e-03 | 7.72e-02 | 49 |
| | mmu00071 | Fatty acid degradation | -4 | 7.69e-05 | 3.39e-03 | 37 |
| | mmu00640 | Propanoate metabolism | -5.1 | 4.08e-06 | 4.49e-04 | 29 |
| | mmu00310 | Lysine degradation | -2.8 | 3.24e-03 | 7.72e-02 | 56 |
| | mmu00380 | Tryptophan metabolism | -2.7 | 4.39e-03 | 8.79e-02 | 33 |
| | mmu00620 | Pyruvate metabolism | -3.4 | 6.88e-04 | 1.92e-02 | 33 |
| | mmu00280 | Valine, leucine and isoleucine degradation | -4.6 | 7.44e-06 | 4.71e-04 | 47 |
| | mmu00532 | Glycosaminoglycan biosynthesis | 2.6 | 7.39e-03 | 4.06e-02 | 20 |



Figure 14: List of KEGG differentially regulated pathways following LAD ligation in transgenic mice expressing STAT1-F77A.

Table 30: List of KEGG differentially regulated pathways following LAD ligation in STAT1-F77A mice

| | KEGG ID | KEGG pathway (STAT1-F77A MI vs sham) | mean | p value | q value | size |
|---------------------|----------|--|------|----------|----------|------|
| Immune Responses | mmu04060 | Cytokine-cytokine receptor interaction | 8.7 | 6.39e-17 | 1.41e-14 | 204 |
| | mmu04062 | Chemokine signalling pathway | 5.7 | 1.53e-08 | 5.60e-07 | 167 |
| | mmu04514 | Cell adhesion molecules (CAMs) | 5.1 | 3.87e-07 | 8.51e-06 | 117 |
| | mmu04810 | Regulation of actin cytoskeleton | 4.2 | 2.03e-05 | 3.19e-04 | 195 |
| | mmu04670 | Leukocyte transendothelial migration | 4.1 | 2.71e-05 | 3.97e-04 | 95 |
| | mmu04510 | Focal adhesion | 2.5 | 6.26e-03 | 3.53e-02 | 192 |
| | mmu04611 | Platelet activation | 2.2 | 1.40e-02 | 7.18e-02 | 116 |
| | mmu04520 | Adherens junction | 2.2 | 1.48e-02 | 7.40e-02 | 70 |
| | mmu04620 | Toll-like receptor signalling pathway | 4.6 | 4.14e-06 | 7.60e-05 | 85 |
| | mmu04666 | FcγR-mediated phagocytosis | 3.2 | 7.22e-04 | 6.62e-03 | 85 |
| | mmu04640 | Hematopoietic cell lineage | 6.2 | 2.89e-09 | 1.59e-07 | 84 |
| | mmu04659 | Th17 cell differentiation | 3.3 | 5.96e-04 | 5.70e-03 | 93 |
| | mmu04657 | IL-17 signalling pathway | 6.1 | 5.81e-09 | 2.56e-07 | 78 |
| | mmu04660 | T cell receptor signalling pathway | 3.4 | 3.77e-04 | 3.77e-03 | 93 |
| | mmu04658 | Th1 and Th2 cell differentiation | 3 | 1.43e-03 | 1.17e-02 | 79 |
| | mmu04662 | B cell receptor signalling pathway | 3 | 1.70e-03 | 1.29e-02 | 69 |
| | mmu04650 | Natural killer cell mediated cytotoxicity | 4.4 | 8.80e-06 | 1.49e-04 | 96 |
| | mmu04672 | Intestinal immune network for IgA production | 3.6 | 2.95e-04 | 3.09e-03 | 34 |
| | mmu04610 | Complement and coagulation cascades | 3 | 1.69e-03 | 1.29e-02 | 59 |
| | mmu04612 | Antigen processing and presentation | 4.9 | 1.83e-06 | 3.66e-05 | 62 |
| Signal Transduction | mmu04151 | PI3K-Akt signalling pathway | 3.1 | 1.02e-03 | 8.96e-03 | 306 |
| | mmu04630 | Jak-STAT signalling pathway | 3.5 | 2.76e-04 | 3.06e-03 | 124 |
| | mmu04668 | TNF signalling pathway | 5.5 | 5.72e-08 | 1.40e-06 | 108 |
| | mmu04064 | NF-κB signalling pathway | 5.6 | 3.62e-08 | 1.14e-06 | 89 |
| | mmu04621 | NOD-like receptor signalling pathway | 5.5 | 4.48e-08 | 1.23e-06 | 146 |
| | mmu04010 | MAPK signalling pathway | 3.9 | 4.86e-05 | 6.69e-04 | 272 |
| | mmu04380 | Osteoclast differentiation | 6.3 | 8.57e-10 | 9.43e-08 | 122 |
| | mmu04066 | HIF-1 signalling pathway | 2.7 | 4.22e-03 | 2.66e-02 | 95 |
| | mmu04350 | TGF-β signalling pathway | 2.6 | 5.50e-03 | 3.36e-02 | 73 |
| | mmu04015 | Rap1 signalling pathway | 2.8 | 2.32e-03 | 1.60e-02 | 188 |
| | mmu04622 | RIG-I-like receptor signalling pathway | 2.8 | 3.34e-03 | 2.22e-02 | 52 |
| | mmu04390 | Hippo signalling pathway | 2.3 | 1.14e-02 | 6.13e-02 | 138 |
| | mmu04115 | p53 signalling pathway | 2.5 | 7.40e-03 | 4.07e-02 | 69 |
| | mmu04623 | Cytosolic DNA-sensing pathway | 3.9 | 1.05e-04 | 1.28e-03 | 46 |
| | mmu03013 | RNA transport | 2.3 | 1.19e-02 | 6.25e-02 | 150 |
| Cellular Processes | mmu04144 | Endocytosis | 2.9 | 2.05e-03 | 1.50e-02 | 240 |
| | mmu04110 | Cell cycle | 2.9 | 2.24e-03 | 1.59e-02 | 119 |
| | mmu04260 | Cardiac muscle contraction | -3.3 | 5.57e-04 | 1.36e-02 | 67 |
| | mmu04218 | Cellular senescence | 3 | 1.40e-03 | 1.17e-02 | 155 |
| | mmu04210 | Apoptosis | 3.5 | 2.79e-04 | 3.06e-03 | 131 |
| | mmu04217 | Necroptosis | 2.2 | 1.52e-02 | 7.44e-02 | 121 |
| | mmu04145 | Phagosome | 6.1 | 2.03e-09 | 1.49e-07 | 137 |
| | mmu04146 | Peroxisome | -4.5 | 5.53e-06 | 2.44e-04 | 78 |
| | mmu03040 | Spliceosome | 2.5 | 5.88e-03 | 3.40e-02 | 127 |
| | mmu04141 | Protein processing in endoplasmic reticulum | 2.5 | 5.83e-03 | 3.40e-02 | 155 |
| | mmu04723 | Retrograde endocannabinoid signalling | -3.6 | 2.02e-04 | 5.56e-03 | 122 |
| | mmu03008 | Ribosome biogenesis in eukaryotes | 3.9 | 9.28e-05 | 1.20e-03 | 72 |
| | mmu00190 | Oxidative phosphorylation | -8.1 | 3.66e-14 | 8.06e-12 | 116 |
| | mmu00020 | Citrate cycle (TCA cycle) | -5.1 | 3.38e-06 | 1.86e-04 | 30 |
| | mmu01212 | Fatty acid metabolism | -2.8 | 3.56e-03 | 6.02e-02 | 49 |
| Metabolism | mmu00071 | Fatty acid degradation | -4.1 | 5.31e-05 | 1.67e-03 | 37 |
| | mmu00640 | Propanoate metabolism | -5.8 | 3.59e-07 | 2.63e-05 | 29 |
| | mmu00380 | Tryptophan metabolism | -3 | 2.01e-03 | 4.01e-02 | 33 |
| | mmu00620 | Pyruvate metabolism | -4.2 | 4.22e-05 | 1.55e-03 | 33 |
| | mmu00280 | Valine, leucine and isoleucine degradation | -5.4 | 2.58e-07 | 2.63e-05 | 47 |
| | mmu00532 | Glycosaminoglycan biosynthesis | 2.8 | 4.19e-03 | 2.66e-02 | 20 |
| | mmu00650 | Butanoate metabolism | -2.8 | 4.39e-03 | 6.90e-02 | 19 |
| | mmu00630 | Glyoxylate and dicarboxylate metabolism | -2.7 | 5.15e-03 | 7.55e-02 | 28 |
| | mmu01200 | Carbon metabolism | -2.9 | 1.98e-03 | 4.01e-02 | 106 |
| | mmu00982 | Drug metabolism - cytochrome P450 | -2.8 | 3.12e-03 | 5.72e-02 | 37 |
| | mmu03320 | PPAR signalling pathway | -2.6 | 5.49e-03 | 7.55e-02 | 61 |



Figure 15: KEGG-regulated pathways following LAD ligation in the two STAT1 genotypes.

3.6 Identification of immune-related pathways differentially regulated after myocardial infarction

In this study, KEGG pathways that were classified as immune processes significantly enriched post-myocardial infarction in both wild-type and STAT1-F77A mice were the following: cytokine-cytokine receptor interaction (Figure 17), chemokine signalling pathway (Figure 18), cell adhesion molecules (CAMs) (Figure 19), regulation of actin cytoskeleton (Figure 20), leukocyte transendothelial migration (Figure 21), focal adhesion, platelet activation, adherens junction, Toll-like receptor signalling pathway, Fc γ R-mediated phagocytosis, hematopoietic cell lineage, Th17 cell differentiation, IL-17 signalling pathway, T cell receptor signalling pathway, Th1 and Th2 cell differentiation, B cell receptor signalling pathway, natural killer cell-mediated cytotoxicity, intestinal immune network for IgA production, complement and coagulation cascades, antigen processing and presentation.

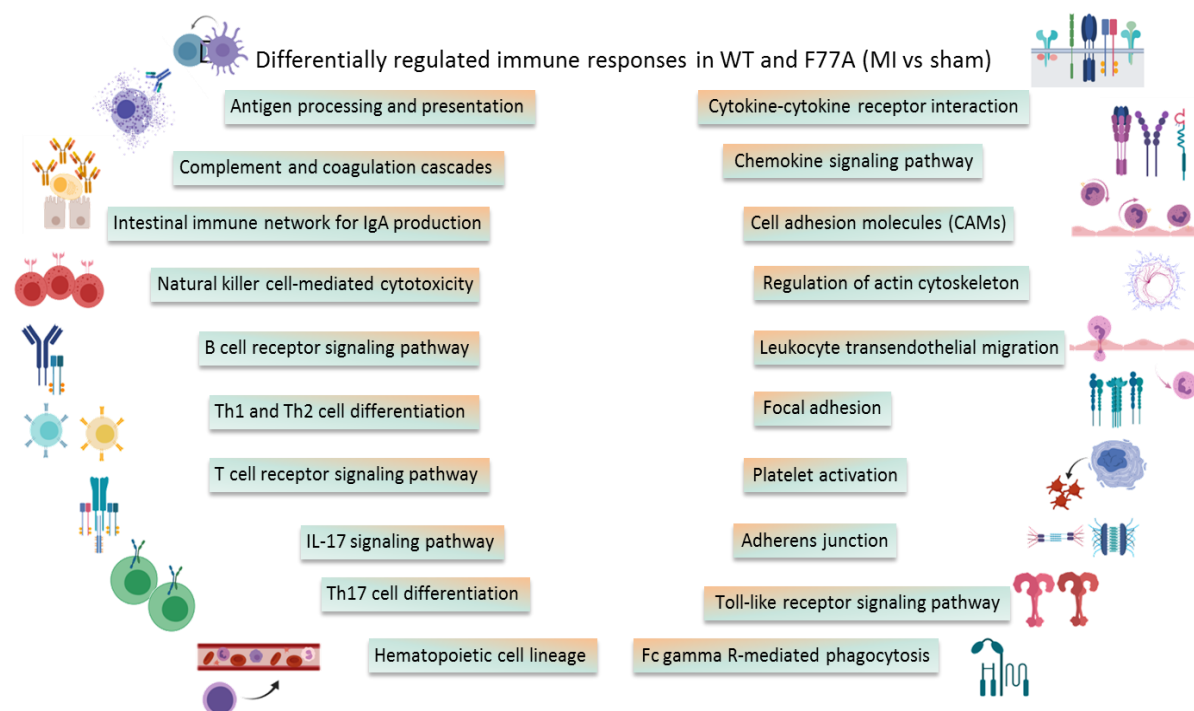


Figure 16: List of KEGG immune responses that were differentially regulated in both STAT1-F77A and wild-type mice.

WT

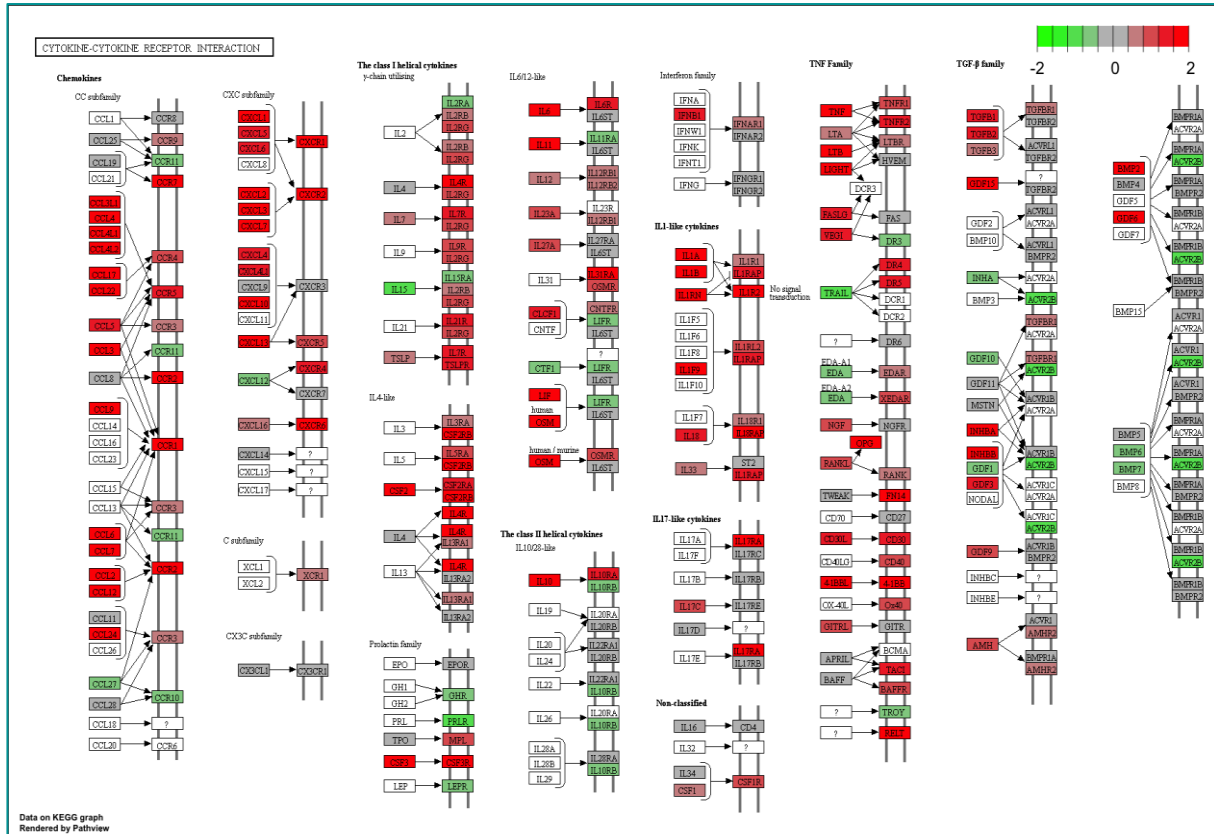
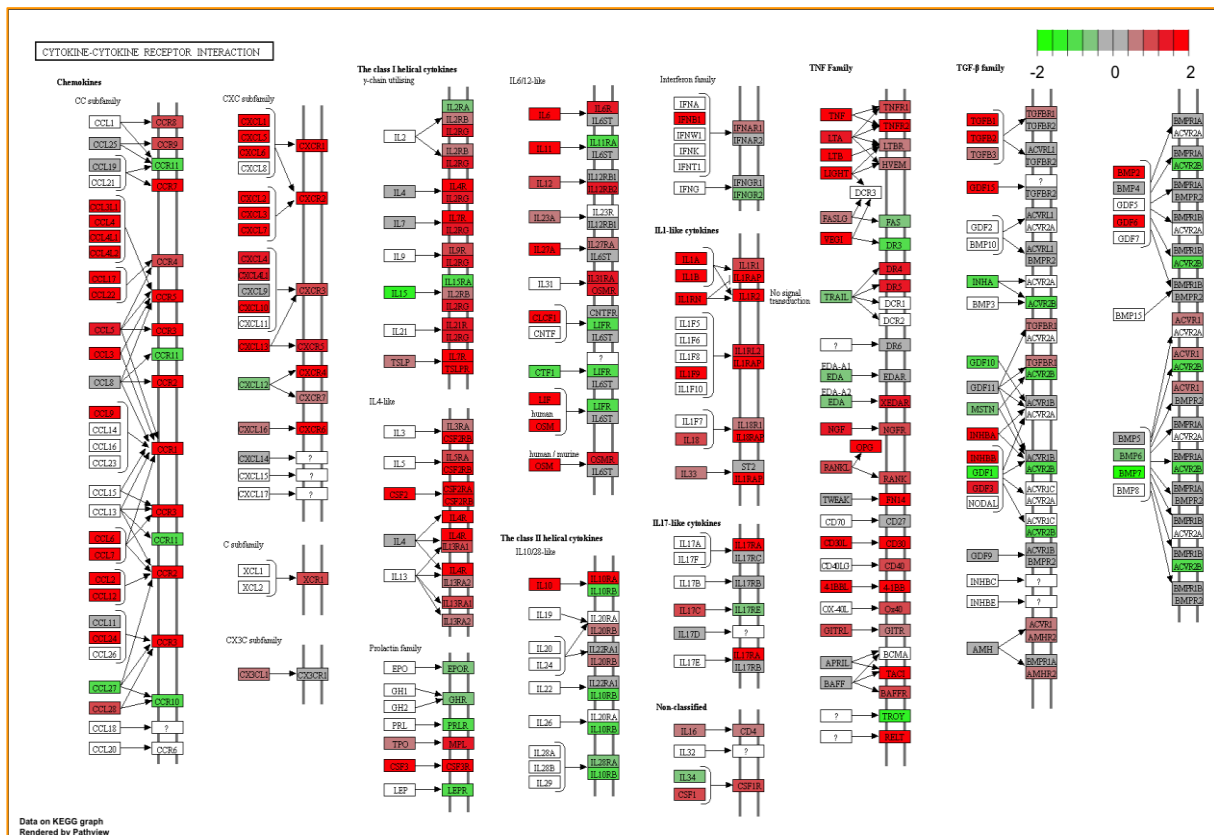
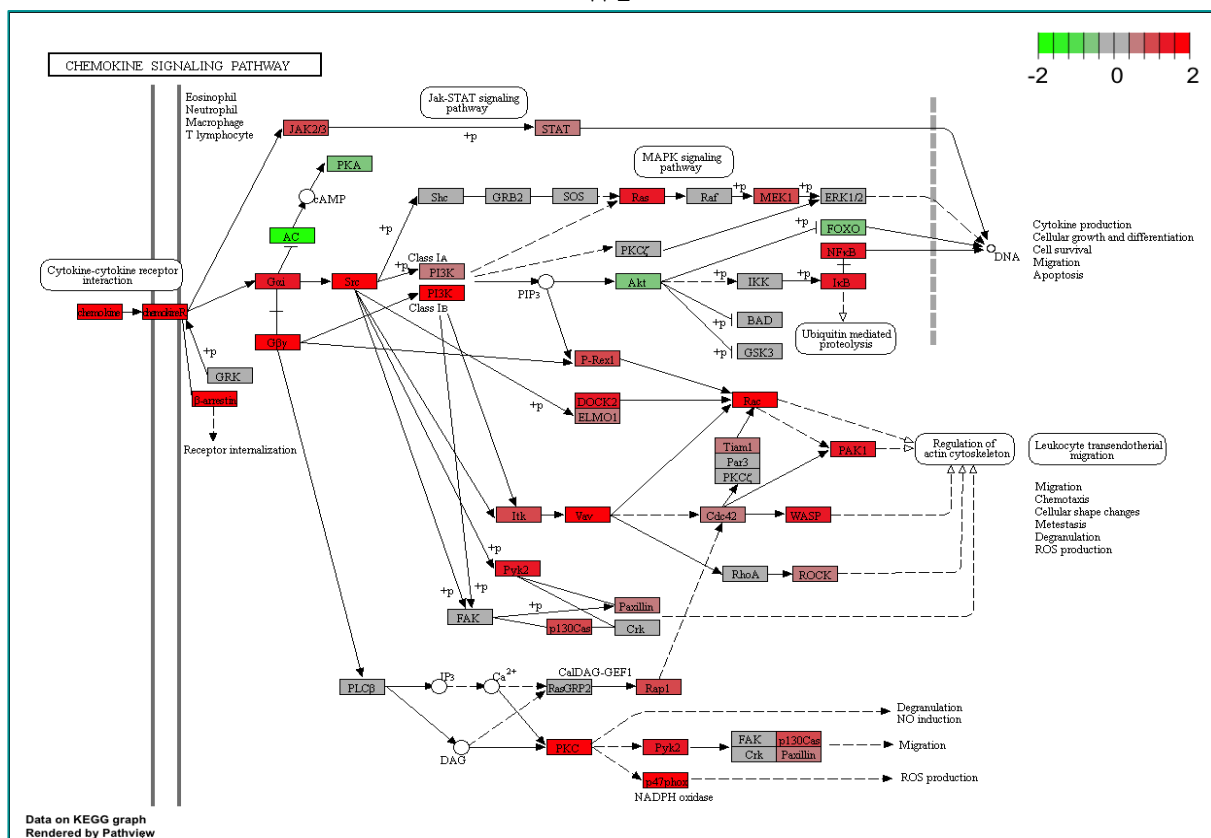
**STAT1-F77A**

Figure 17: Pathview of the “cytokine-cytokine receptor interaction” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

WT



STAT1-F77A

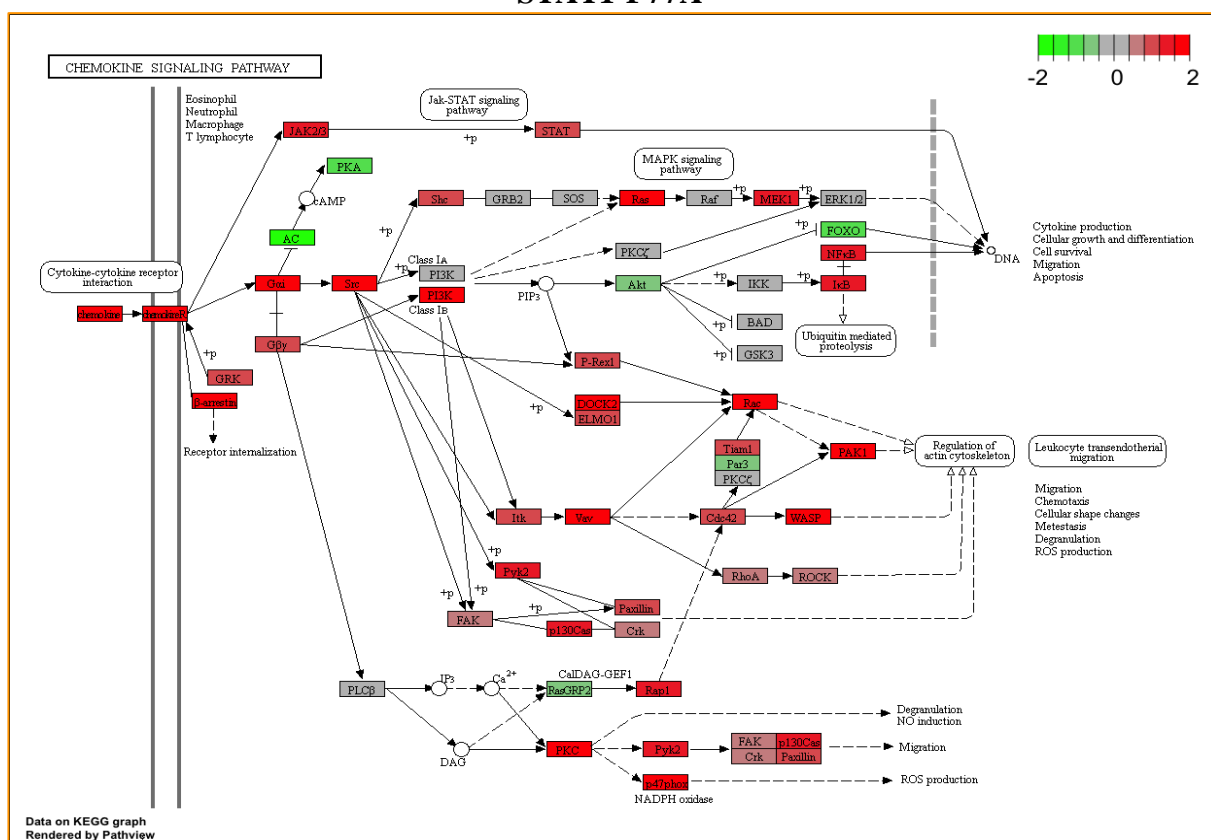


Figure 18: Pathview of the “chemokine signalling” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

WT

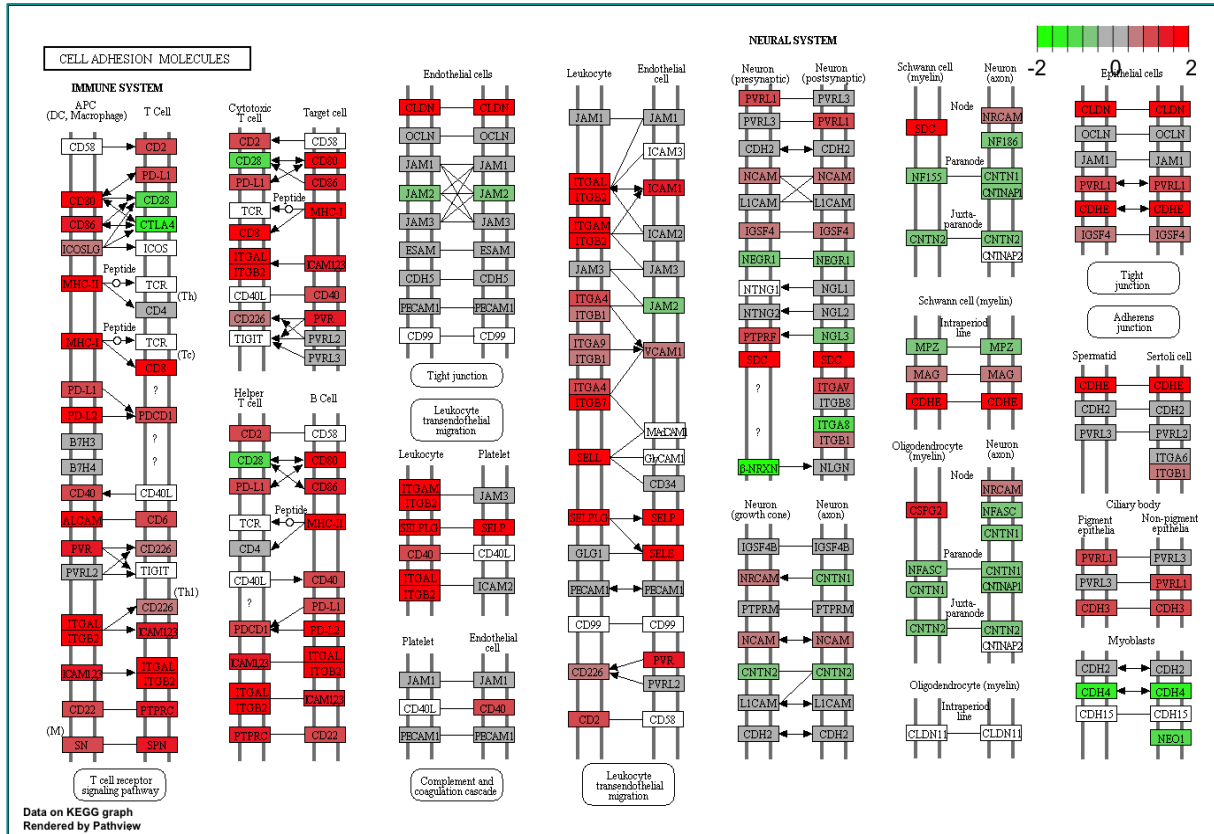
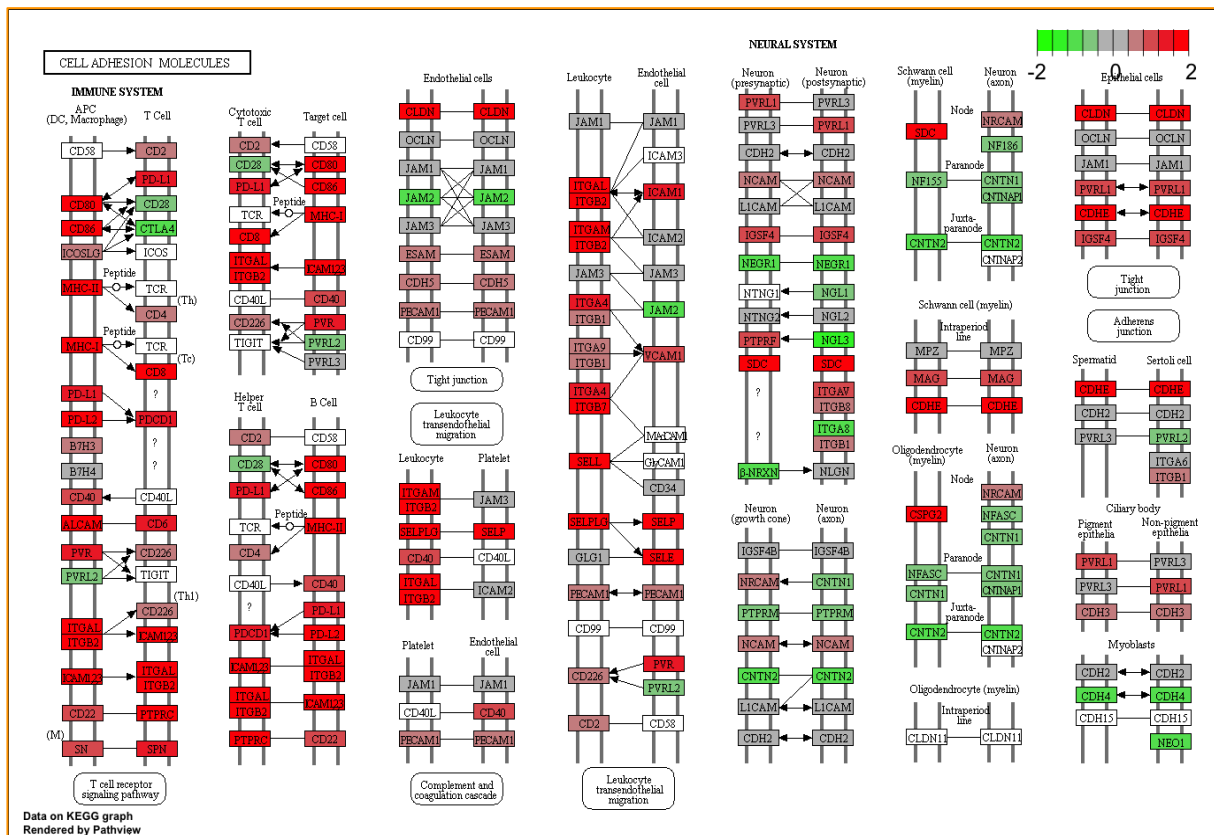
**STAT1-F77A**

Figure 19: Pathview of the “cell adhesion molecules (CAMs)” KEGG pathway in the two STAT1 genotypes following myocardial infarction.

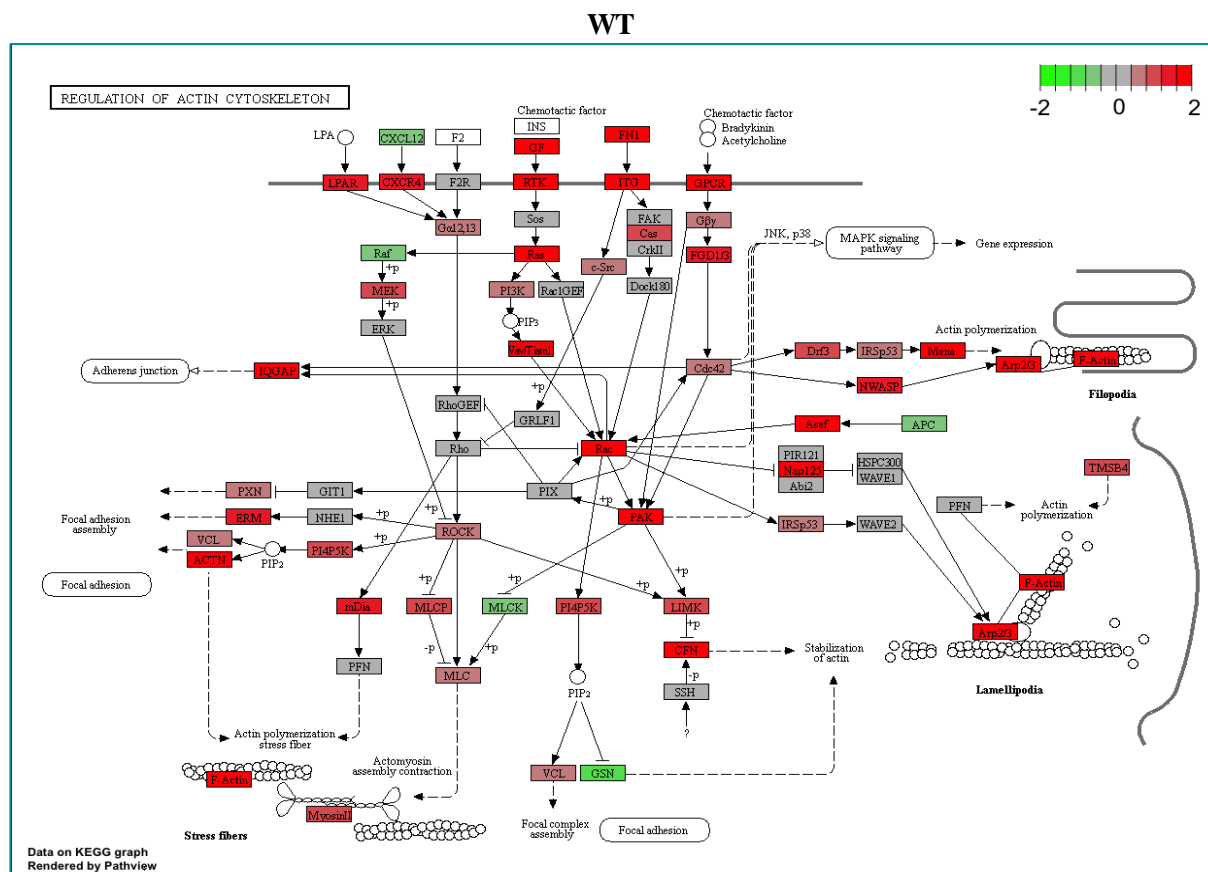
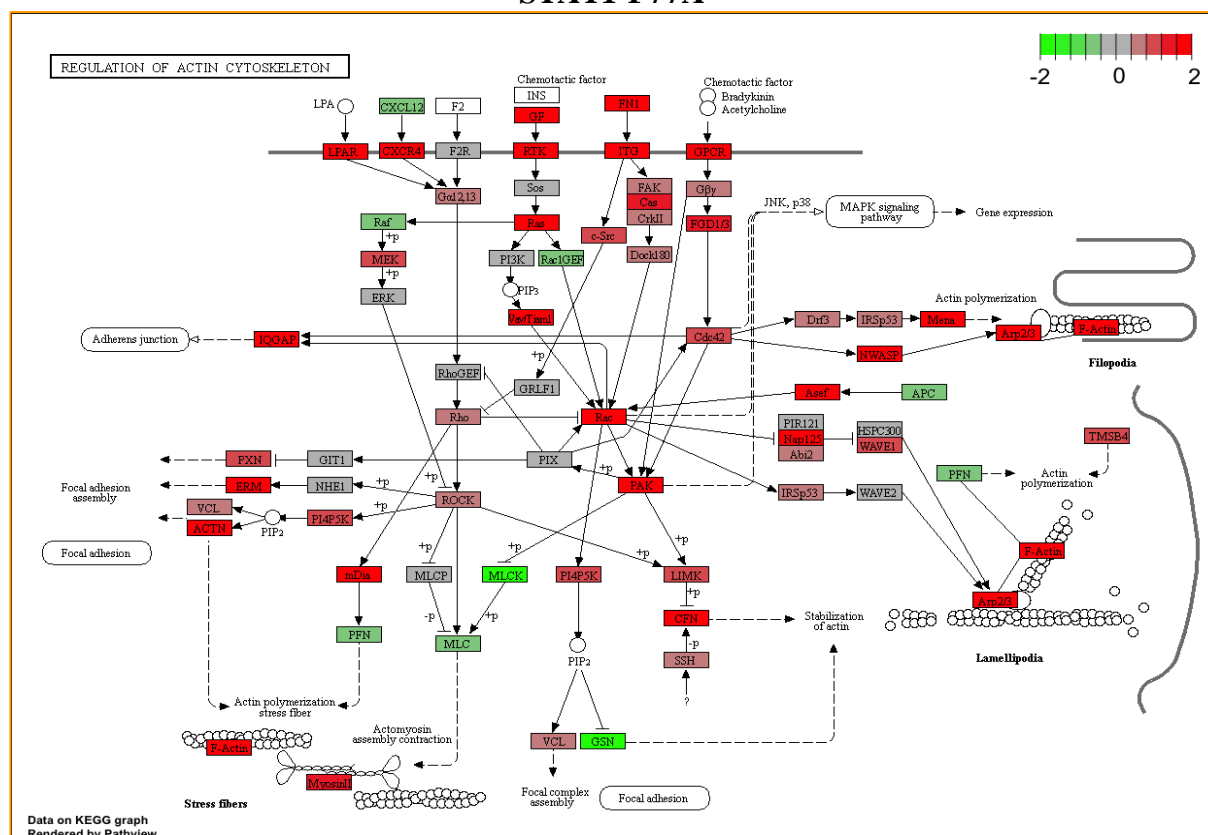
**STAT1-F77A**

Figure 20: Pathview of the “regulation of actin cytoskeleton” KEGG pathway in mice with with wild-type or mutant *Stat* allele following LAD ligation.

[illegible][illegible]

3.7 Identification of signal pathways and cellular processes differentially regulated after LAD ligation

Since signal transduction pathways transmit extracellular stimuli inside the cell thereby regulating the expression of genes with important physiological functions, we performed differential expression analysis to identify the most significantly enriched signal pathways. Signal transduction pathways that were associated with myocardial infarction and were genotype-independent are the following: PI3K-Akt signalling (Figure 23), JAK-STAT signalling (Figure 24), TNF signalling (Figure 25), NF- κ B signalling (Figure 26), NOD-like receptor signalling (Figure 27), MAPK signalling, osteoclast differentiation, HIF-1 signalling, TGF- β signalling, Rap1 signalling, RIG-I-like receptor signalling, Hippo signalling, p53 signalling, and cytosolic DNA-sensing pathway.

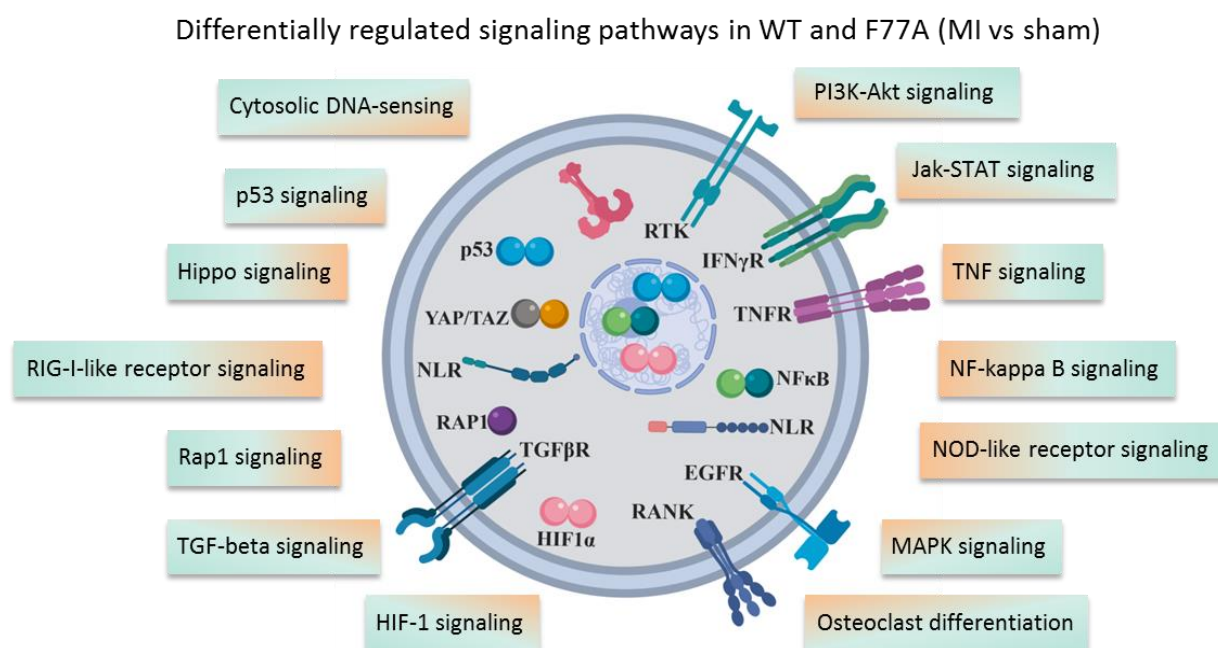
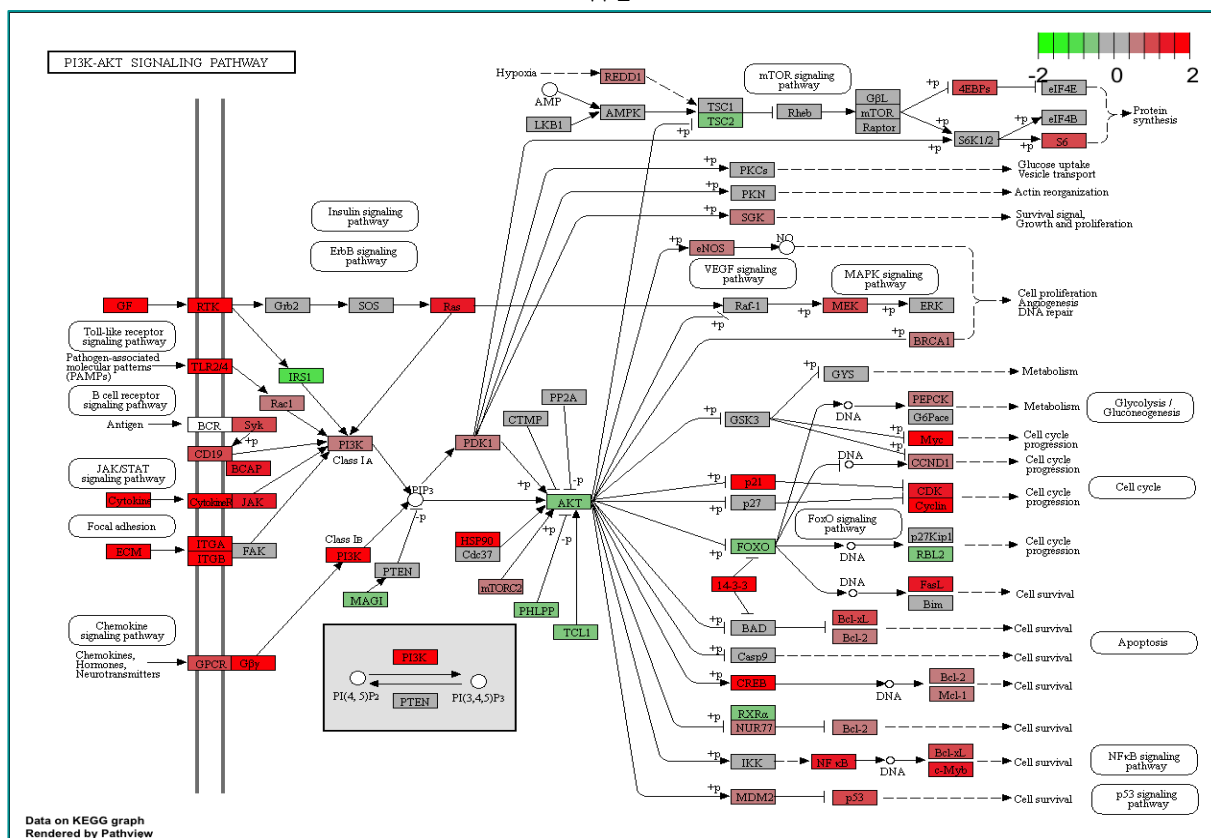


Figure 22: List of KEGG immune responses and signal transduction pathways that are differentially regulated in between STAT1-F77A and wild-type mice.

WT



STAT1-F77A

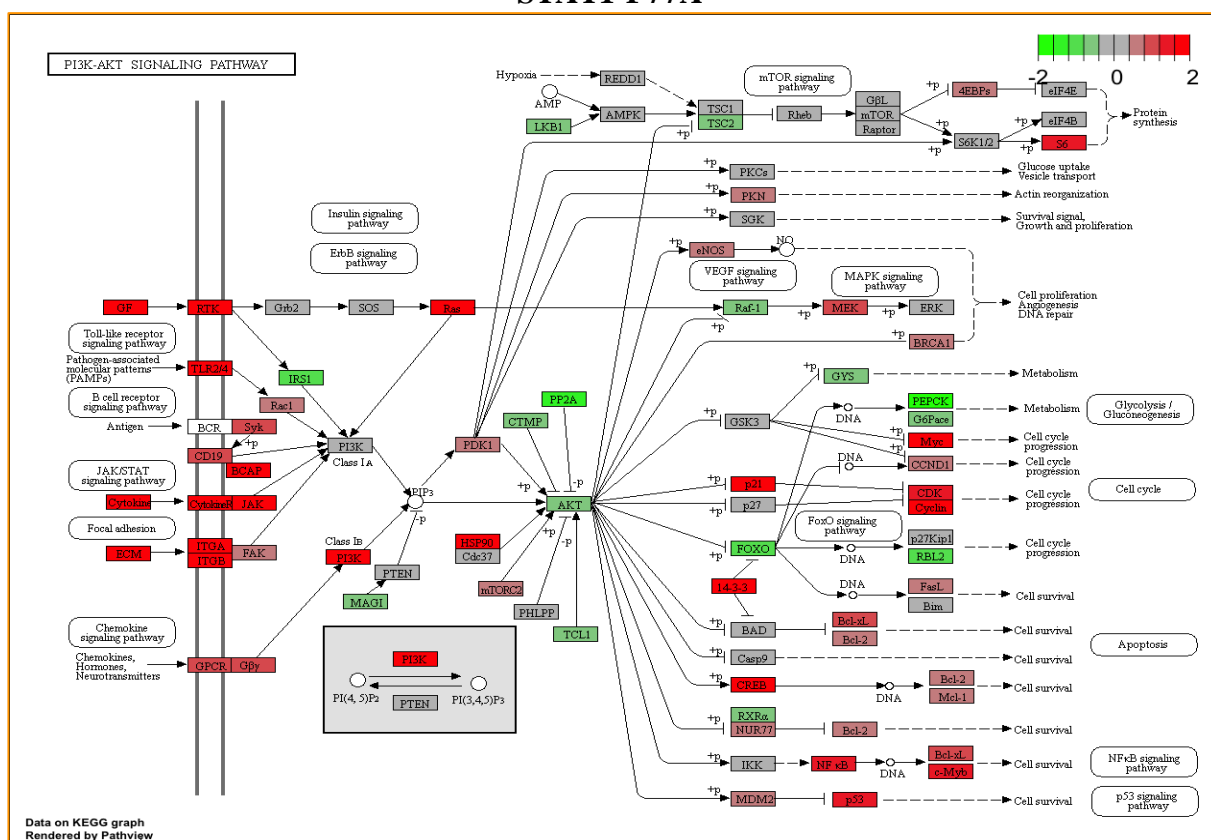


Figure 23: Pathview of the “PI3K-Akt signalling” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

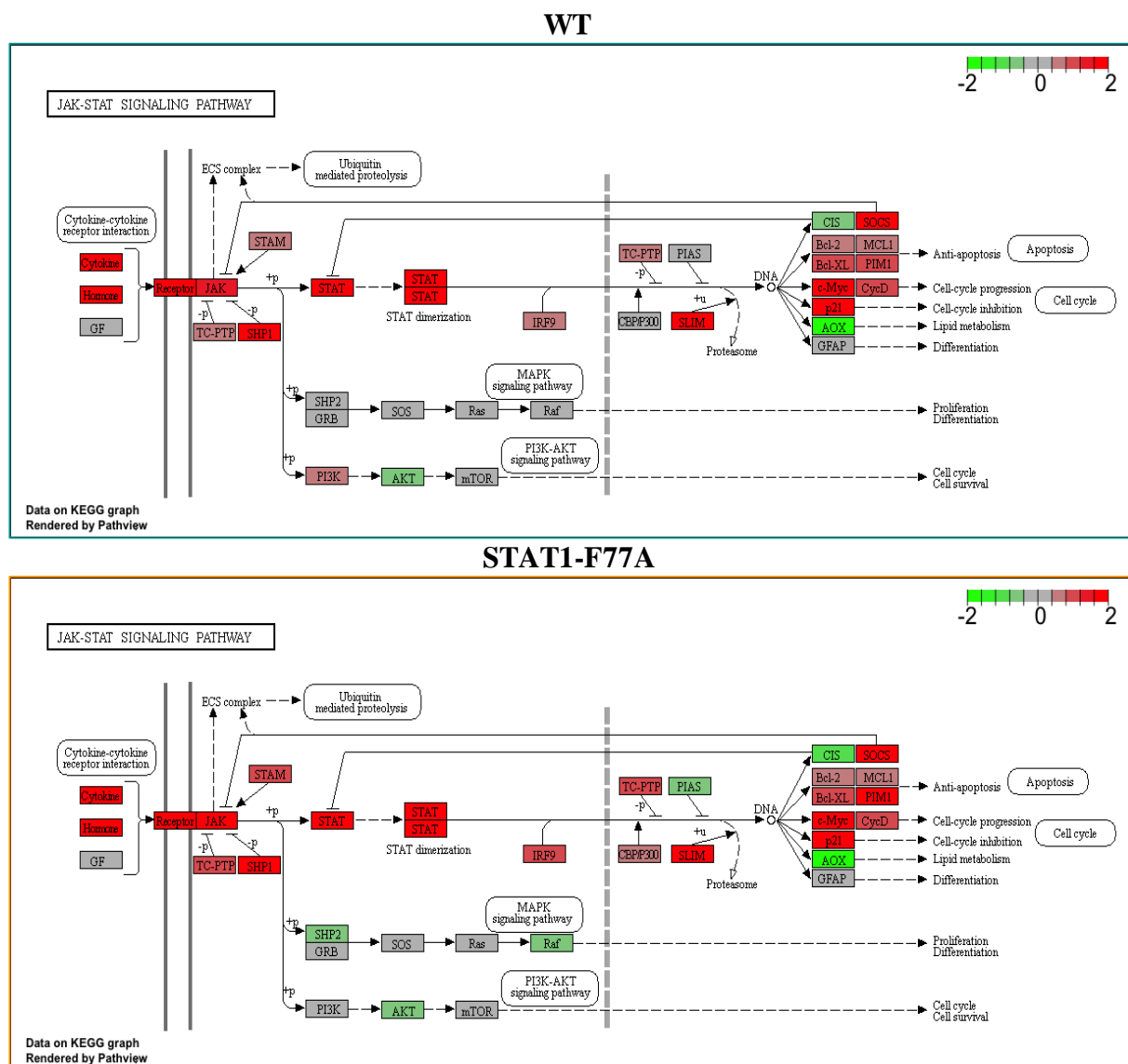


Figure 24: Pathview of the “Jak-STAT signalling” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

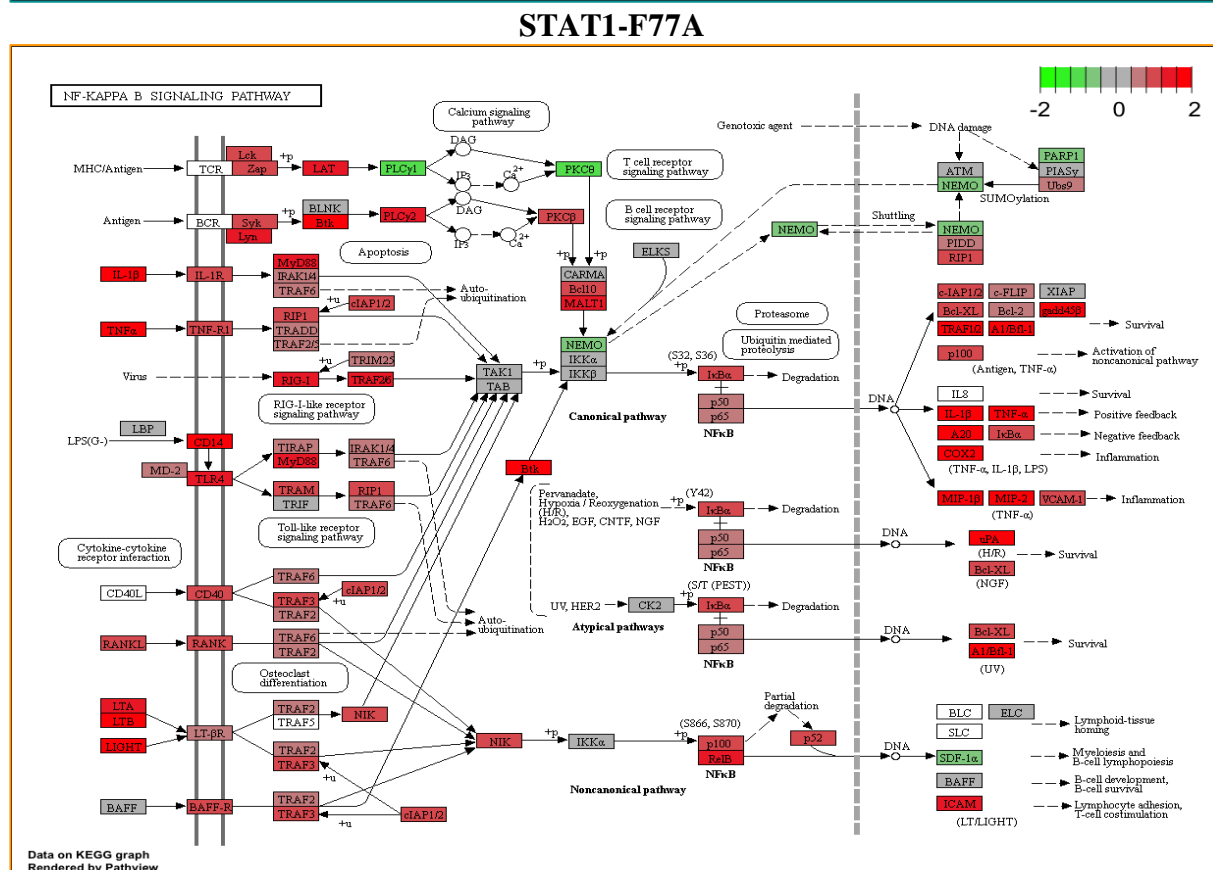
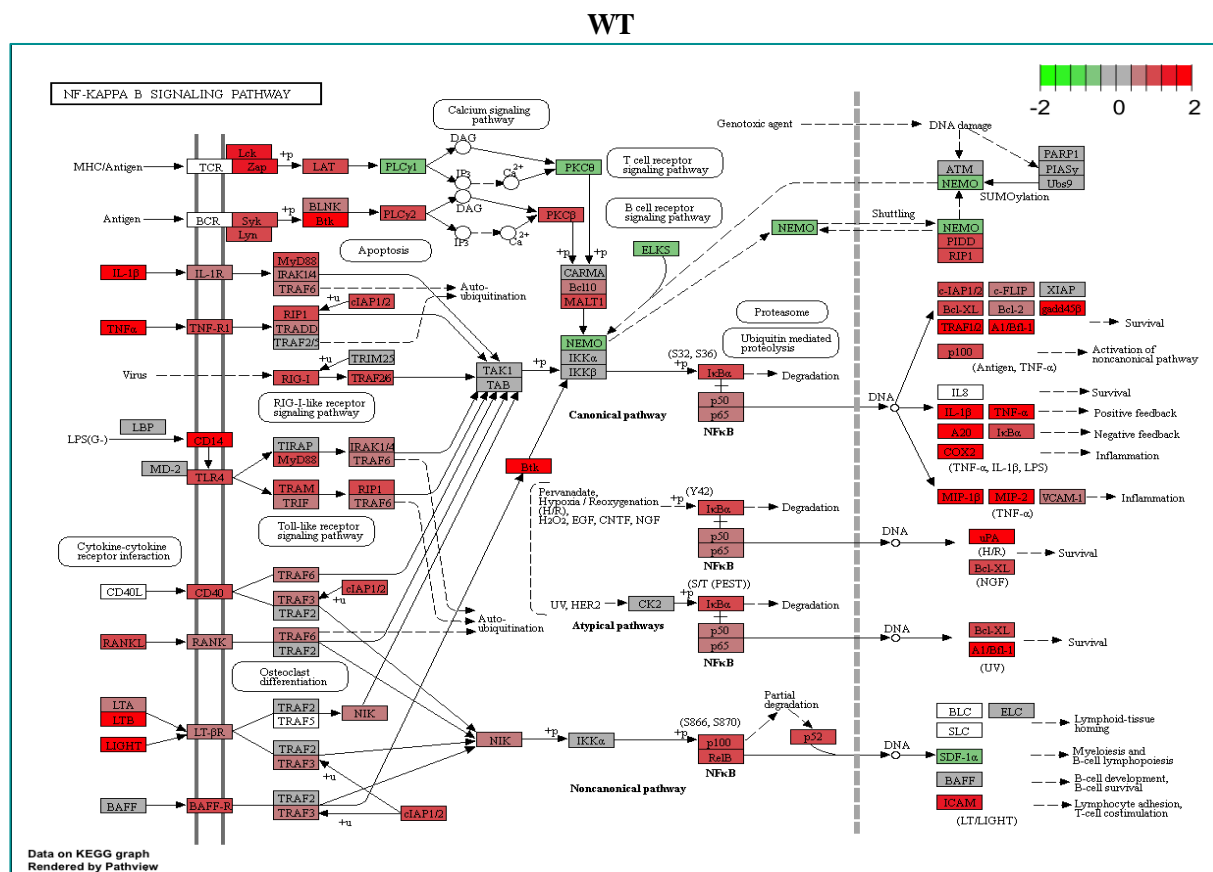
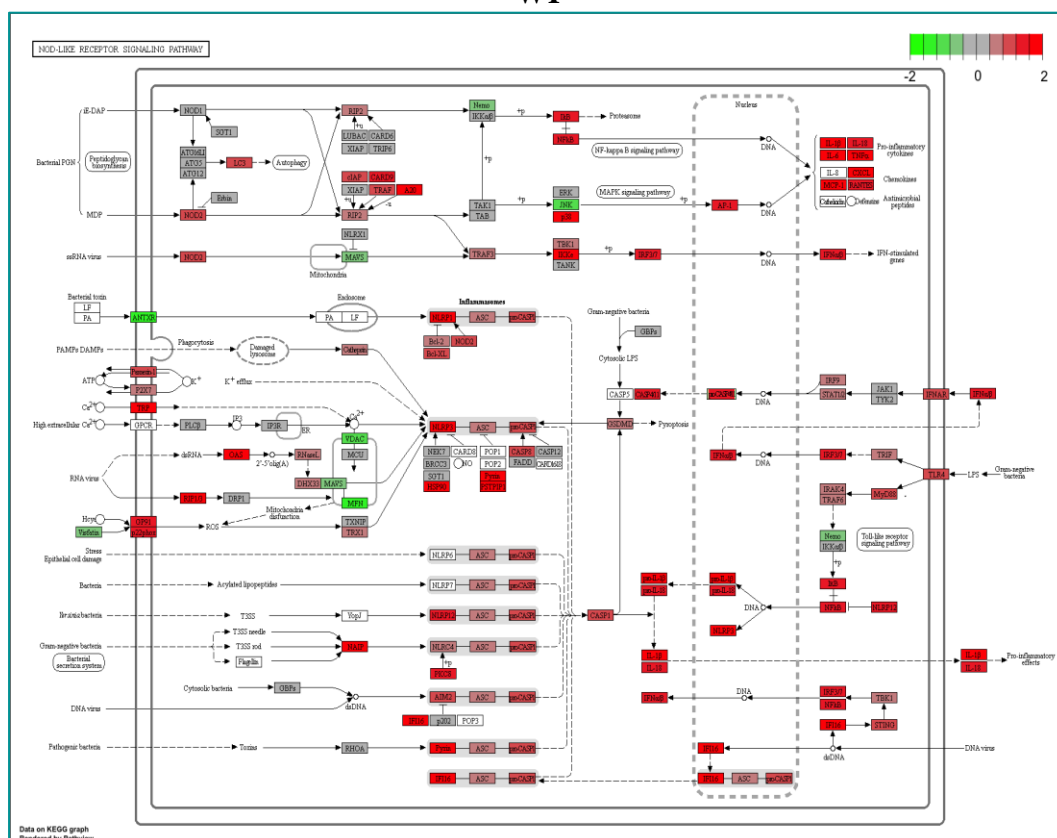


Figure 26: Differential expression of components in the “NF-κB signalling” KEGG pathway in the two *Stat1* genotypes following LAD ligation.

WT



STAT1-F77A

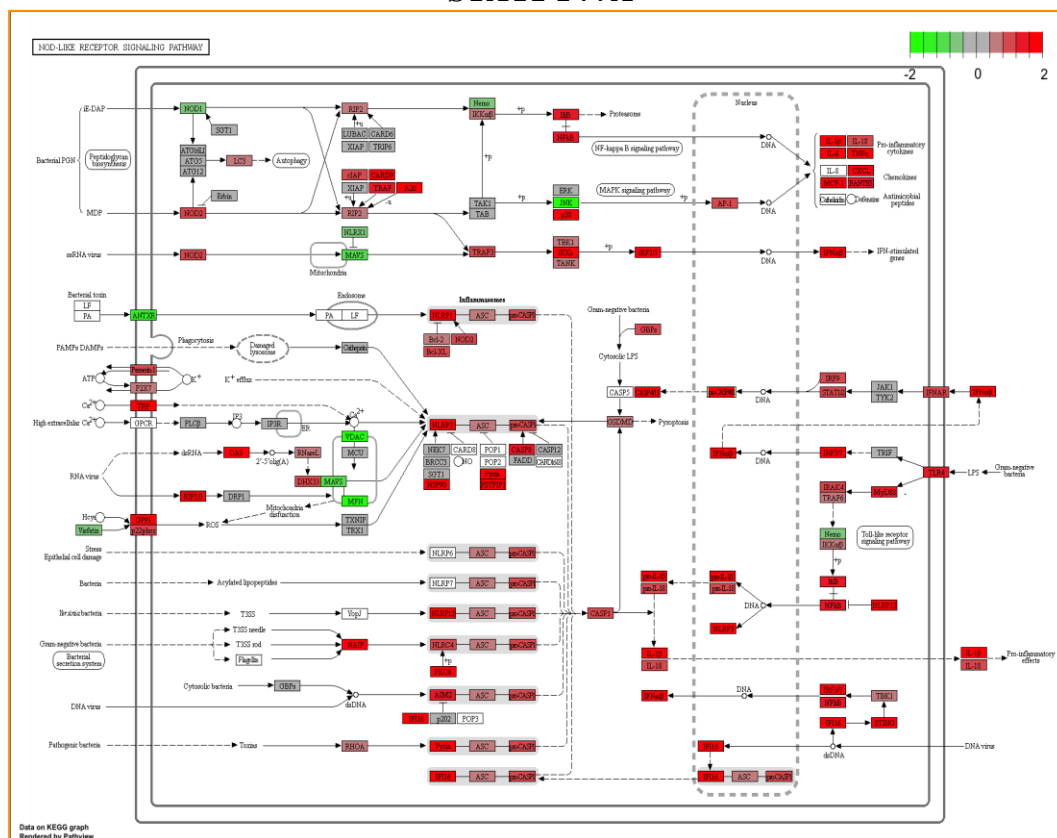


Figure 27: Pathview of the “NOD-like receptor signalling” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

Additionally, the top significantly perturbed cellular processes associated with MI which are genotype-independent include: endocytosis (Figure 29), cell cycle (Figure 30), cellular senescence (Figure 31), apoptosis (Figure 32), necroptosis (Figure 33), phagosome, peroxisome, spliceosome, protein processing in endoplasmic reticulum, and retrograde endocannabinoid signalling and ribosome biogenesis in eukaryotes. Interestingly, we found cardiac muscle contraction to be uniquely differentially regulated in STAT1-F77A (MI vs sham) comparison only ($q\text{-value}=1.36\times 10^{-2}$), while ribosome pathway was significantly differentially regulated in wild-type (MI vs sham) comparison only ($q\text{-value}=3.94 \times 10^{-2}$).

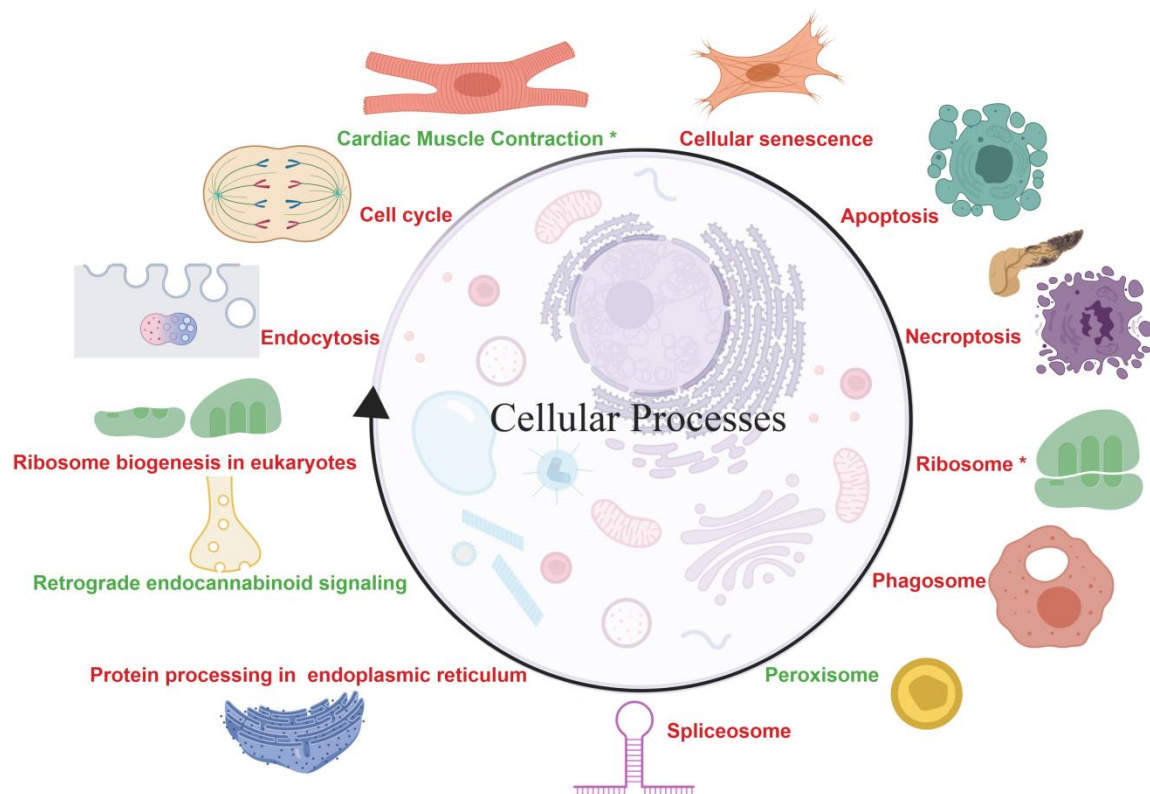


Figure 28: List of KEGG differentially regulated cellular processes following LAD ligation in wild-type and STAT1-F77A mice. Upregulated pathways are displayed in red and downregulated pathways are in green. *Cardiac muscle contraction is uniquely differentially regulated in STAT1-F77A (MI vs sham) comparison only, while ribosome pathway is uniquely differentially regulated in wild-type (MI vs sham) comparison only.

The diagram illustrates the complex pathways of endocytosis and vesicle trafficking. It is organized into several main sections:

- Clathrin-dependent endocytosis:** This pathway is shown on the left, involving receptors like TGF β , transferrin, and growth factor (e.g., EGF). It features adaptor proteins such as SARA, SARM, and AP-2, leading to the formation of clathrin-coated pits. The process is regulated by the TGF β -beta signaling pathway and involves ubiquitination and the ESCRT machinery.
- Clathrin-independent endocytosis:** This pathway is shown on the right, involving receptors like IL2R and LMR1. It features adaptor proteins such as PIP3K, PLD, and RhoA, leading to the formation of clathrin-independent vesicles. The process is regulated by the PI(3,4,5)P3 signaling system and involves the formation of a clathrin-coated vesicle.
- Fast recycling:** This pathway involves the rapid return of receptors to the cell surface, mediated by the ERC (early endosome recycling compartment) and the ESCRT machinery.
- Slow recycling:** This pathway involves the slower return of receptors to the cell surface, mediated by the ARF6-dependent recycling pathway and the ESCRT machinery.
- Early endosome (pH 6.2):** This compartment is shown in the center, where various receptors and adaptor proteins are sorted for recycling or degradation. It is characterized by the presence of VPS29, VPS35, and VPS36.
- Late endosome and multivesicular body (pH 5.2):** This compartment is shown at the bottom, where the degradation of internalized material occurs. It is characterized by the presence of VPS36, VPS22, and VPS25.
- Trans Golgi network and Golgi body:** These compartments are shown on the left, where the sorting of receptors and adaptor proteins for endocytosis occurs. The Golgi body is characterized by the presence of SNX1/2 and SNX5.

The diagram also includes a color-coded pH scale from -2 to 2, indicating the pH of different compartments. The color scale ranges from red (-2) to green (2).

ENDOCYTOSIS

Clathrin-dependent endocytosis

Stimulated by: ARF6, PIP3K, PLD, LDL, Cytokine: cytokine receptor interaction

Receptors: TGF β , transferrin, growth factor (e.g. EGF), EGF, LDL, Cytokine: cytokine receptor interaction

Adaptor proteins: SARA, cPML, SARM2, SARM3, SARM4, SARM5, SARM6, SARM7, SARM8, SARM9, SARM10, SARM11, SARM12, SARM13, SARM14, SARM15, SARM16, SARM17, SARM18, SARM19, SARM20, SARM21, SARM22, SARM23, SARM24, SARM25, SARM26, SARM27, SARM28, SARM29, SARM30, SARM31, SARM32, SARM33, SARM34, SARM35, SARM36, SARM37, SARM38, SARM39, SARM40, SARM41, SARM42, SARM43, SARM44, SARM45, SARM46, SARM47, SARM48, SARM49, SARM50, SARM51, SARM52, SARM53, SARM54, SARM55, SARM56, SARM57, SARM58, SARM59, SARM60, SARM61, SARM62, SARM63, SARM64, SARM65, SARM66, SARM67, SARM68, SARM69, SARM70, SARM71, SARM72, SARM73, SARM74, SARM75, SARM76, SARM77, SARM78, SARM79, SARM80, SARM81, SARM82, SARM83, SARM84, SARM85, SARM86, SARM87, SARM88, SARM89, SARM90, SARM91, SARM92, SARM93, SARM94, SARM95, SARM96, SARM97, SARM98, SARM99, SARM100

Clathrin-coated pit

Clathrin-coated vesicle

Fast recycling

Slow recycling

ARF6-dependent recycling

Caveola vesicle

Caveosome

To ER

Clathrin-independent endocytosis

Stimulated by: ARF6, Sec, PIP3K, PLD, H β 2R, MHC1, folic acid, Vln entry Potocytosis, Cereola

Receptors: ARF6, Sec, PIP3K, PLD, H β 2R, MHC1, folic acid, Vln entry Potocytosis, Cereola

Adaptor proteins: SARA, cPML, SARM2, SARM3, SARM4, SARM5, SARM6, SARM7, SARM8, SARM9, SARM10, SARM11, SARM12, SARM13, SARM14, SARM15, SARM16, SARM17, SARM18, SARM19, SARM20, SARM21, SARM22, SARM23, SARM24, SARM25, SARM26, SARM27, SARM28, SARM29, SARM30, SARM31, SARM32, SARM33, SARM34, SARM35, SARM36, SARM37, SARM38, SARM39, SARM40, SARM41, SARM42, SARM43, SARM44, SARM45, SARM46, SARM47, SARM48, SARM49, SARM50, SARM51, SARM52, SARM53, SARM54, SARM55, SARM56, SARM57, SARM58, SARM59, SARM60, SARM61, SARM62, SARM63, SARM64, SARM65, SARM66, SARM67, SARM68, SARM69, SARM70, SARM71, SARM72, SARM73, SARM74, SARM75, SARM76, SARM77, SARM78, SARM79, SARM80, SARM81, SARM82, SARM83, SARM84, SARM85, SARM86, SARM87, SARM88, SARM89, SARM90, SARM91, SARM92, SARM93, SARM94, SARM95, SARM96, SARM97, SARM98, SARM99, SARM100

Phosphatidylinositol signaling system

lipid raft

Caveola vesicle

Caveosome

To ER

Early endosome pH 6.2

Late endosome and multivesicular body pH 5.2

Lysosome

Trans Golgi network

Golgi body

SNARE interactions in vesicular transport

ESCRT-0

ESCRT-I

ESCRT-II

ESCRT-III

disassembly of ESCRTs

AAA-ATPase

ADP

ATP

UBPY

AMSH

deubiquitinating enzymes

SNX12

SNX5

SNX13

SNX14

SNX15

SNX16

SNX17

SNX18

SNX19

SNX20

SNX21

SNX22

SNX23

SNX24

SNX25

SNX26

SNX27

SNX28

SNX29

SNX30

SNX31

SNX32

SNX33

SNX34

SNX35

SNX36

SNX37

SNX38

SNX39

SNX40

SNX41

SNX42

SNX43

SNX44

SNX45

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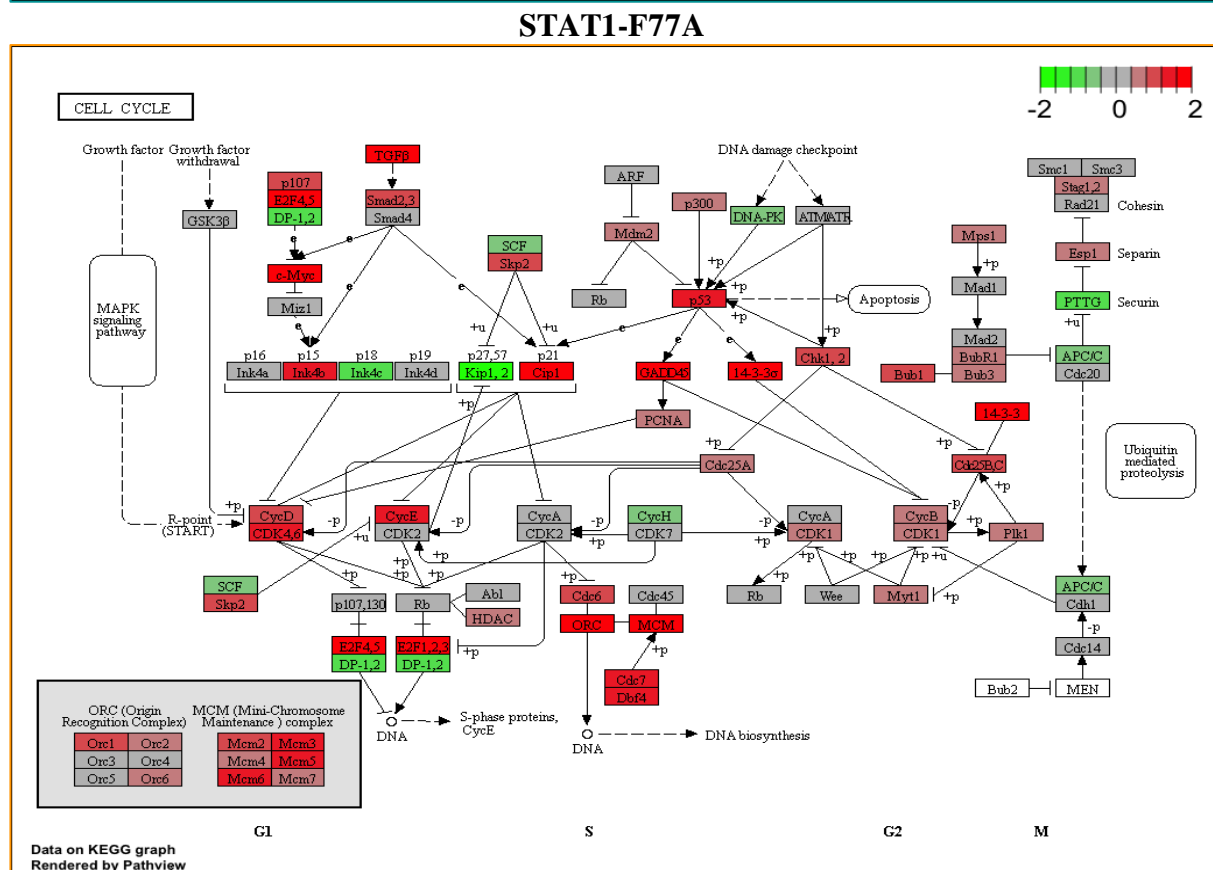
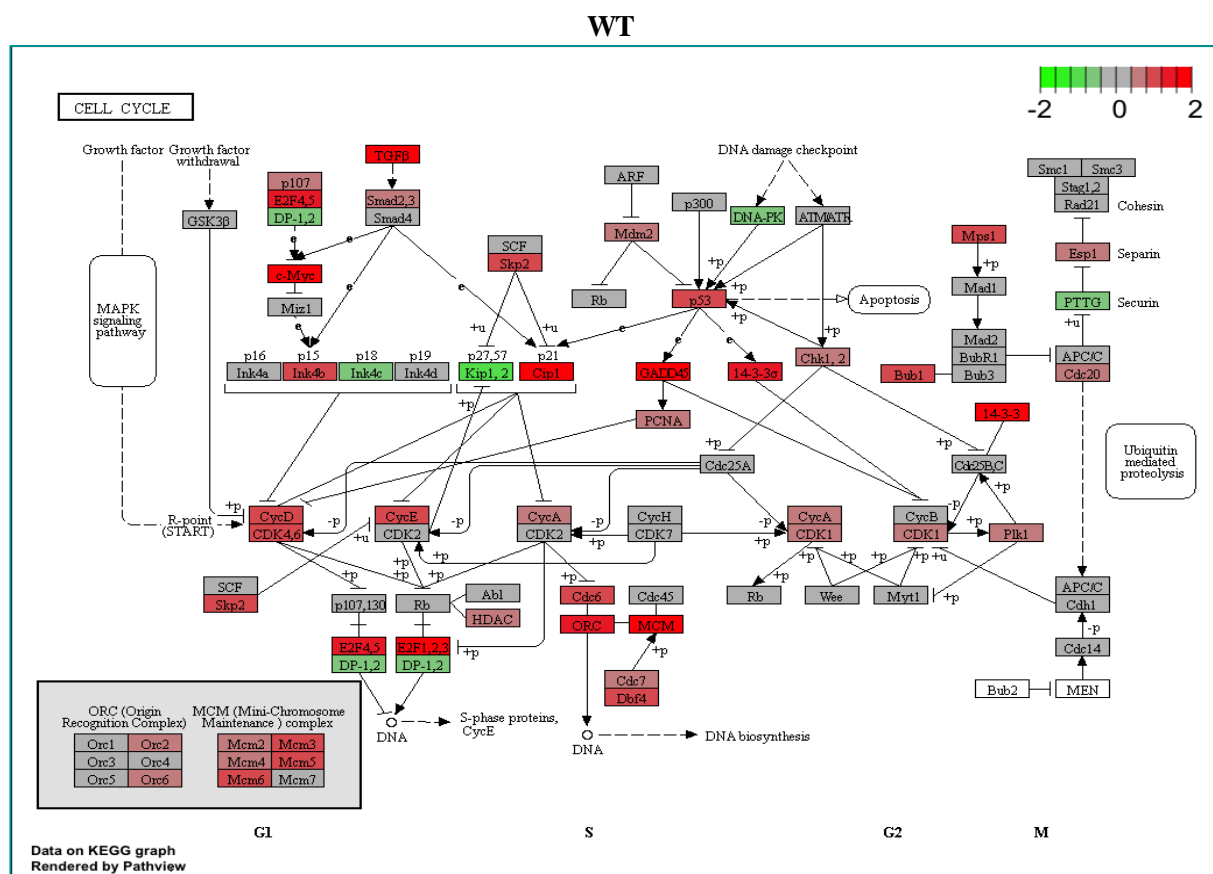
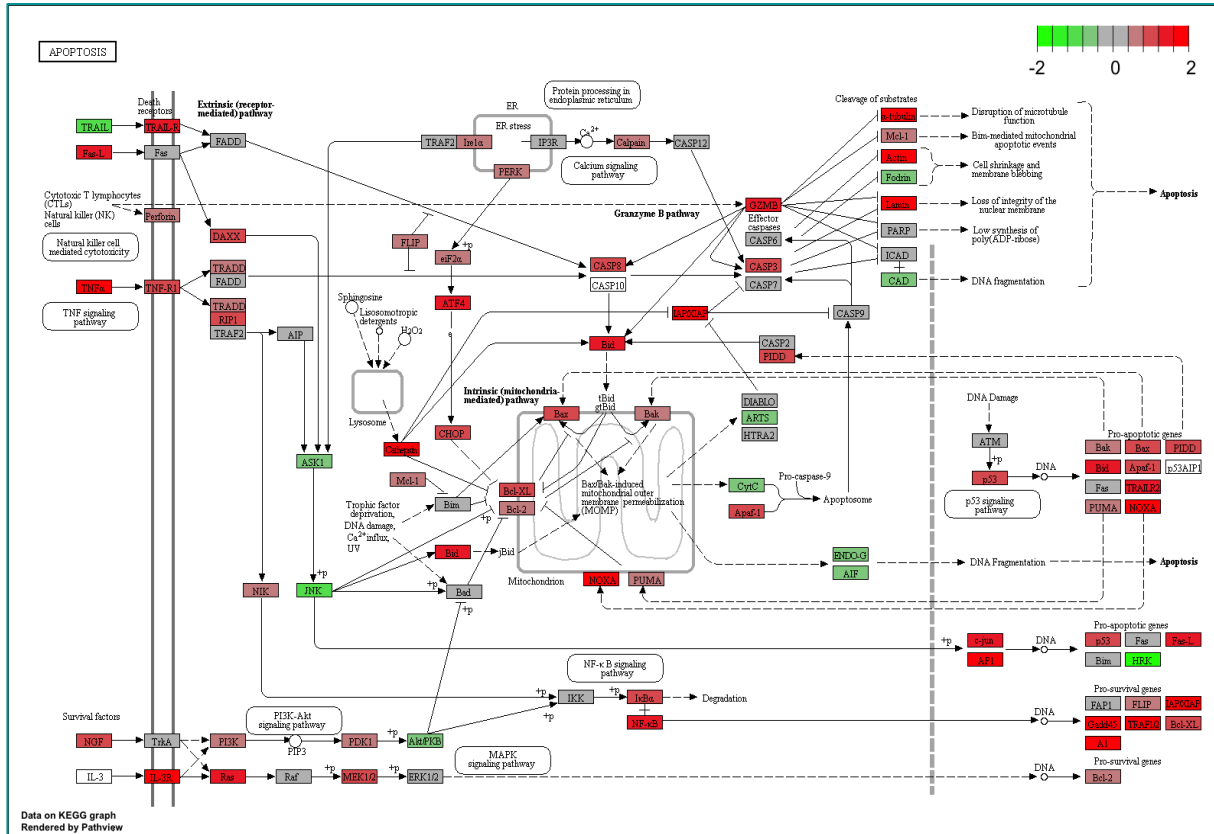


Figure 30: Pathview of the “cell cycle” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

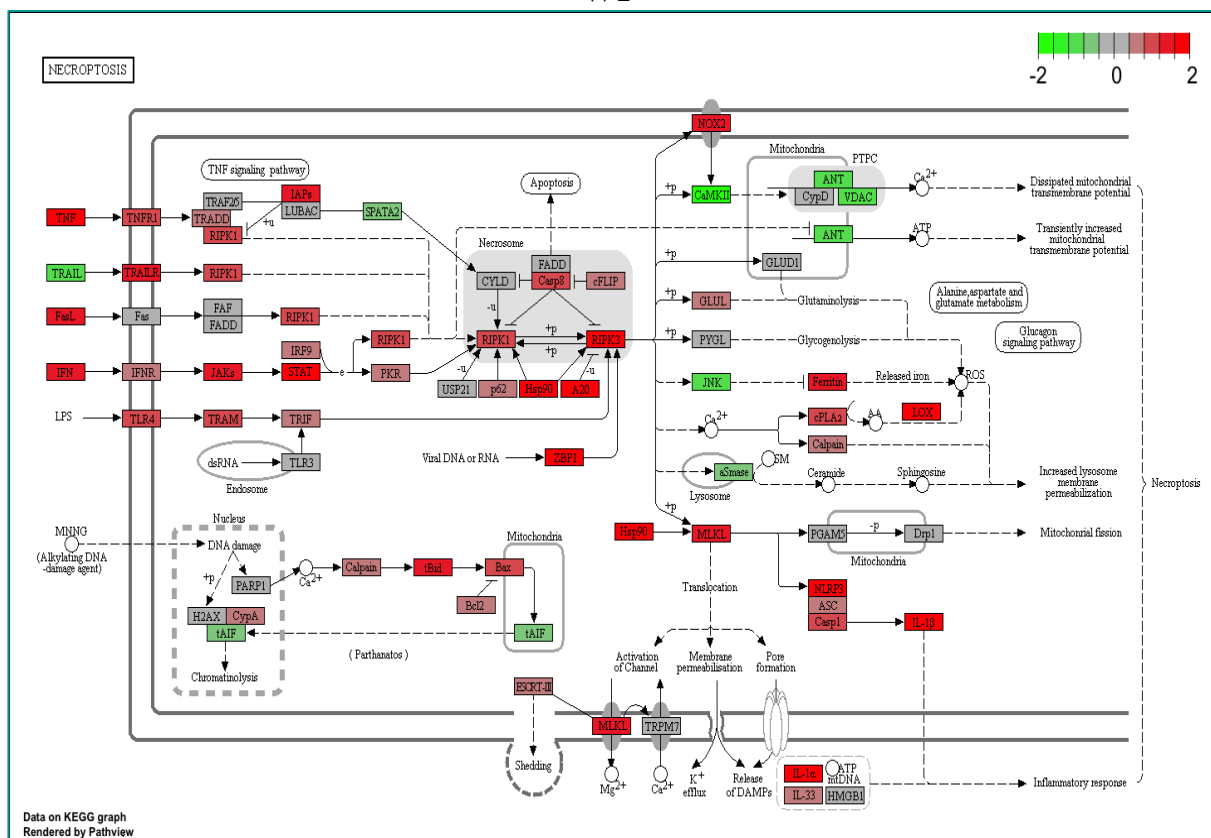




WT



WT



STAT1-F77A

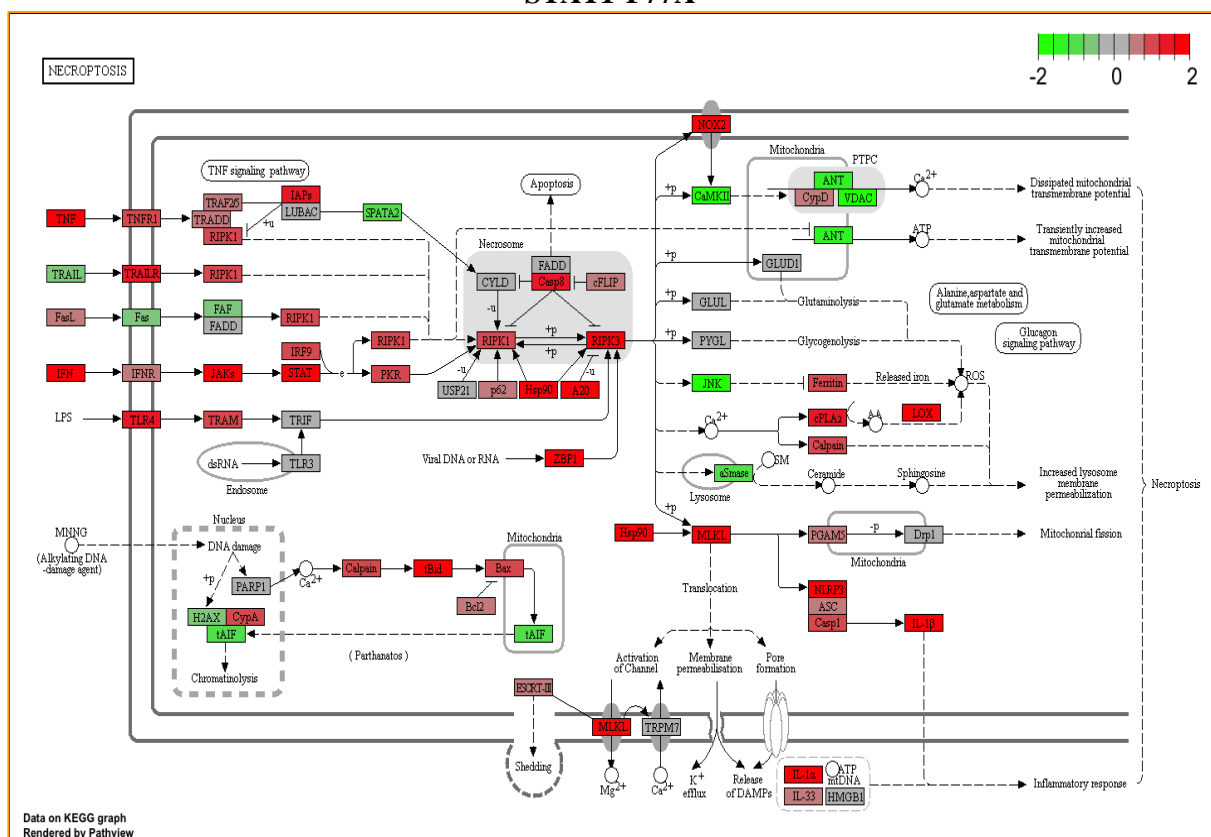
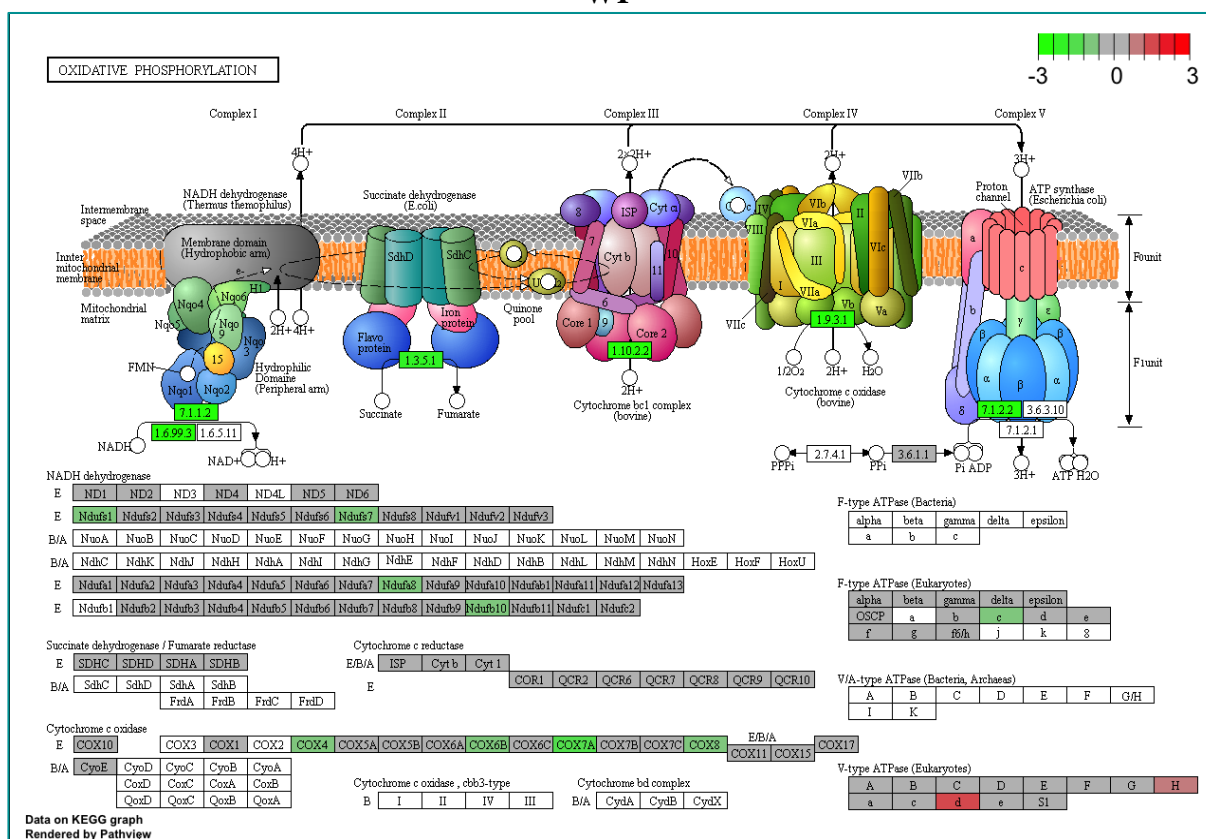


Figure 33: Pathview of the “necroptosis” KEGG pathway in mice expressing wild-type STAT1 and STAT1-F77A following LAD ligation.

3.8 Gene set enrichment analysis revealed changes in metabolic signature following myocardial infarction

The heart has the capacity to metabolize various types of substrates to synthesize ATP. In this study, we observed that both processes of energy production and energy utilization are dysregulated in the failing heart. Differential analysis revealed downregulation of metabolic pathways, involving oxidative phosphorylation (Figure 34), citrate cycle (TCA cycle) (Figure 35), fatty acid metabolism (Figure 36), fatty acid degradation (Figure 37), propanoate metabolism (Figure 38), tryptophan metabolism, pyruvate metabolism, valine, leucine and isoleucine degradation and glycosaminoglycan biosynthesis in both wild-type and STAT1-F77A following myocardial infarction. Whereas lysine degradation was only significantly differentially regulated in wild-type (MI vs sham) comparison, other pathways significantly differentially regulated in STAT1-F77A (MI vs sham) were butanoate metabolism, glyoxylate and dicarboxylate metabolism, carbon metabolism, drug metabolism-cytochrome P450, PPAR signalling pathway, and metabolism of xenobiotics by cytochrome P450. Similarly, performing hierarchical clustering analysis showed that the 1428 DEGs which are downregulated in STAT1-F77A have similar functions, which are involved in the regulation of metabolic pathways related to oxidative phosphorylation, citrate cycle (TCA cycle), fatty acid metabolism, fatty acid degradation, propanoate and tryptophan metabolism, pyruvate metabolism, and valine, leucine and isoleucine degradation. In addition, those genes were associated with glycosaminoglycan biosynthesis, butanoate metabolism, glyoxylate and dicarboxylate metabolism, carbon metabolism, drug metabolism and cytochrome P450, and PPAR signalling pathway (Figure 39). From these data, we hypothesize that the differential gene expression pattern between animals from the two genotypes resulted from a higher immune cell infiltration in STAT1-F77A mice, demonstrating a metabolic shift from oxidative phosphorylation towards glycolysis, which is more prominent in the knock-in mice.

WT



STAT1-F77A

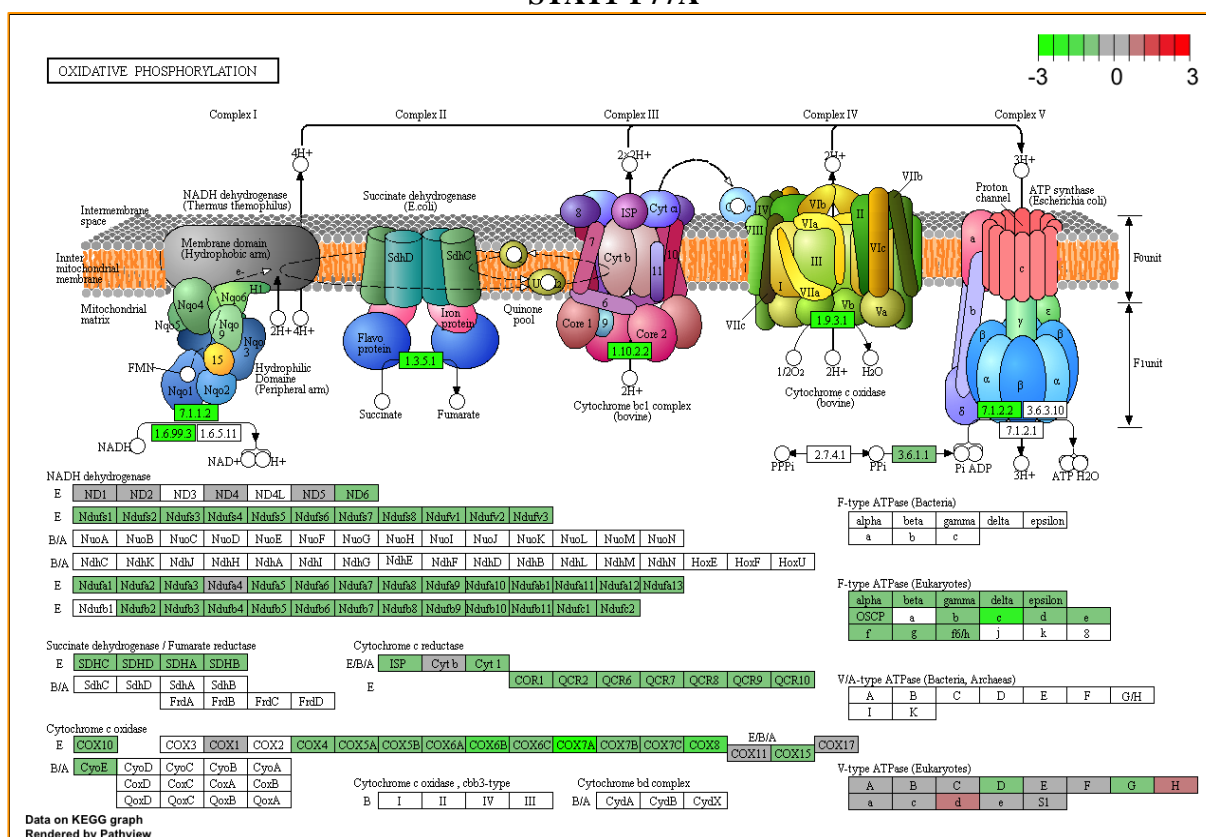
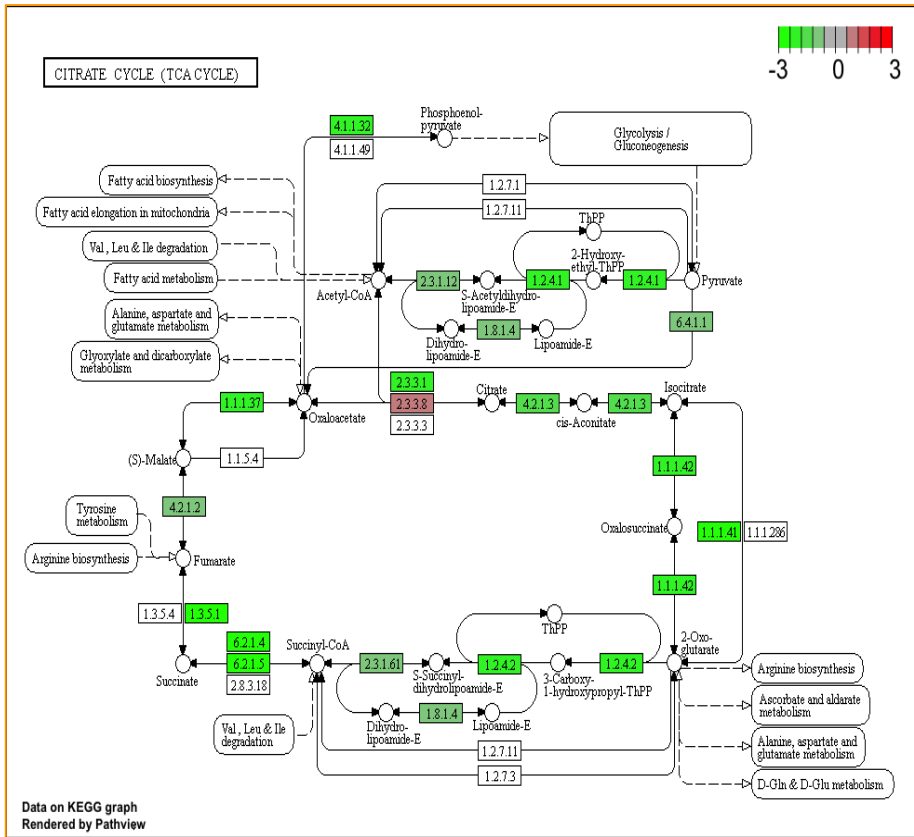


Figure 34: Pathview of the “oxidative phosphorylation” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

STAT1-F77A



WT

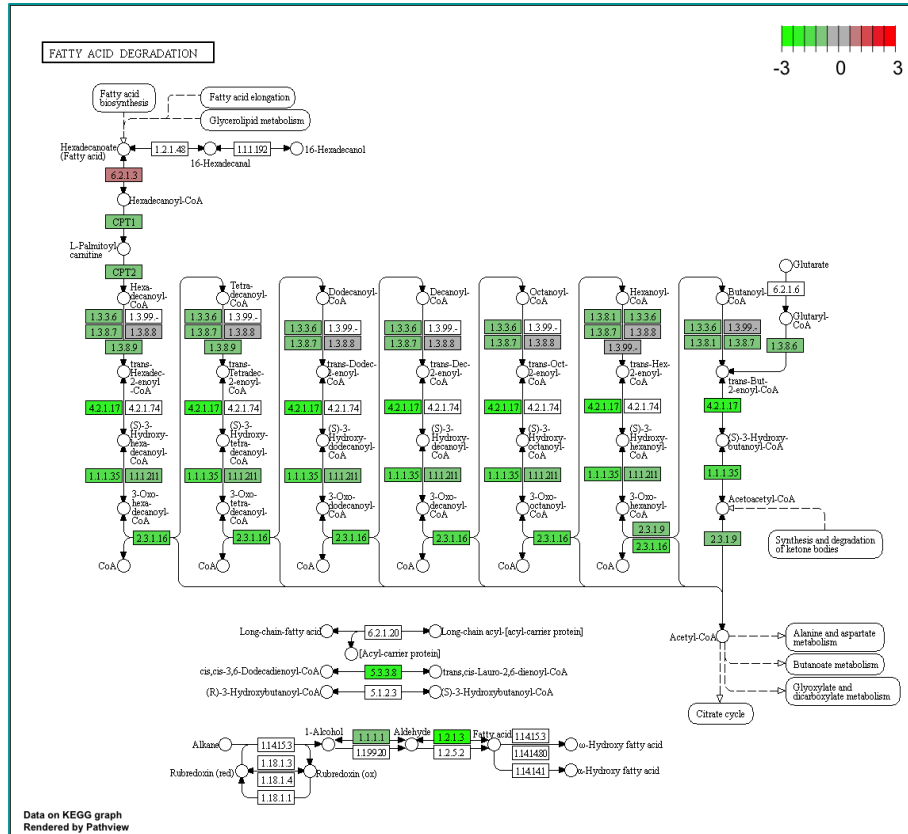
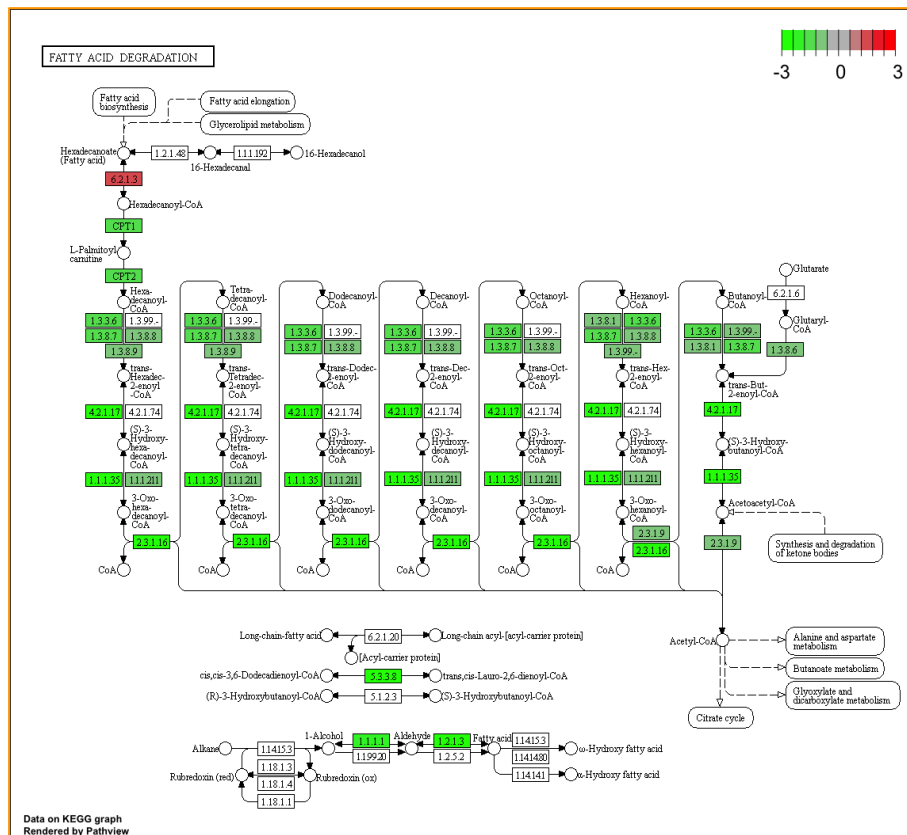
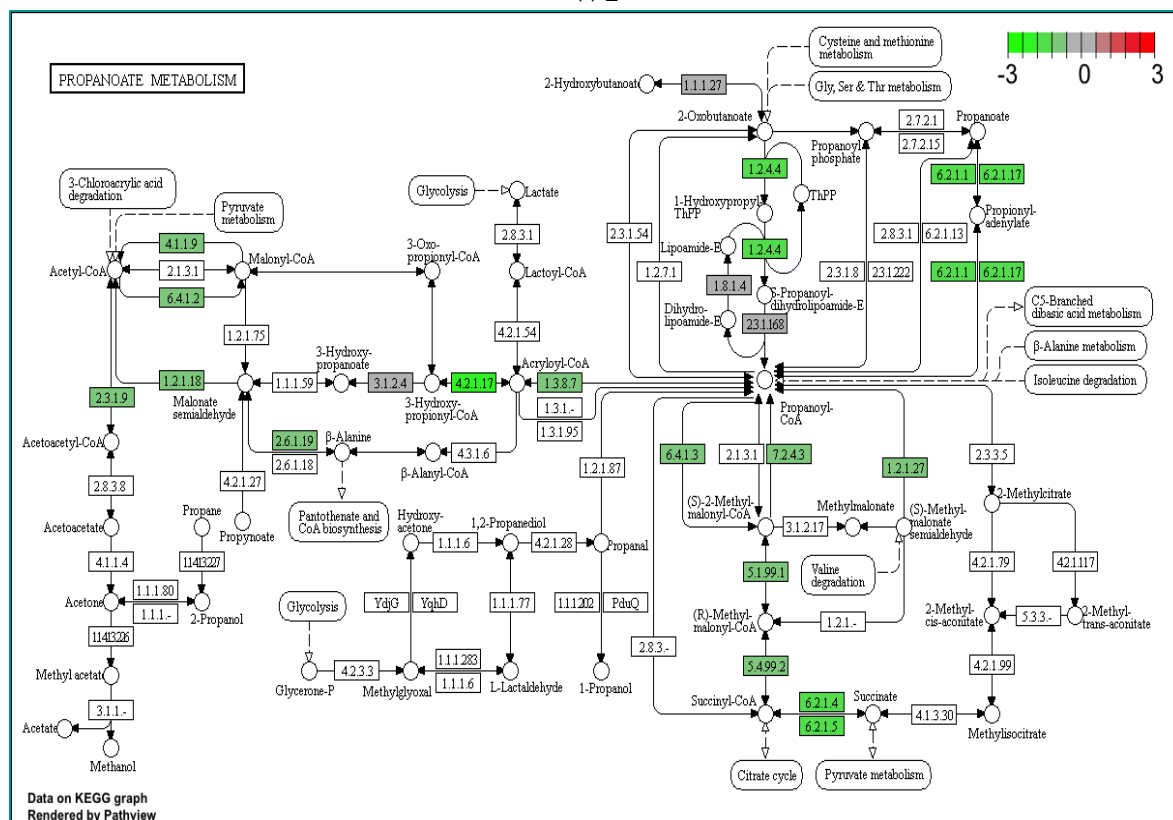
**STAT1-F77A**

Figure 37: Pathview of the “fatty acid degradation” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.

WT



STAT1-F77A

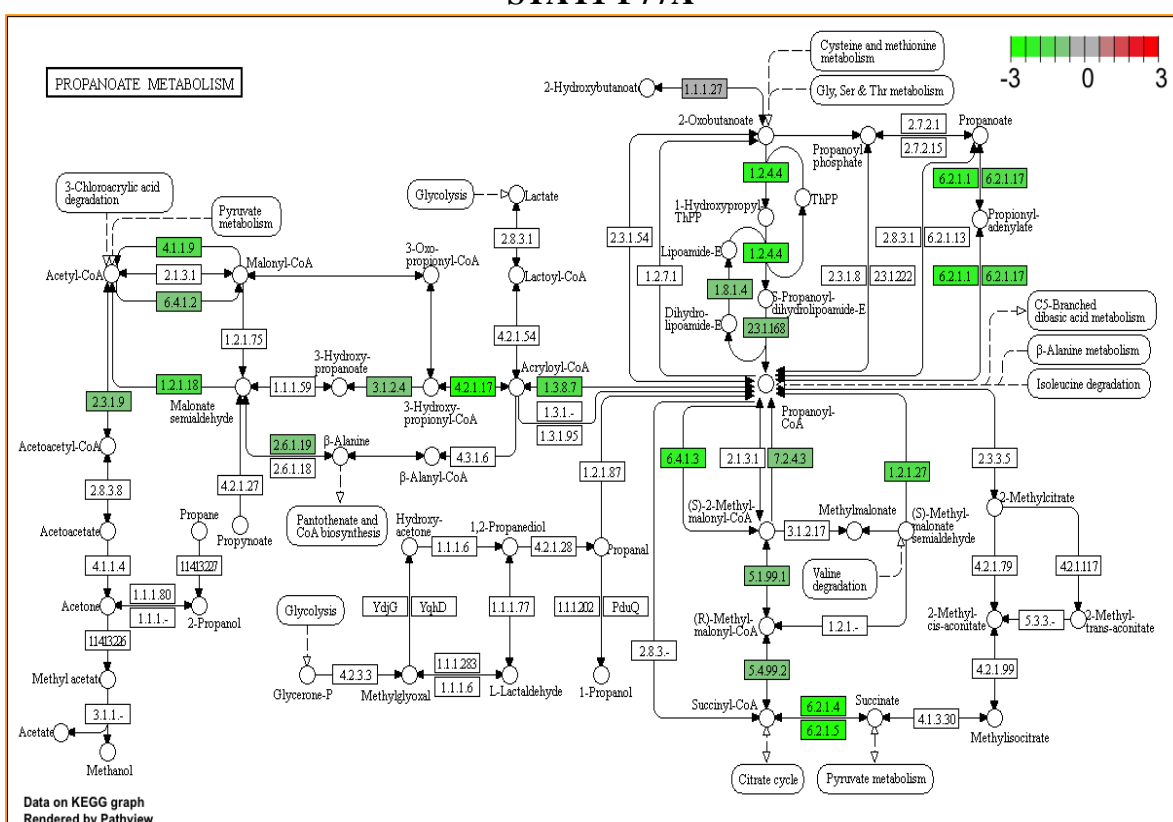


Figure 38: Pathview of the “proanoate metabolism” KEGG pathway in wild-type and STAT1-F77A mice following LAD ligation.



Figure 39: Gene set enrichment analysis of differentially downregulated genes in STAT1-F77A post-MI.

3.9 Differentially regulated pathways in F77A as compared to wild-type mice in the infarcted area following LAD ligation

Using KEGG pathway enrichment analysis, 18 KEGG pathways were identified when comparing STAT1-F77A and wild-type mice post-LAD ligation, among which oxidative phosphorylation, cardiac muscle contraction, citrate cycle were observed to be the top three in mean enrichment score (Figure 40). Table 31 below illustrates the KEGG pathway ID, KEGG name, mean enrichment score, *p*-value, *q*-value, and the number of genes in each respective pathway comparing STAT1-F77A and wild-type mice post-MI. Furthermore, gene set enrichment analysis revealed differential expression of cardiac muscle contraction pathway only in STAT1-F77A mice at day 1 post-MI (Figure 41).

Table 31: List of KEGG differentially regulated pathways comparing STAT1-F77A and wild-type mice post-MI.

| KEGG ID | KEGG pathway | mean | <i>p</i> -value | <i>q</i> -value | size |
|----------|--|------|-----------------|-----------------|------|
| mmu00190 | Oxidative phosphorylation | -8.2 | 1.48e-14 | 3.26e-12 | 116 |
| mmu04260 | Cardiac muscle contraction | -4.9 | 1.33e-06 | 1.46e-04 | 67 |
| mmu00020 | Citrate cycle (TCA cycle) | -4.1 | 7.43e-05 | 5.45e-03 | 30 |
| mmu00280 | Valine, leucine and isoleucine degradation | -3.8 | 1.36e-04 | 6.53e-03 | 47 |
| mmu00640 | Propanoate metabolism | -3.9 | 1.48e-04 | 6.53e-03 | 29 |
| mmu04621 | NOD-like receptor signalling pathway | 4 | 3.46e-05 | 7.61e-03 | 146 |
| mmu04723 | Retrograde endocannabinoid signalling | -3.5 | 3.08e-04 | 1.13e-02 | 122 |
| mmu03008 | Ribosome biogenesis in eukaryotes | 3.4 | 5.00e-04 | 3.86e-02 | 72 |
| mmu04380 | Osteoclast differentiation | 3.3 | 5.26e-04 | 3.86e-02 | 122 |
| mmu00620 | Pyruvate metabolism | -3.1 | 1.34e-03 | 4.22e-02 | 33 |
| mmu04657 | IL-17 signalling pathway | 3 | 1.60e-03 | 6.46e-02 | 78 |
| mmu04144 | Endocytosis | 3 | 1.65e-03 | 6.46e-02 | 240 |
| mmu04668 | TNF signalling pathway | 2.9 | 1.85e-03 | 6.46e-02 | 108 |
| mmu04060 | Cytokine-cytokine receptor interaction | 2.8 | 2.43e-03 | 6.46e-02 | 204 |
| mmu03013 | RNA transport | 2.8 | 2.46e-03 | 6.46e-02 | 150 |
| mmu04062 | Chemokine signalling pathway | 2.8 | 2.64e-03 | 6.46e-02 | 167 |
| mmu01200 | Carbon metabolism | -2.8 | 3.11e-03 | 8.56e-02 | 106 |
| mmu04218 | Cellular senescence | 2.7 | 4.17e-03 | 9.17e-02 | 155 |

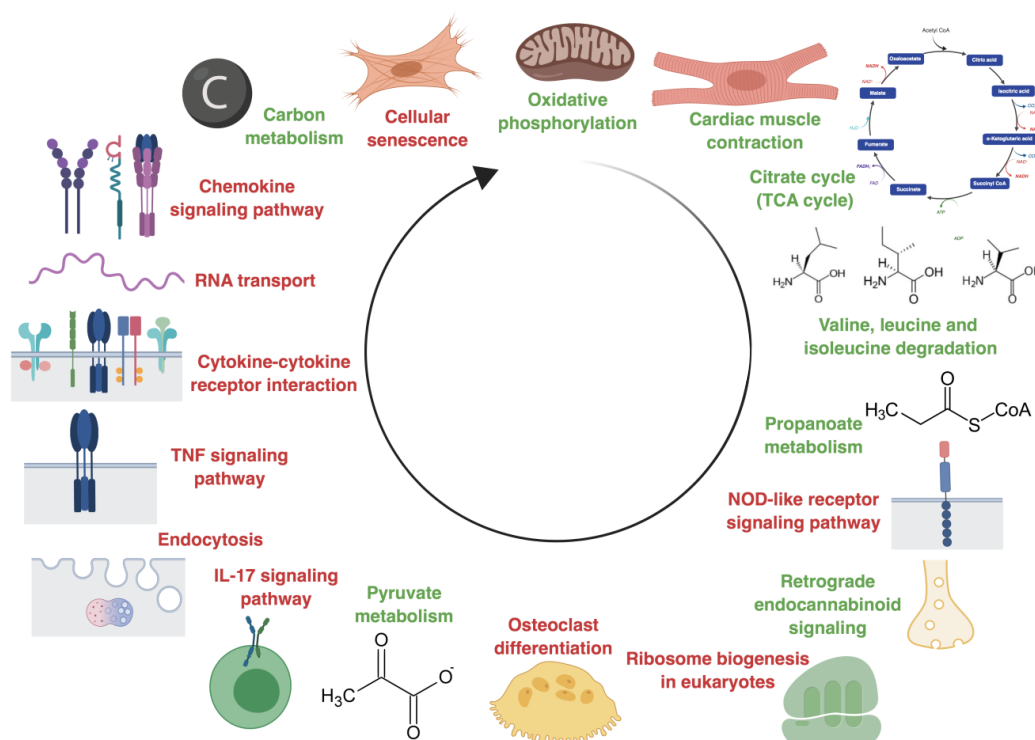
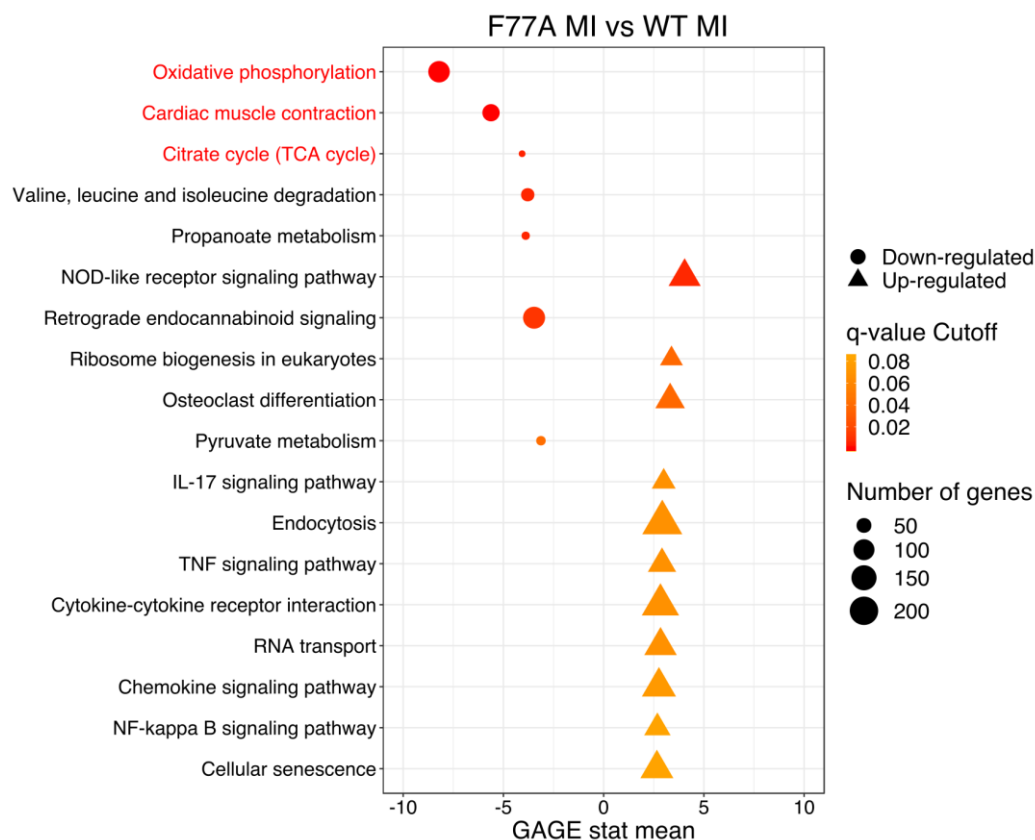


Figure 40: List of KEGG differentially regulated pathways comparing STAT1-F77A MI and wild-type MI mice. GSEA-enriched pathways with q -value < 0.1 are shown.

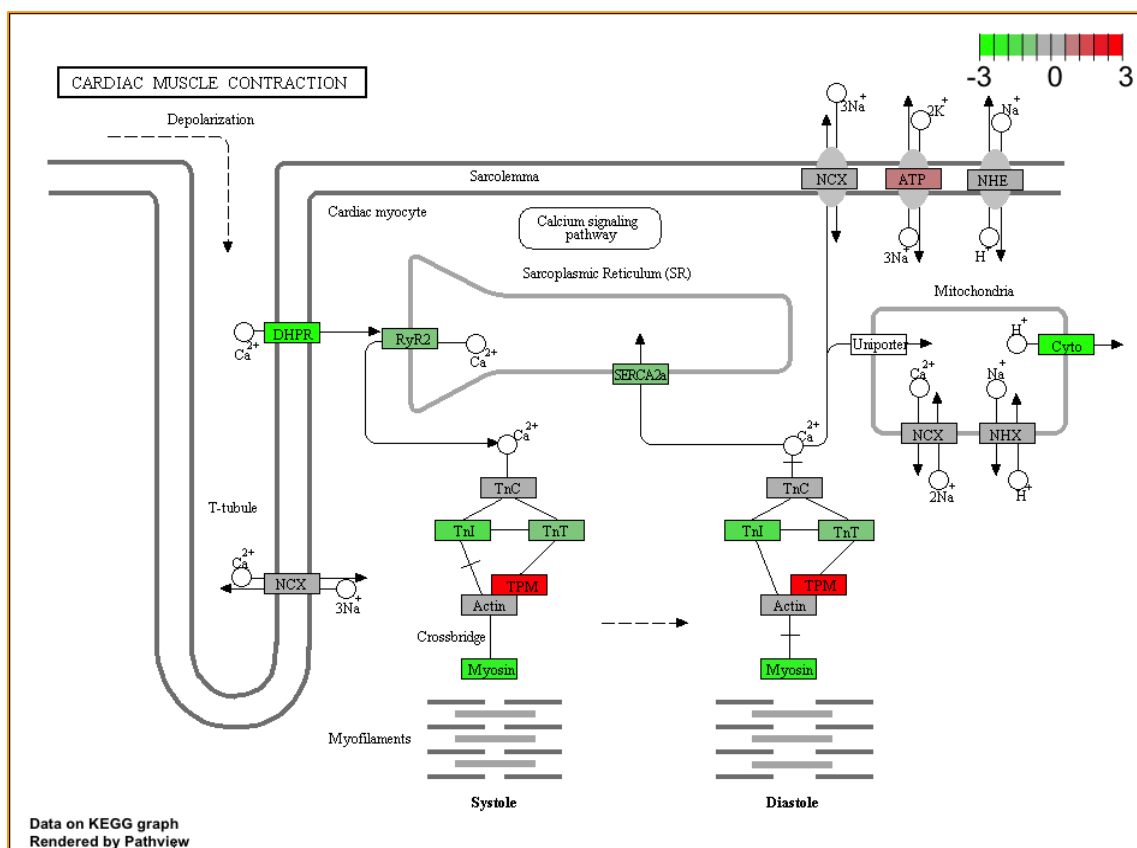
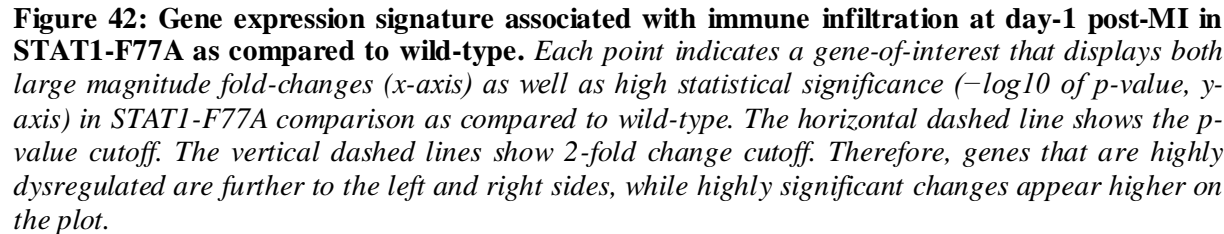


Figure 41: Pathview of the “cardiac muscle contraction” KEGG pathway in STAT1-F77A following LAD ligation.

3.10 A gene expression signature indicative of immune cells infiltration detected in F77A mice following LAD ligation

Recent studies have demonstrated the prognostic and predictive roles of immune markers in several inflammatory diseases. Nevertheless, additional characterization of these markers in the context of myocardial infarction is evidently warranted. We aimed at identifying putative target genes that are differentially regulated in STAT1-F77A as compared to wild-type. A list of immune markers signifying an immunologically active microenvironment was identified in STAT1-F77A mice with higher cut off value ($-\log_{10}(\text{padj})=11$) (Figure 42 and Table 32).



87

Table 32: Genes associated with immune cells activation at day 1 following LAD ligation in STAT1-F77A

| Gene | log ₂ fold change | log ₁₀ padj | KEGG pathway |
|----------------|------------------------------|------------------------|---|
| <i>Ccl2</i> | 3.08 | 1.34e+01 | Chemokine signalling pathway |
| <i>Ccl22</i> | 3.31 | 1.17e+01 | Chemokine signalling pathway |
| <i>Ccr1</i> | 3.38 | 1.40e+01 | Chemokine signalling pathway |
| <i>Cd14</i> | 2.97 | 1.14e+01 | MAPK signalling pathway |
| <i>Csf2rb</i> | 3.26 | 1.38e+01 | Jak-STAT signalling pathway |
| <i>Csf3r</i> | 3.62 | 1.48e+01 | PI3K-Akt signalling pathway |
| <i>Cxcl10</i> | 3.76 | 1.23e+01 | RIG-I-like receptor signalling pathway |
| <i>Cxcl2</i> | 4.51 | 1.79e+01 | Chemokine signalling pathway |
| <i>Cxcl3</i> | 4.76 | 1.90e+01 | Chemokine signalling pathway |
| <i>Cxcr2</i> | 4.19 | 1.75e+01 | Endocytosis |
| <i>Ereg</i> | 3.71 | 1.18e+01 | PI3K-Akt signalling pathway |
| <i>F10</i> | 3.69 | 1.36e+01 | Complement and coagulation cascades |
| <i>Fgf23</i> | 4.89 | 1.89e+01 | Ras signalling pathway |
| <i>Fpr1</i> | 3.88 | 1.59e+01 | Rap1 signalling pathway |
| <i>H2-Q10</i> | 3.69 | 1.37e+01 | Endocytosis |
| <i>Hspa1a</i> | 4.12 | 1.33e+01 | Protein processing in endoplasmic reticulum |
| <i>Hspa1b</i> | 3.94 | 1.20e+01 | Protein processing in endoplasmic reticulum |
| <i>Il18rap</i> | 3.29 | 1.26e+01 | Cytokine-cytokine receptor interaction |
| <i>Il1b</i> | 3.89 | 1.66e+01 | Necroptosis |
| <i>Il1f9</i> | 3.90 | 1.65e+01 | Cytokine-cytokine receptor interaction |
| <i>Il1r2</i> | 4.41 | 1.66e+01 | Hematopoietic cell lineage |
| <i>Il1rn</i> | 3.67 | 1.35e+01 | Cytokine-cytokine receptor interaction |
| <i>Il6</i> | 4.09 | 1.49e+01 | HIF-1 signalling pathway |
| <i>Inhba</i> | 3.87 | 1.24e+01 | TGF- β signalling pathway |
| <i>Lilrb4</i> | 2.90 | 1.16e+01 | Osteoclast differentiation |
| <i>Mefv</i> | 3.72 | 1.33e+01 | NOD-like receptor signalling pathway |
| <i>Mmp9</i> | 3.82 | 1.57e+01 | Leukocyte transendothelial migration |
| <i>Oas3</i> | 3.55 | 1.22e+01 | NOD-like receptor signalling pathway |
| <i>Osm</i> | 3.96 | 1.44e+01 | PI3K-Akt signalling pathway |
| <i>Plaur</i> | 3.69 | 1.42e+01 | Complement and coagulation cascades |
| <i>Ptgs2</i> | 4.27 | 1.46e+01 | NF- κ B signalling pathway |
| <i>S100a8</i> | 3.66 | 1.75e+01 | IL-17 signalling pathway |
| <i>S100a9</i> | 3.79 | 1.72e+01 | IL-17 signalling pathway |
| <i>Sell</i> | 3.44 | 1.45e+01 | Cell adhesion molecules (CAMs) |
| <i>Selp</i> | 4.03 | 1.85e+01 | Cell adhesion molecules (CAMs) |

| | | | |
|-----------------|------|----------|--|
| <i>Serpine1</i> | 3.66 | 1.21e+01 | p53 signalling pathway |
| <i>Thbs1</i> | 3.82 | 1.58e+01 | Focal adhesion |
| <i>Timp1</i> | 2.89 | 1.14e+01 | HIF-1 signalling pathway |
| <i>Tnc</i> | 3.66 | 1.34e+01 | Focal adhesion |
| <i>Tnfrsf9</i> | 3.95 | 1.16e+01 | Cytokine-cytokine receptor interaction |
| <i>Tnfrsf14</i> | 3.57 | 1.26e+01 | NF- κ B signalling pathway |

padj: Benjamini-Hochberg adjusted p -value

3.11 Analysis of STAT1-target genes associated with myocardial infarction

Since a perturbation in STAT1-target gene networks is linked to multiple autoimmune diseases and tumorigenesis, it is noteworthy to investigate the global transcriptional network of STAT1-target genes after myocardial infarction (Cao *et al.*, 2017). Using a list of 1,441 ChIP-Seq-based STAT1 target genes in IFN- γ -treated HeLa cells published by Satoh and Tabunoki (2013), we constructed a functional protein association network of 132 STAT1-target genes that are differentially regulated in wild-type mice (Figure 43) and another protein association network of 180 STAT1-target genes differentially regulated in STAT1-F77A (Figure 44) at day 1 following myocardial infarction. A fold change >1 was used as a cut-off value.

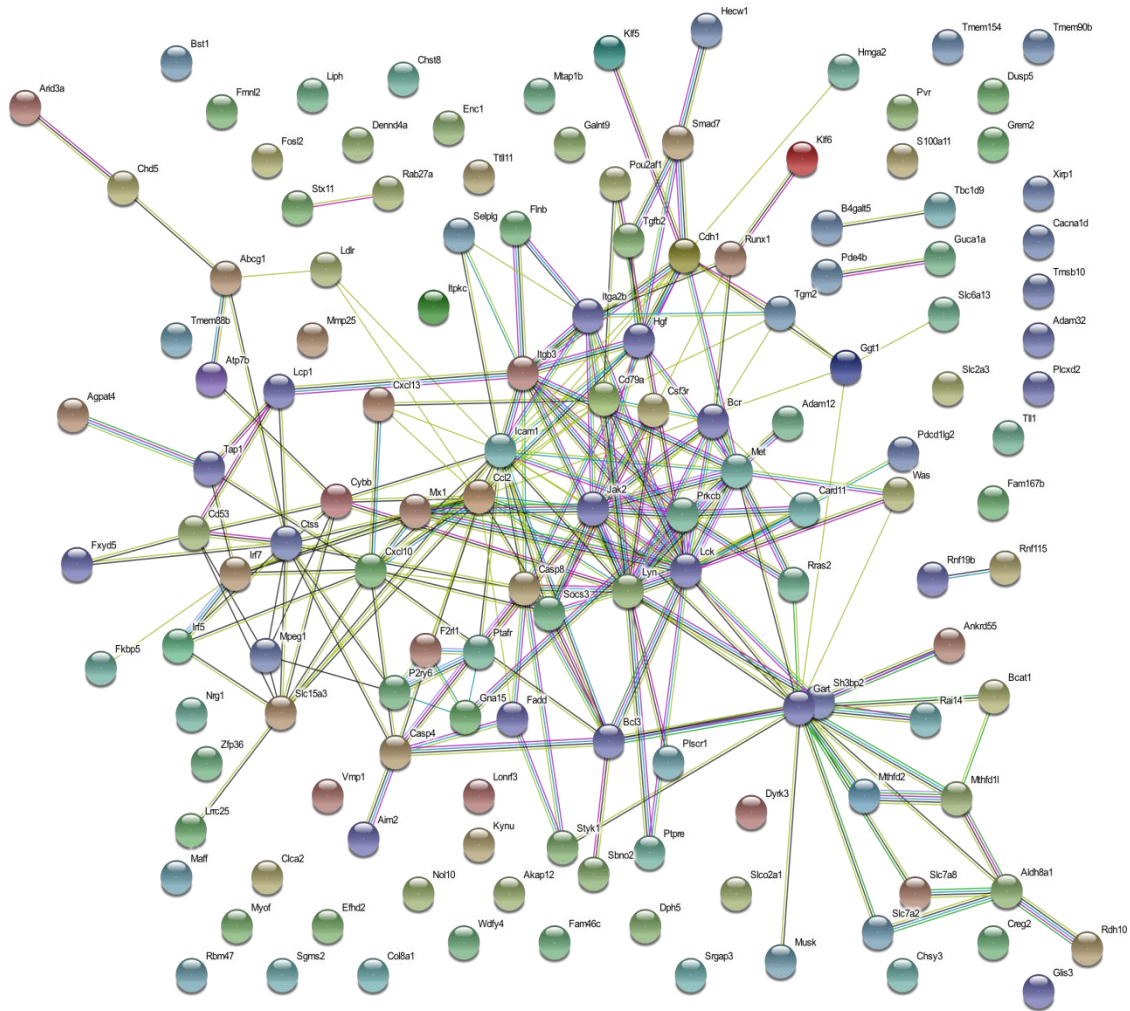


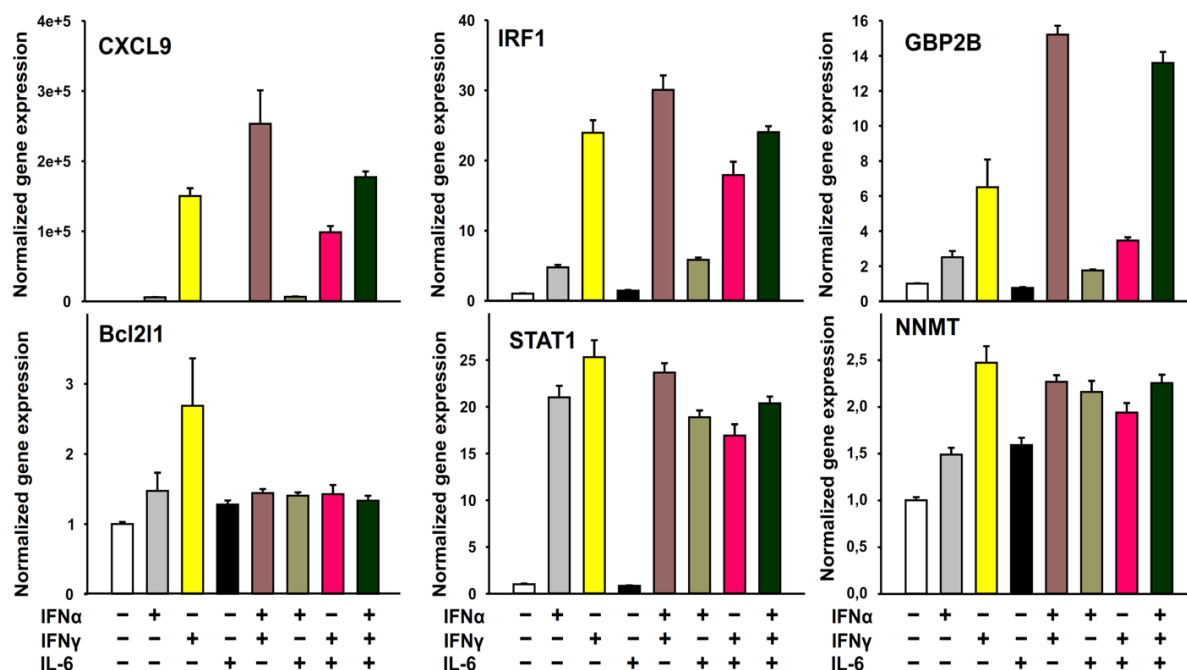
Figure 43: A functional protein association network of 132 STAT1 target genes differentially regulated in wild-type mice post-MI.

3.12 Co-stimulatory effects of interferon- γ , interferon- α , and interleukin-6 in MHEC-5 cells

91

cells compared to cells treated with IFN- γ only (Figure 45A), whereas co-stimulation with IL-6 and IFN- γ did not alter the level of STAT1 tyrosine phosphorylation in comparison to cells treated with IFN- γ only (Figure 45B).

A)



B)

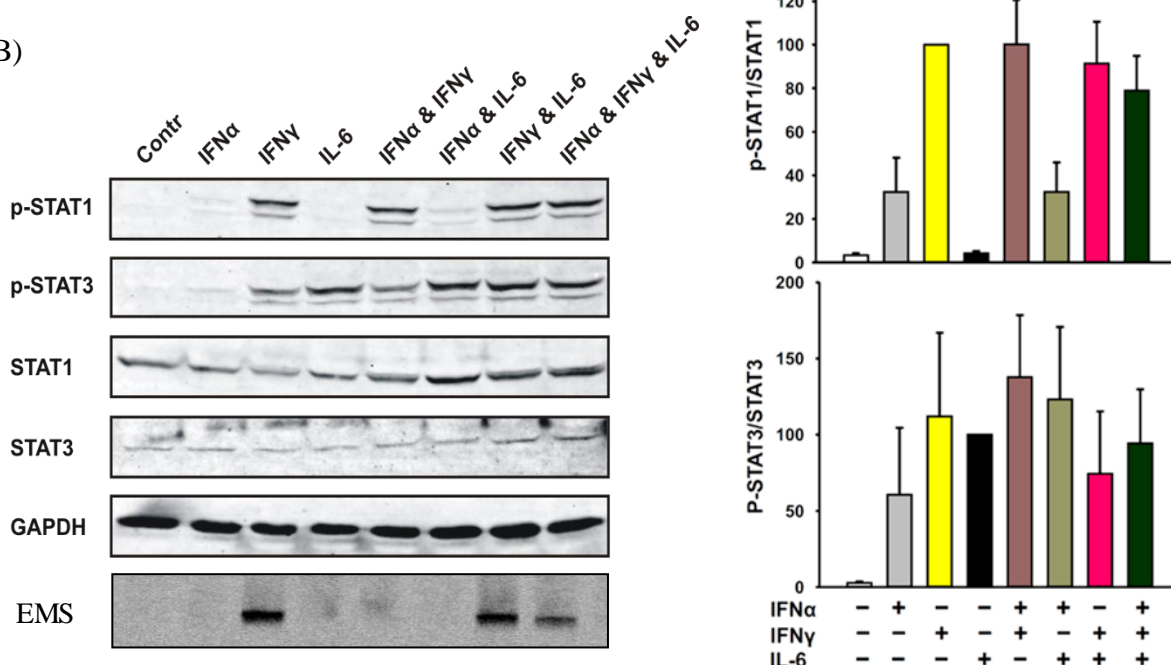


Figure 45: Co-stimulatory effects of interferon- γ , interferon- α , and interleukin-6 in MHEC-5 cells on (A) the induction of STAT1 target genes and (2) the expression and phosphorylation levels of STAT1 and STAT3 proteins

3.13 Generation of STAT1- and STAT3-knockdown HL-60 cell lines

In addition to acting as cytoskeletal proteins, the ezrin, radixin and moesin (ERM) proteins act as signaling adaptors and play a fundamental role regulating invasion and cellular cortical tension, both of which are required for efficient transendothelial migration. Given the complex phenotype presented in the STAT1-F77A mice challenged with MI, STAT1 and STAT3 were systematically knocked down as a proof of concept study to determine if reducing STAT1 expression in neutrophils would interfere with recruitment and transendothelial migration. Lentiviral vectors with short hairpin RNA (shRNA) were used to transfect the human leukemia cell line HL-60. Immunoblotting was utilized to assess the relative protein levels in STAT1 and STAT3 shRNA-transfected cells compared to cells transfected with control shRNA. Total STAT1 levels were reduced following transfection with 3 different anti-STAT1 shRNA, whereas total STAT3 levels were reduced using 5 different anti-STAT3 shRNA viral clones, as can be seen in Figure 46. HL-60 cells that are transfected with lentiviral shSTAT1 clone#3 and HL-60 cells transfected with shSTAT3 clone#5 were used in the following experiments, as they have shown the highest percentage of knockdown according to the ratios of STAT1 and STAT3 proteins to β -actin in each lane in comparison to shRNA control.

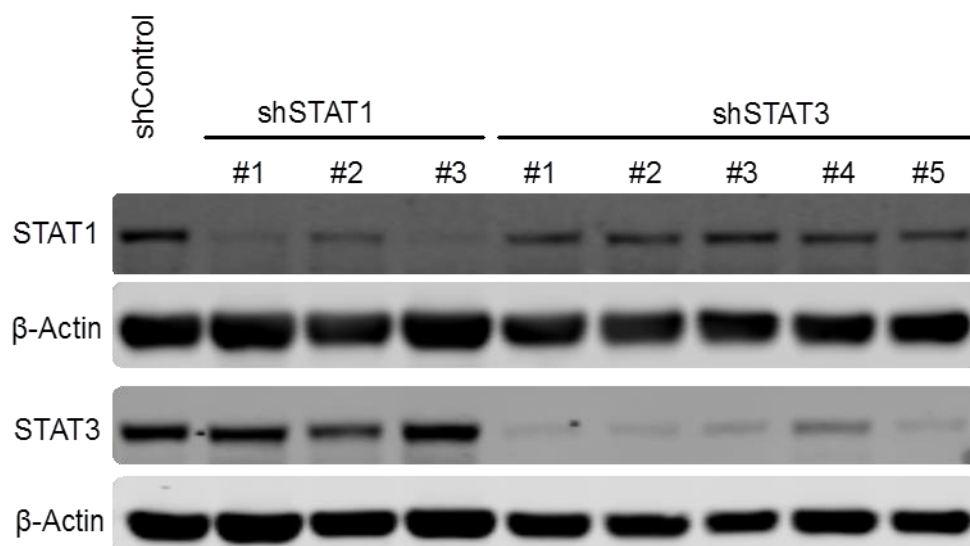


Figure 46: shRNA knockdown of STAT1 and STAT3 in HL60 untreated cells

3.14 DMSO-induced differentiation results in upregulation of STAT1 and STAT3 in HL-60 cells

In order to further characterize the mechanisms involved in HL-60 cells differentiation, we examined the kinetics of STAT1 and STAT3 expression in response to 1.3% DMSO stimulation. Protein extracts were collected at days 0, 1, 2, 3, 4, and 5 and were subjected to immunoblot analyses. DMSO stimulation led to the induction of STAT1 and STAT3 expression, where the peak of STAT1 expression was at day 4, and the peak of STAT3 expression was at day 5 in STAT1 and STAT3-knockdown cells, respectively (Figure 47). These data suggest that STATs signalling is involved in the differentiation of HL-60 cells.

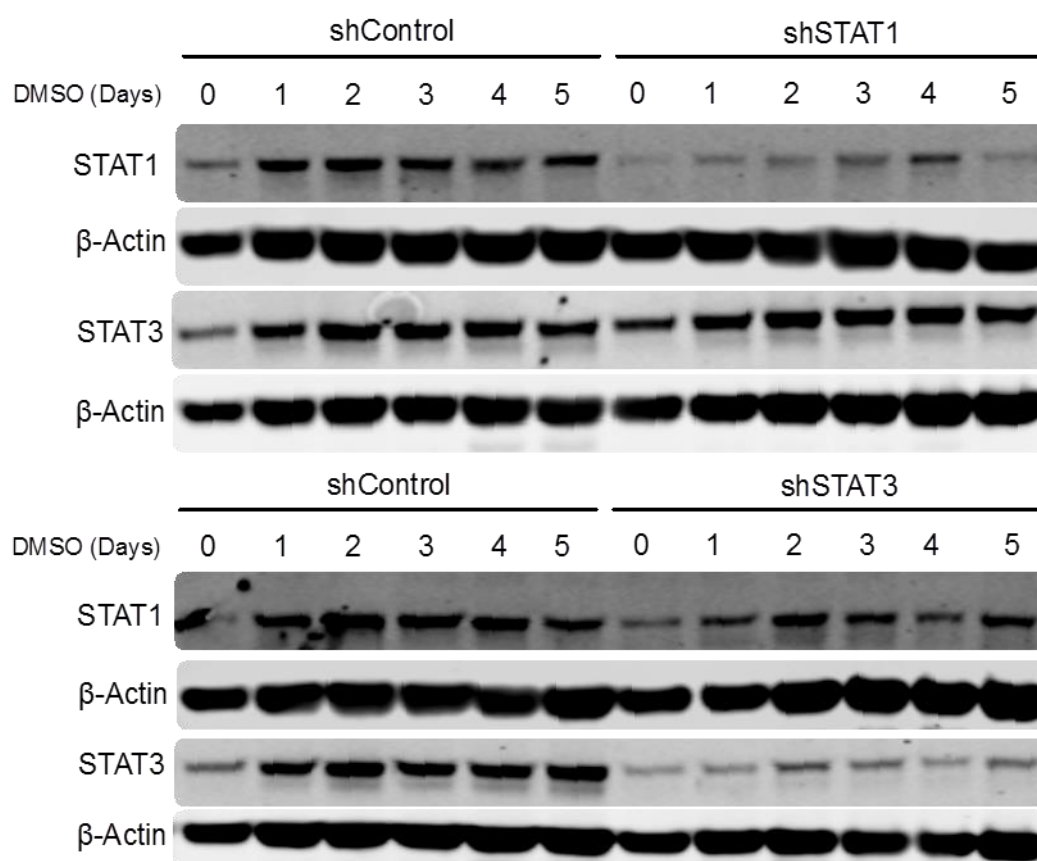


Figure 47: STAT1 and STAT3 expression levels in control as well as STAT1- and STAT3-knockdown HL-60 cells differentiated with DMSO

3.15 Decreased phosphorylation levels of STAT1 and STAT3 in HL-60 knockdown cells

To further characterize the generated STAT1- and STAT3-knockdown cells, we differentiated HL-60 cells with 1.3% DMSO and analyzed them at day 4 for the phosphorylation levels of STAT1 and STAT3. Cells were left unstimulated or stimulated with IFN- γ (50 ng/ml), IL-6

(10 ng/ml) or both IFN- γ and IL-6 for 6 and 24 hours. Our results show that STAT1 phosphorylation levels upon stimulation with IFN- γ or both IFN- γ and IL-6 are significantly increased, whereas STAT3 phosphorylation is induced in IFN- γ , IL-6 or both IFN- γ and IL-6 stimulation conditions as compared to unstimulated cells. Additionally, STAT1 and STAT3 phosphorylation levels are significantly decreased in STAT1-knockdown and STAT3-knockdown HL-60 cells, respectively, as compared to control shRNA transfected cells. Moreover, immunoblots of STAT1 and STAT3 demonstrated that downregulation of STAT1 in HL-60 cells by shRNA expression had no effect on STAT3 expression level. However, the knockdown of STAT3 in HL-60 cells by shRNA had a negative effect on the expression levels of STAT1. Furthermore, the phosphorylation levels of STAT1 were reduced in STAT3-knockdown HL-60 cells as compared to control shRNA (Figure 48 and Figure 49).

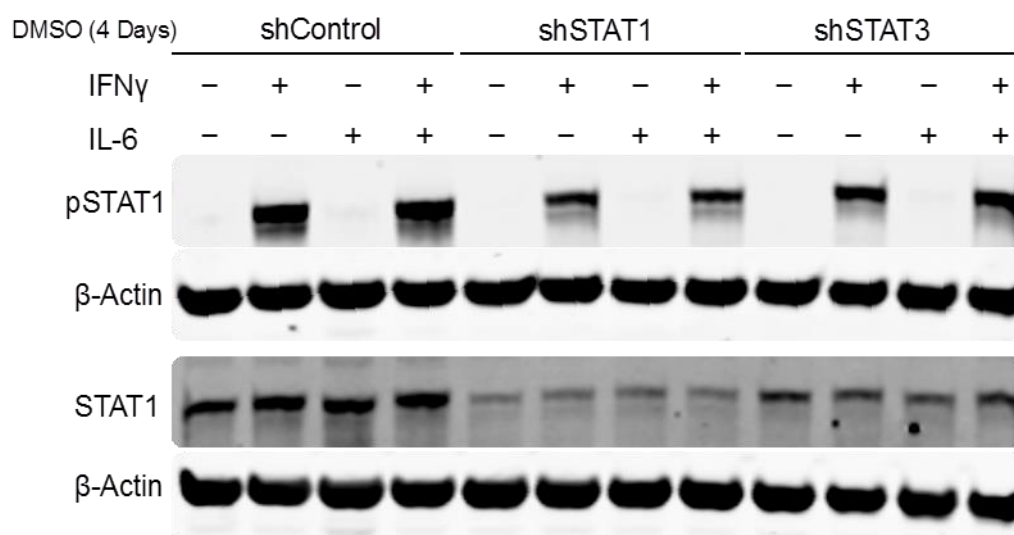


Figure 48: STAT1 expression levels and phosphorylation in HL-60 cells differentiated with DMSO and stimulated at day 4 with IFN- γ , IL-6 or the combination of both for 45 min.

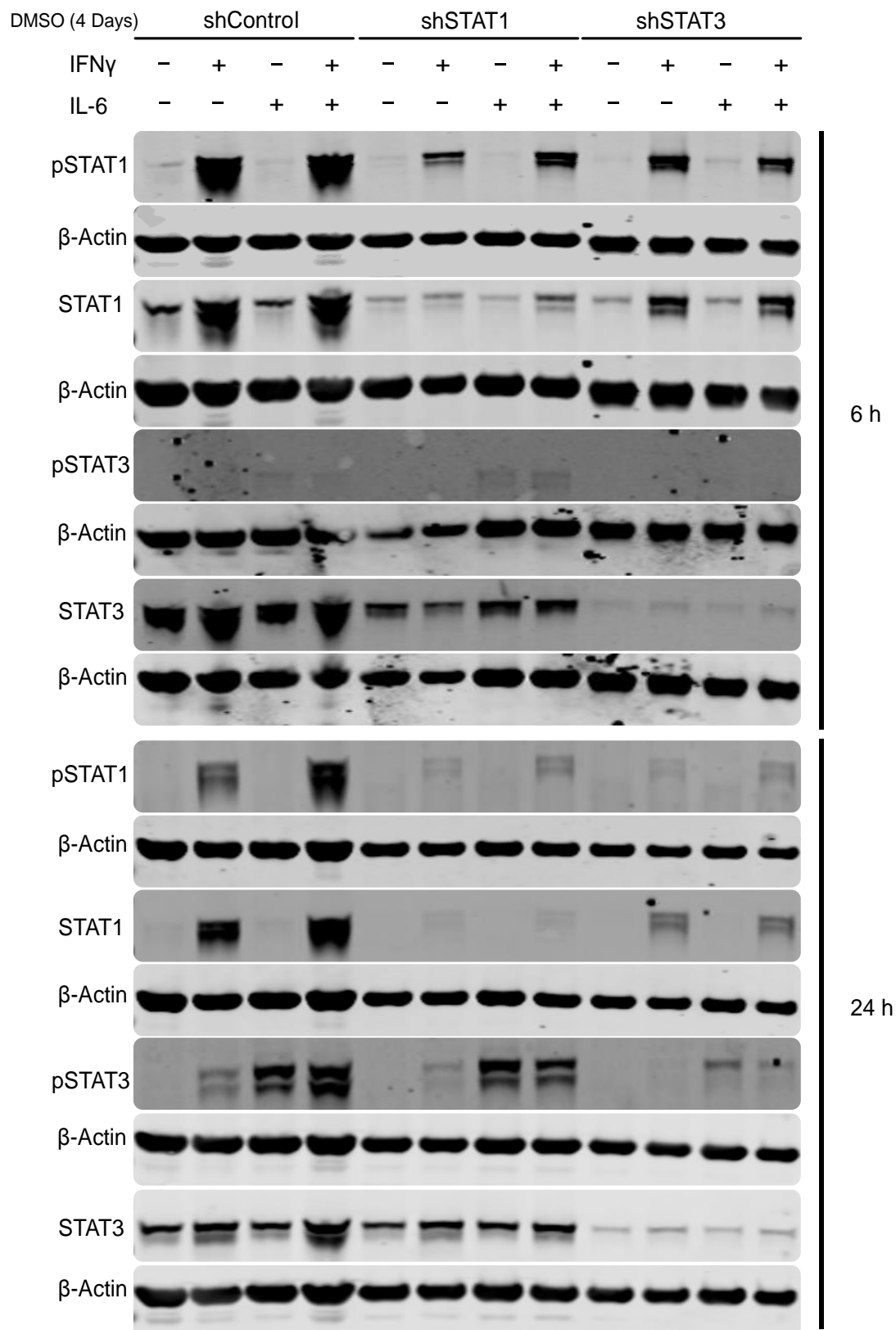


Figure 49: STAT1 and STAT3 expression and phosphorylation in control, shSTAT1, and shSTAT3 HL-60 cells differentiated with DMSO. Stimulation was performed with IFN- γ , IL-6 or combination of both for 6h, 24h at analyzed at day 4

3.16 Downregulation of ezrin in STAT1- and STAT3-knockdown HL-60 cells

Ezrin, a member of the ezrin/radixin/moesin (ERM) family of proteins, cross-links the cytoskeleton and the cell membrane mediating various cellular processes. Here, to investigate the expression of ezrin, we blotted for ezrin in control and STAT1 and STAT3-knockdown HL-60 cells that are differentiated with DMSO (day 4). In this preliminary data we found that the knockdown of STAT1 and STAT3 by shRNA decreased the expression of ezrin in HL-60 cells when left untreated or treated with IFN- γ and IL-6 or a combination of both for 24 hours compared to control shRNA cells (Figure 50).

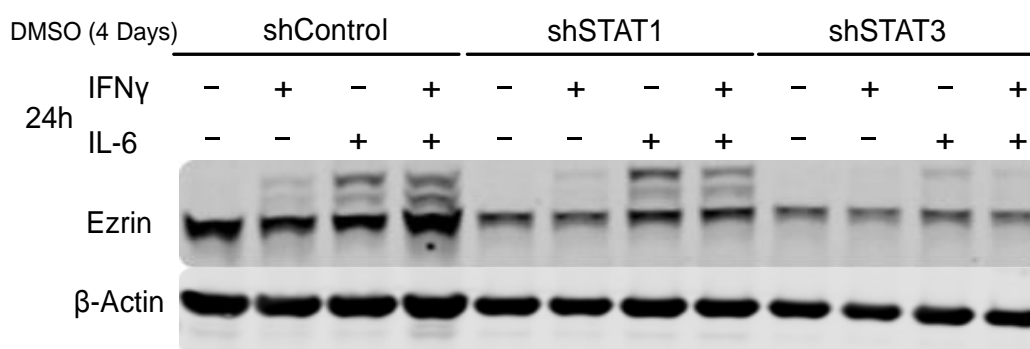


Figure 50: Ezrin expression levels in control, STAT1- and STAT3-knockdown HL-60 cells differentiated with 1.3% DMSO. Cells were analyzed at day 4 and left unstimulated or stimulated with IFN- γ , IL-6 or combination of both for 24 hours.

3.17 Reduced transendothelial migration in HL-60 cells with decreased STAT1 and STAT3 expression

Parallel plate flow chamber assays were used for live-cell imaging to investigate the role of STAT1 and STAT3 on the recruitment and transmigration of neutrophil-like HL-60 cells to activated endothelial monolayers under constant flow conditions. While tracking of transmigrating cells have shown a reduced transendothelial migration in both STAT1 and STAT3-knockdown HL-60 cells, STAT3-knockdown resulted in a more pronounced inhibition of transmigration across the HUVEC endothelial monolayer as compared to control cells (n=3) (Figure 51). Interestingly, STAT3 knockdown has further impacted cell polarity and directional persistence in HL-60 cells.

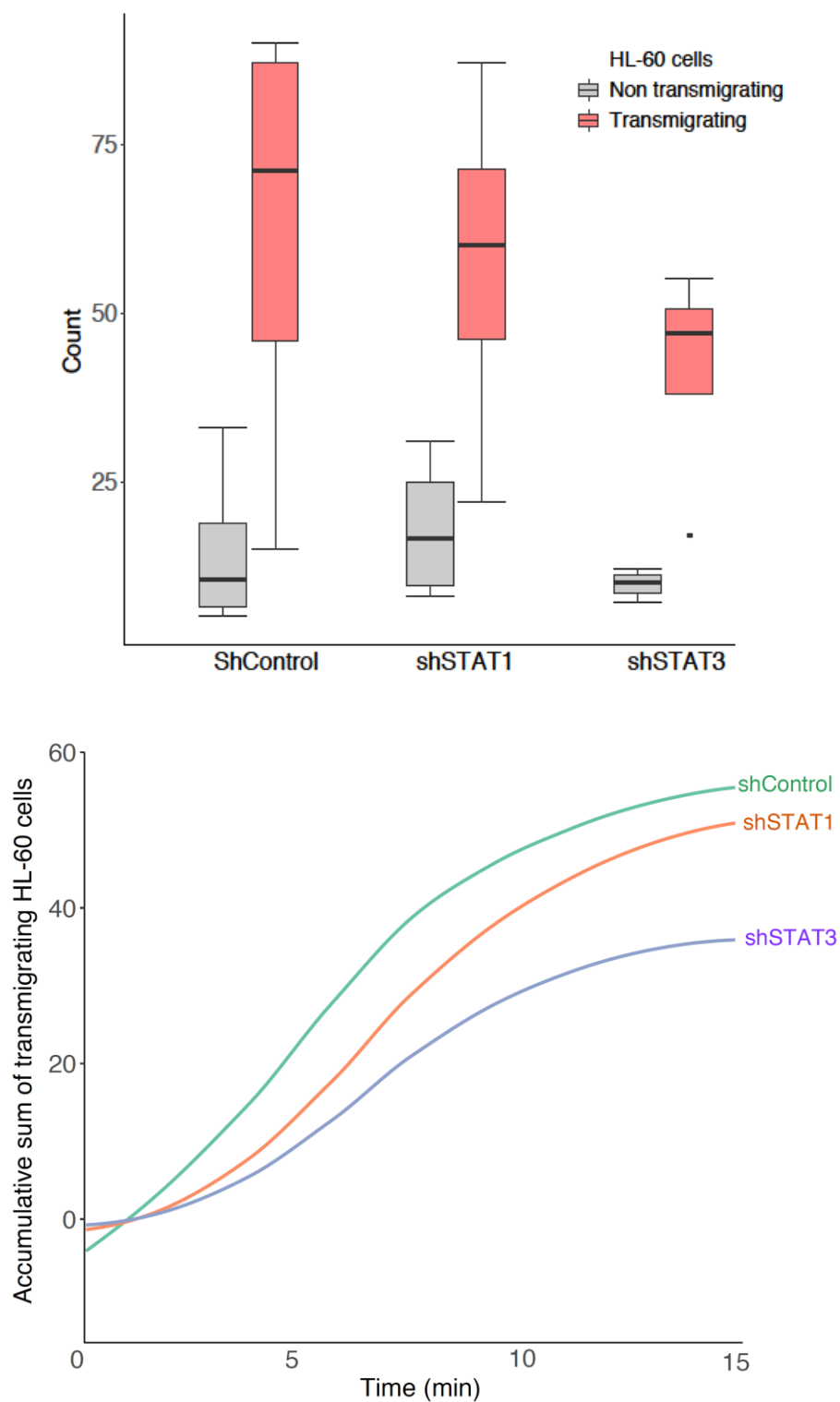


Figure 51: Extravasation analysis of HL-60 cells across HUVEC monolayer in STAT1 and STAT3 shRNA-transfected cells compared to cells transfected with control shRNA. All cells were differentiated with DMSO and analyzed at day 4.

4. Discussion

The Janus kinase-signal transducer and activator of transcription (JAK-STAT) signalling pathway has been identified as a critical regulator of several physiological responses following cardiac insult including oxidative damage, myocarditis, and cardiac hypertrophy. Moreover, elevated levels of circulating cytokines have been recognized to activate the JAK-STAT signal transduction and modulate the functional status of heart patients (Rumalla *et al.*, 2002; Jug *et al.*, 2009; Parissis *et al.*, 2004; Ruppert and Meyer, 2007; Simon *et al.*, 1998; Ueland *et al.*, 2015; Wagner and Siddiqui, 2012).

Interferon signalling is initiated by binding of the cytokine to the extracellular domain of its cognate receptor. To facilitate differential regulation of type I and type II interferon signalling, interferon- α receptor recruits STAT1 as well as STAT2, while interferon- γ receptor exclusively recruits STAT1. The STAT1 homodimer is formed by interactions between the SH2 domains of two phosphorylated STAT1 protomers. The STAT1-F77A point mutant, with alanine substituted for phenylalanine at position 77, is defective in forming stabilized tetramers on DNA and shows no cooperative DNA binding (Begitt *et al.*, 2014).

In this project, the role of STAT1-cooperative DNA binding, its transcriptional consequences in the infarcted myocardium, and its impact on cardiac function have been investigated in a mouse model of tetramerisation-deficient STAT1 signalling. To this end, permanent occlusion of the left anterior descending coronary artery was performed to experimentally induce infarction in the left ventricle in wild-type and knock-in STAT1-F77A mice, whereas sham-operated mice were used as controls to determine baseline characteristics.

To assess cardiac structure and function following sham operation and LAD ligation, we analyzed cardiac chamber dimensions, fractional shortening, intracardial volumes, the ejection fraction, and wall thickness using echocardiographic ultrasound imaging with two dimensional views of the heart (short and long axis). Subsequently, RNA-seq technology was used to delineate the role of STAT1-cooperative DNA binding by analyzing RNA samples from the infarcted tissue in wild-type and knock-in mice in comparison to sham-operated left ventricular myocardium (Wang *et al.*, 2009).

Assessment of echocardiographic parameters in wild-type and STAT1-F77A mice demonstrated a crucial role of STAT1 signalling, where IFN- γ -irresponsive female mice lacking cooperative DNA binding were protected from adverse cardiac remodeling (Figure 4). This finding is consistent with previous observations, in which mice deficient in interferon regulatory factor 3 (IRF3) or type I IFN receptor (IFNAR) have an improved survival rate

following MI as compared to control mice (King *et al.*, 2017). In the same study, single-cell RNA-seq analysis of 4215 isolated leukocytes from infarcted and healthy heart tissues revealed an activation of the IRF3-interferon axis in cardiac macrophages which had driven a fatal response.

Moreover, treatment of mice with an IFNAR-neutralizing antibody post-MI abrogated the interferon response and improved left ventricular size, contractile function, and survival (King *et al.*, 2017). Similar results were also reported in another study, in which STAT1-deficient mice had a significantly smaller infarct size compared to wild-type mice. The authors observed an upregulation of autophagy in STAT1-deficient hearts, characterized by light chain 3 (LC3)-I/LC3-II conversion and an increased expression of cellular autophagy markers Atg12 and Beclin 1, suggesting that the absence of functional STAT1 signalling is protective within the context of myocardial infarction by enhancing autophagy (McCormick *et al.*, 2012).

It has been previously reported that STAT1 activates *Fas/FasL* genes and apoptosis in cardiac myocytes, when subjected to ischemia/reperfusion injury, which was mediated by the phosphorylation of serine residue 727, but not the tyrosine residue 701 of STAT1 (Stephanou *et al.*, 2001). Further *in vitro* experiments have shown that STAT1 interacts directly with p53 to regulate the expression of pro-apoptotic target genes, such as *Bax*, *Noxa*, and *Fas*. These data provide further support for the role of STAT1 in promoting DNA damage-induced apoptosis (Townsend *et al.*, 2004). Similarly, a previous study by Stephanou and co-workers demonstrated a higher susceptibility to ischemia-induced cell death in STAT1-transfected cells compared to cells transfected with control plasmid. The authors have also reported an activation of STAT1 and caspase-1 processing in myocytes following ischemia/reperfusion injury using an *ex vivo* model of intact heart (Stephanou *et al.*, 2000). Over the last decade, the literature has further accumulated enough data to convincingly establish a role for the JAK/STAT signalling pathway in ischemia-reperfusion injury and ischemic post-conditioning (Barry *et al.*, 2007; Boengler *et al.*, 2010; Dawn *et al.*, 2004; Goodman *et al.*, 2008; Goodman *et al.*, 2011; Gross *et al.*, 2006; Heusch *et al.*, 2012; Huffman *et al.*, 2008; Kodama *et al.*, 1998; Smith *et al.*, 2010; Xuan *et al.*, 2005). Of particular importance are the experiments by several groups concerning the cardioprotective effects of STAT3 during ischemia/reperfusion injury as well as under pressure overload (Bolli *et al.*, 2011; Butler *et al.*, 2006; Fuglestad *et al.*, 2008; Heusch *et al.*, 2011; Hilfiker-Kleiner *et al.*, 2004; Jacoby *et al.*, 2003; Kunisada *et al.*, 2000; Oshima *et al.*, 2005; Pedretti and Raddatz, 2011; Smith *et al.*, 2004; Zhuo *et al.*, 2011).

Numerous studies in this fast-moving field have now convincingly shown that abrupt reduction in blood flow through the coronary artery can expose the cells to hypoxia, resulting in an irreversible cell damage and necrosis (Semenza *et al.*, 2014). Necrosis, characterized by cytoplasmic granulation and cellular swelling, leads to cell membrane rupture and breakdown of organelles, which consequently culminates in the activation of a severe inflammatory response (Kung *et al.*, 2011). However, the mechanisms of how STAT1 cooperative DNA binding plays a role in mediating this MI-associated inflammatory response have remained unexplored.

In this study, next-generation sequencing (RNA-seq) technology was used to analyze the transcriptome of infarcted tissue in STAT1-F77A mice as compared to wild-type littermates. This approach has gained high-resolution data without any previous knowledge about the sequences under exploration. Within the ongoing efforts to construct a comprehensive model which can predict phenotypes from the transcriptomic data, KEGG (Kyoto Encyclopedia of Genes and Genomes) tries to answer different biological questions by integrating the recent understanding of molecular functions and processes at the level of genes, proteins, and pathways (Kanehisa and Goto, 2000).

Interestingly, our gene set enrichment analysis has revealed PI3K-Akt and JAK-STAT signalling pathways as the top enriched signal transduction pathways following myocardial infarction. A previous study has demonstrated that the activation of PI3K (p110 α) in mice protects against myocardial infarction-induced heart failure (Lin *et al.*, 2010). In our data set, transcriptomic analysis of cardiac tissue at day 1 following myocardial infarction revealed prominent gene expression changes in wild-type mice and STAT1 tetramerisation-deficient mice. We observed a significantly higher number of differentially expressed genes in STAT1-F77A mice, since 2337 genes were differentially regulated with a $|\log_2(\text{fold change})| > 1$ and a $p\text{-value} < 0.05$, including 909 upregulated genes and 1428 downregulated genes compared to wild-type mice (Figure 8).

Utilizing KEGG database, we have identified common and unique differentially expressed pathways in both wild-type and STAT1-F77A animals following LAD ligation. These were enriched in four categories including immune processes, signal transduction, cellular processes, and metabolism (Figure 13 and Figure 14). The top enriched immune processes that are upregulated post-MI in both wild-type and knock-in mice are cytokine-cytokine receptor interactions, chemokine signalling pathways, cell adhesion molecules, regulation of actin cytoskeleton, and leukocyte transendothelial migration.

These data are in line with prior studies where gene expression data sets of MI patients were analyzed. For instance, Kontou et al. has identified using blood samples of 93 MI patients and 89 healthy controls differentially expressed genes involved in inflammation, wound healing, nuclear pore proteins, and RNA processing and transport (Kontou *et al.*, 2018). In patients with ST-elevation myocardial infarction (STEMI), Muller and colleagues have used custom-designed DNA array to analyze blood samples at the site of occlusion and in the peripheral circulation during primary percutaneous coronary intervention. Validation of identified genes by qPCR has demonstrated an upregulation of chemokines (*Ccl2*, *Ccl18* and *Cxcl12*), cell-extracellular matrix (*Fnl*, *Cdh5* and *Spp1*), and lipoprotein modification genes (*Apoe*) in local versus peripheral blood (Muller *et al.*, 2011).

Moreover, current research has shown that low serum levels of CCL2 chemokine (≤ 100.9 pg/mL) in 803 patients with acute coronary syndrome were significantly associated with fatal and non-fatal myocardial infarction (Leocádio *et al.*, 2019). Using microarray and protein networks to analyze blood samples from 127 acute MI patients, Devaux and co-workers have demonstrated the involvement of vascular endothelial growth factor B (*Vegf-b*), thrombospondin-1 (*Thbs1*) and placental growth factor (*Pgf*) as potential biomarkers (Devaux *et al.*, 2010). In addition, using RNA-seq of MI patients' blood samples, Zhao and colleagues have identified several immune-related genes, including interferon-induced protein with tetratricopeptide repeats 3 (*Ifit3*), interferon-induced GTP-binding protein (*Mx1*), and major histocompatibility complex class II DQ α 1 (*Hla-dqa1*) as potentially relevant for myocardial infarction (Zhao *et al.*, 2018).

In a recently published study in mice, several pro-inflammatory transcripts have been detected during the first week post-myocardial infarction, including tumor necrosis factor- α (*Tnf- α*), monocyte chemoattractant protein-1 (*Mcp-1*), interleukin-6 (*Il-6*), and interleukin-1 β (*Il-1 β*). Administration of an TNF- α inhibitor has been reported to adversely affect cardiac function, suggesting that inflammatory responses play a protective role in the early stage of myocardial infarction (Wang *et al.*, 2018). Furthermore, recent research has shown in mice that depleting neutrophils from the infarcted myocardium at day 1, using a monoclonal anti-Gr-1 antibody, had resulted in a downregulation of IFN- γ and TNF- α mRNA transcripts, higher mortality rates and worsened cardiac function compared to infarcted controls (Finger *et al.*, 2019). Collectively, these observations indicated that IFN- γ mediates a sequential chemotactic cellular immune response that regulates survival and cardiac function.

On the other hand, administration of IL-10 has been shown to improve left ventricular function in rats following MI, which was associated with a significant downregulation of pro-

inflammatory cytokines and chemokine levels including TNF- α , IL-6, and MCP-1. This effect was also accompanied by decreased infiltration of macrophages in the myocardium (Jung *et al.*, 2017; Stumpf *et al.*, 2008). Experimental evidence indicates that transcriptional repression of monocyte interleukin-10 (IL-10) is mediated through STAT1 signalling (VanDeusen *et al.*, 2006). Thus, to inhibit the excessive recruitment of monocyte to the infarcted myocardium, Wang and co-workers have developed poly(ethylene glycol) (PEG)-distearoylphosphatidylethanolamine (PEG-DSPE) micelles loaded with a CCR2 (C-C chemokine receptor type 2) antagonist. These micelles were further surface-decorated with an anti-CCR2 antibody for efficient targeting. Both non-targeted and CCR2-targeting micelles exhibited a tendency to improve cardiac function (Wang *et al.*, 2018). These experiments highlight the crucial effects immune cells play in cardiac remodeling post-MI.

Our results from DESeq2 analysis revealed a set of immune-activating cell markers that were differentially expressed at a higher fold change in the early phase post-MI in STAT1-F77A mice as compared to their wild-type littermates (Figure 42). In particular, the expression profile in the myocardium of STAT1-F77A mice following LAD ligation had identified candidate genes involved in chemokines and their receptors, adhesion molecules, cytokines and their receptors, growth factors and their receptors, coagulation cascade proteins, and heat shock proteins. The top ten differentially upregulated genes in the infarcted myocardium of STAT1-F77A mice were *Fgf23*, *Cxcl3*, *Slfn4*, *Rab44*, *Cxcl2*, *Cd177*, *Gm5483*, *Il1r2*, *Cd300lf*, and *Slfn1*.

Moreover, it is noteworthy to highlight that STAT1-F77A mice had a downregulation in the contractile machinery components compared to wild-type mice (Figure 41). This finding might be a result of a higher ratio of infiltrating immune cells to cardiac cells in the infarcted area, which is essential for cardiac repair processes. This is very interesting in light of accumulating research pinpointing to the inflammatory response as a key factor in ventricular remodeling, where it is associated with chemokine signalling as well as recruitment and migration of different immune cell subsets (Azevedo *et al.*, 2016; Frangogiannis, 2014; Liu *et al.*, 2015; Prabhu and Frangogiannis, 2016; Xia and Frangogiannis, 2017).

To further explore the molecular drivers of myocardial infarction that are under the control of STAT1, we performed a gene set enrichment analysis using the results of the DESeq2 differential expression analysis. Our study in the transgenic mouse line adds to previous findings by identifying top downregulated metabolic pathways post-MI, which are oxidative phosphorylation, citrate cycle, fatty acid metabolism, fatty acid degradation, and propanoate metabolism (Nojiri *et al.*, 2006; Stanton *et al.*, 2000; van Bilsen *et al.*, 2004; Xu *et al.*, 2011).

In the present study, we demonstrated a higher decrease in the expression of a multitude of genes involved in mitochondrial oxidative phosphorylation, citrate cycle, and substrate utilization in the infarcted myocardium in STAT1-F77A mice. These findings are interesting given that STAT1 has been identified as a transcriptional modulator of Warburg effect in the context of tumorigenesis, regulating genes involved in glycolysis, the citrate cycle and mitochondrial oxidative phosphorylation (Pitroda *et al.*, 2009).

It is important to point out that the highest energy yield in the normal heart comes from metabolizing and consuming non-carbohydrate substrates, such as fatty acids. However, under certain pathological conditions, glucose becomes a preferential substrate in producing energy per oxygen consumption. These alterations are driven by an intricate transcriptional regulatory network that feeds into factors including substrate availability and workload (Nagoshi *et al.*, 2011). Modifications in the oxidative phosphorylation machinery as a result of inflammation-induced NF- κ B activation culminate in changes in energy requirement to support systolic and diastolic function (Gordon *et al.*, 2011; Remels *et al.*, 2018). Indeed, it has been suggested that shifting substrate utilization from fatty acids to glucose can improve cardiac function and slow the progression of heart failure (Lionetti *et al.*, 2011).

It has recently been shown that macrophages exhibit distinctive transcriptomic profiles during the first week of myocardial infarction. At day 3 post-MI, macrophages displayed an upregulation of genes involved in proliferation, phagocytosis, mitochondrial function and oxidative phosphorylation, further suggesting an important role for metabolic reprogramming in macrophage polarization (Mouton *et al.*, 2018).

In order to evaluate post-infarction remodeling in a swine model of MI, Yang and co-workers have utilized label-free quantitative proteomics and immunohistochemical analyses (Yang *et al.*, 2017). Subsequent experiments in their model have shown a significant downregulation of proteins involved in energy metabolism, suggesting dysfunctional myocardial energetics and mitochondrial impairment in the peri-infarct border zone. Numerous other mitochondrial processes were also similarly downregulated, including branched-chain amino acid catabolism, β -oxidation, fatty acid metabolism, Krebs cycle, and the formation of iron-sulfur clusters (Yang *et al.*, 2017). These results are in line with our observation that MI interferes at a global transcriptional level with oxidative phosphorylation. Taken together, these findings clearly support that the imbalance of cardiac substrate metabolism plays a significant role in the pathogenesis of heart failure (Birkenfeld *et al.*, 2019; Noordali *et al.*, 2018).

We have postulated that STAT1-F77A mice will demonstrate a similar phenotype as mice expressing hyperactive STAT3 with respect to cardioprotective actions. Phillips *et al.*

suggested that STAT3 is conceivably regulating mitochondrial machinery through transcriptional regulation or indirect signalling pathways (Phillips *et al.*, 2010). However, according to current concepts activated STAT3 enhances mitochondrial oxidative phosphorylation and promotes transformation by oncogenic Ras. The observed downregulation of oxidative phosphorylation in STAT1-F77A mice may possibly be attributed to the dependence of infiltrating immune cells on glycolysis as a major pathway of energy production which is essential for the restoration of homeostasis (Buck *et al.*, 2017).

Our RNA-seq analysis has several limitations as the data indicate the average gene expression pattern using millions of cells which can mask biologically relevant signatures between different cell types. Therefore, additional studies using single-cell RNA-seq (scRNA-seq) technology will be required to overcome this limitation and determine the transcriptional program in individual cell populations including cardiomyocytes, fibroblasts, endothelial cells, and immune cells with unparalleled resolution (Farbehi *et al.*, 2019).

Although there are plenty of studies investigating the role of the immune system in myocardial infarction (Lai *et al.*, 2019), many aspects of immunological modulation by STAT1 and STAT3 in the infarcted heart still require further elucidation. Therefore, the overarching aim of the next experiments were to study the balance of STAT1 and STAT3 activation in murine heart endothelial cells. Several studies have reported STAT1 and STAT3 playing opposing roles in regulating multiple cellular processes including survival, proliferation, motility and inflammation (Avalle *et al.*, 2012). In our data, we observed that upon co-stimulation with IFN- γ and IL-6 in MHEC-5 cells, the RNA expression level of STAT1 target genes were reduced as compared to cells stimulated with IFN- γ only, however, the phosphorylation levels of STAT1 and STAT3 were similar in both conditions.

The results presented here open up new possibilities for analyzing the role of STAT1:STAT3 heterodimers in modifying signals downstream of the receptor by integrating different signal inputs. Sadreev and colleagues recognized the importance of STAT heterodimer species in regulating immune responses (Sadreev *et al.*, 2018). The authors postulated that the rearrangement of various STAT dimers formed by stimulation with different extracellular cytokines can regulate T-cell plasticity. They further suggested that the competing STAT complexes can indirectly inhibit each other depending on extracellular cytokine concentrations. An interesting aspect of these findings is that they highlight the biological role of STAT heterodimeric complexes which have remained enigmatic (Delgoffe and Vignali, 2013; Sadreev *et al.*, 2018).

The focus of my project was then directed towards studying the role of STAT1 and STAT3 proteins in controlling transendothelial migration and the regulation of the cytoskeletal ERM component ezrin in the neutrophil-like HL-60 cell line. In light of their importance in ischemic heart disease, neutrophils have been extensively studied, with excessive infiltration contributing to adverse cardiac remodeling in the injured myocardium. Accordingly, directing detrimental neutrophils away from the site of cardiac injury is an interesting avenue to investigate (de Oliveira *et al.*, 2016).

In a parallel plate flow chamber assay, analysis of STAT1 and STAT3 knockdown HL-60 cells showed a reduced transendothelial migration across an endothelial monolayer compared to control cells. Therefore, STAT1 and STAT3 knockdown highlighted defects in neutrophils' transendothelial migration, as modelled in HL-60 cells. Our original observation indicated that ezrin expression levels are lower in STAT1 and STAT3 knockdown HL-60 cells as compared to control cells following differentiation with DMSO at day 4. The recently published paper by Ivetic *et al.* provided an interesting model for monocyte behavior during transendothelial migration. Herein, the authors described a novel physiological function of sequential binding of ezrin and moesin to L-selectin, which modulates L-selectin shedding and hence protrusion dynamics and polarity. In our data, it is interesting to note that STAT3 knockdown in HL-60 cells may result in disorganized cell polarity and loose directional persistence. These novel data are in agreement with earlier reports, in which STAT3 knockdown in mouse embryonic fibroblasts resulted in increased Rac1 activity, which regulates directional persistence and generation of actin stress fibers, thus inducing a random mode of migration (Teng *et al.*, 2009). The authors have also established a direct binding of STAT3 to β -PIX, which is a known Rac1 activator, suggesting a possible mechanism for STAT3 in modulating actin cytoskeleton reorganization and inducing directional movement (Teng *et al.*, 2009). Other studies have focused on the role of IL-6 in promoting epithelial-to-mesenchymal transition and metastatic behavior via STAT3 signalling pathway within the context of carcinogenesis (Yadav *et al.*, 2011). Nevertheless, the physiological significance of these data is yet to be evaluated within the context of myocardial infarction. These studies would pave the way for more sophisticated experiments involving the re-introduction of shRNA immune constructs of mutant STAT1-F77A. Alternatively, the more recently developed CRISPR/Cas9 platform of gene editing could be used to engineer the F77A mutant.

With recent advancements in STAT1 signalling, current concepts are continually evolving to describe the molecular and cellular processes in the failing heart. In concert with prior studies,

the observed upregulation of immune cell markers and adhesion molecules and the significant decrease in the expression of oxidative phosphorylation genes in STAT1-F77A as compared to wild-type mice may be beneficial in the early phase to meet the demands of repair processes through metabolic rewiring and immune cells recruitment. Nowadays, a compelling body of experimental evidence indicates that innate and adaptive immune cells play a fundamental role in cardiac remodeling (Aubert *et al.*, 2013). Certainly, our finding that immunity-related pathways are significantly upregulated following myocardial infarction is in good agreement with previous studies showing increased expression of genes involved in inflammation, chemotaxis, blood agglutination, and immunity in the myocardial tissue of MI-operated mice (Li *et al.*, 2019). Furthermore, we observed a downregulation of a multitude of genes that are involved in ATP production when STAT1 tetramer activity is suppressed. The heart fails when its generation or utilization of free energy in the process of contraction is inadequate for the cardiac load imposed. These observations may help explain the why STAT1-F77A mice are protected from adverse cardiac remodeling. Overall, these results provide evidence for a plausible link between interferon signalling and the modulation of gene signature related to mitochondrial ATP synthesis. Our findings will encourage further investigations to study if these metabolic changes observed at the transcriptional level of the entire genome are reflected by parallel changes in the enzymatic activity of the proteins as well as the levels of metabolites. Therapeutic interventions which modify substrate utilization will be a promising target to optimize cardiac metabolism and function. Hence, investigating how does STAT signalling affect the utilization of alternative fuel sources in cardiac cells in addition to the major substrates utilized will be of major importance for an understanding of cardiac metabolic reprogramming.

5. References

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Biography

Asmma Doudin was born in 1991 in Amman, Jordan. She received her secondary education in Fatima Al-Zahra Primary and Secondary School in Sharjah. She graduated and received her bachelor's degree in Applied Biology/Biotechnology (2012) from the College of Science and her Master's in Molecular Medicine (2014) from the College of Medicine at the University of Sharjah. Following her graduation Asmma worked as a research assistant at Sharjah Institute of Medical Research (2015). In the year 2016, she joined the International Research Training Group 1816 which is funded by the German Research Foundation. The experimental work for her Ph.D. thesis was conducted in the Molecular Psychocardiology Laboratory at the Department of Psychosomatic Medicine at Georg-August University of Göttingen, Germany, and at the Cardiovascular Department at King's College London, UK. Asmma was involved in research projects studying the molecular actions of STAT transcription factors in the pathophysiology of sterile inflammatory diseases, including murine myocardial infarction and experimental autoimmune encephalitis. She is also a young researcher in the German Centre for Cardiovascular Research (DZHK) and has published papers on the relationship between serum 25-hydroxyvitamin D and red blood cell indices in German adolescents.